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MITOGENOME ANNOUNCEMENT

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The complete mitochondrial genomes of *Ceratitis rosa* and *Ceratitis quilicii*, members of the *Ceratitis* FAR species complex (Diptera: Tephritidae)

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ABSTRACT

Ceratitis FAR is an African species complex comprising insect pests of great economic interest and obscure species limits. Here, we report the mitochondrial genomes of two members of the FAR complex, namely *Ceratitis rosa* and the recently characterized *Ceratitis quilicii*. A phylogenetic analysis based on PCGs of available Tephritidae mitogenomes is presented. The current mitochondrial sequences from the FAR complex could contribute toward the resolution of phylogenetic relationships and species limits within this taxonomically challenging group, which is also an important issue for the development of environment-friendly and species-specific control methods, such as the sterile insect technique (SIT).

Ceratitis rosa (Karsch, 1887), the Natal fruit fly, and Ceratitis quilicii (De Meyer, Mwatawala & Virgilio sp. Nov) are African insect pests and members of the Ceratitis FAR species complex (Diptera: Tephritidae) which also includes Ceratitis fasciventris and Ceratitis anonae. Ceratitis quilicii has been recently described as a different species from C. rosa based on molecular genetics, morphometrics, developmental physiology, behavior and sexual compatibility, chemo ecology, and environmental preferences data (reviewed in De Meyer et al. 2016). The two species are almost identical (only adult males exhibit minor differences in the mid tibia) and have overlapping distribution in eastern and southern Africa (De Meyer 2001; De Meyer et al. 2016; Copeland et al. 2006). They are both highly destructive as they attack a great number of wild and cultivated plants causing great economic consequences (De Meyer 2001; De Meyer et al. 2002, 2016; Copeland et al. 2006). Furthermore, there is concern about their invasive potential, since C. rosa has been reported in the islands of Mauritius and La Réunion (White et al. 2001). Later it was found that the La Reunion populations of C. rosa belong to the group R2 which was subsequently described as C. quilicii (Virgilio et al. 2013; De Meyer et al. 2016). In this study we present the first complete mitochondrial genomes of C. rosa and C. quilicii, which could be useful for further analyses aiming to ascertain the species number and clarify the evolutionary relationships within the FAR complex.

The analyzed specimens came from C. rosa and C. quilicii laboratory colonies held at the Insect Pest Control Laboratory (IPCL, Seibersdorf, Vienna). The original C. rosa colony was established at the International Center of Insect Physiology and Ecology (ICIPE, Kenya) from insects from guava fruits collected from Kibarani, Msambweni district (S 04°19'62.8"; E 039°32′41.1″; 34 masl). The original *C. quilicii* colony was established at the Citrus Research International (CRI, South Africa) from samples of infested Jambos and loquat collected from Pretoria (S 25°45'13.7"; E 28°13'45"). Both colonies have been verified by experts and used in previous studies (Tanga et al. 2015, 2018). Total genomic DNA was extracted from the whole body of individual adult flies using the DNeasy Blood and Tissue kit (Qiagen). DNA from specimens of the above colonies are kept in IPCL. Library construction and sequencing were performed by Macrogen using pair-ended $(2 \times 250 \text{bp})$ HiSeq 2500 technology.

Mitogenomes were assembled from the quality-trimmed sequencing reads using the mitochondrial baiting and iterative mapping algorithm MITObim (Hahn et al. 2013) with default parameters. The mitogenome of *C. fasciventris* (NC_035497) was used as the initial template. Annotation was performed using the MITOS WebServer (Bernt et al., 2013) followed by manual curation using BLASTn (Altschul et al. 1997) and Clustal Omega (Sievers et al. 2011) alignments to the mitogenomes of *C. capitata* (NC_00857) and *C. fasciventris* (NC_035497). For the confirmation of tRNA

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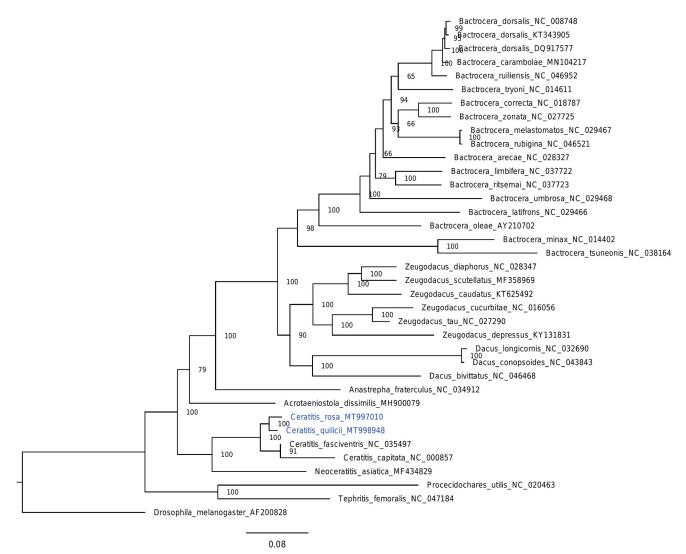


Figure 1. Phylogenetic tree based on the 13 PCGs of Tephritidae mitogenomes. Phylogeny was inferred in W-IQ-TREE. Numbers at nodes are for bootstrap percentages from 1000 replicates. Numbers following species names are GenBank accession numbers. The sequences analyzed in the present study are presented in blue.

annotations, the tRNAscan-SE (Lowe and Eddy 1997) was used. Maximum-likelihood analysis based on the 13 PCGs from Tephritidae species was inferred in W-IQ-TREE (Trifinopoulos et al. 2016) using the General Time Reversible model with empirical base frequencies and invariable site plus discrete Gamma mode (GTR + F + I + G4) and ultrafast bootstrap analysis (1000 replicates).

The length of the *C. rosa* and *C. quilicii* mitogenomes was 16,047 and 16,035 bp and the A + T content 77.4 and 77.5%, respectively. Each mitogenome contains 13 PCGs (*ND1-6, ND4L, COI-III, ATP8, ATP6,* and *CYTB*) two rRNA (*12S* and *16S rRNA*) and 22 tRNA genes, and one major non-coding sequence, the control region (CR), following the typical organization of insect mitogenome (Cameron 2014). Six PCGs start with ATG, five with ATT, one with ATA and one with TCG, while ten terminate with TAA, one with TAG, and two with an incomplete stop-codon (T). The length of the CR is 1028 bp for *C. rosa* and 1036 bp for *C. quilicii*. The longest intergenic spacer lies between *tRNA^{IIe}* and *tRNA^{GIn}* for both species, similarly to *C. fasciventris* (Drosopoulou et al. 2017). The two analyzed mitogenomes present 97.75% sequence identity to each other and a very high identity to the

mitogenome of *C. fasciventris* (97.85% for *C. rosa* and 98.32% for *C. quilicii*). The phylogenetic analysis among available Tephritidae mitogenomes clusters the currently analyzed species in a well-supported clade (Figure 1), confirming their close phylogenetic relationships. The availability of additional FAR mitogenome sequences in the future is expected to provide useful information for the better understanding of species identities within the complex, which is necessary for the development and application of environment-friendly control methods, such as the SIT.

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Disclosure statement

No potential conflict of interest was reported by the author(s).

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Data availability statement

The genome sequence data that support the findings of this study are openly available in GenBank of NCBI at https://www.ncbi.nlm.nih.gov/ under the accession no. MT997010 (*C. rosa*) and MT998948 (*C. quilicii*). The associated BioProject, SRA, and Bio-Sample numbers are PRJNA680883, SRR13181787 and SRR13171161, and SAMN16925622 and SAMN16925623, respectively.

References

- Altschul S. 1997. Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. Nucleic Acids Research. 25(17): 3389–3402. doi:10.1093/nar/25.17.3389.
- Bernt M, Donath A, Jühling F, Externbrink F, Florentz C, Fritzsch G, Pütz J, Middendorf M, Stadler P F. 2013. MITOS: Improved de novo metazoan mitochondrial genome annotation. Mol. Phylogenet. Evol. 69(2): 313–319. doi:10.1016/j.ympev.2012.08.023.
- Cameron S L. 2014. Insect Mitochondrial Genomics: Implications for Evolution and Phylogeny. Annu Rev Entomol. 59(1):95–117. doi:10. 1146/annurev-ento-011613-162007.
- Copeland R, Wharton RA, Luke Q, De Meyer M, Lux S, Zenz N, Machera P, Okumu M. 2006. Geographic distribution, host fruit, and parasitoids of African fruit fly pests *Ceratitis anonae*, *Ceratitis cosyra*, *Ceratitis fasciventris*, and *Ceratitis rosa* (Diptera: Tephritidae) in Kenya. Ann Entomol Soc Am. 99(2):261–278.
- De Meyer M. 2001. Distribution patterns and host-plant relationships within the genus *Ceratitis* MacLeay (Diptera: Tephritidae) in Africa. Cimbebasia. 17:219–228.
- De Meyer M, Copeland RS, Lux SA, Mansell M, Quilici S, Wharton R, White IM, Zenz NJ. 2002. Annotated check list of host plants for Afrotropical fruit flies (Diptera: Tephritidae) of the genus *Ceratitis*. Vol. 27. Brussels (Belgium): R. Museum Cent.
- De Meyer M, Mwatawala M, Copeland RS, Virgilio M. 2016. Description of new *Ceratitis* species (Diptera: Tephritidae) from Africa, or how

morphological and DNA data are complementary in discovering unknown species and matching sexes. Eur J Taxon. 233:1–23.

- Drosopoulou E, Pantelidou C, Gariou-Papalexiou A, Augustinos AA, Chartomatsidou T, Kyritsis GA, Bourtzis K, Mavragani-Tsipidou P, Zacharopoulou A. 2017. The chromosomes and the mitogenome of *Ceratitis fasciventris* (Diptera: Tephritidae): two genetic approaches towards the *Ceratitis* FAR species complex resolution. Sci Rep. 7(1): 4877.
- Hahn C, Bachmann L, Chevreux B. 2013. Reconstructing mitochondrial genomes directly from genomic next-generation sequencing reads A baiting and iterative mapping approach. Nucleic Acids Res. 41(13): e129.
- Lowe TM, Eddy SR. 1997. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. Nucleic Acids Res. 25(5): 955–964.
- Sievers F, Wilm A, Dineen D, Gibson TJ, Karplus K, Li W, Lopez R, McWilliam H, Remmert M, Söding J, et al. 2011. Fast, scalable generation of high-quality protein multiple sequence alignments using Clustal Omega. Mol Syst Biol. 7:539.
- Tanga CM, Khamis FM, Tonnang HEZ, Rwomushana I, Mosomtai G, Mohamed SA, Ekesi S. 2018. Risk assessment and spread of the potentially invasive *Ceratitis rosa* Karsch and *Ceratitis quilicii* De Meyer, Mwatawala & Virgilio sp. Nov. using life-cycle simulation models: implications for phytosanitary measures and management. PLoS One. 13(1):e0189138.
- Tanga CM, Manrakhan A, Daneel J-H, Mohamed SA, Khamis F, Ekesi S. 2015. Comparative analysis of development and survival of two Natal fruit fly *Ceratitis rosa* Karsch (Diptera, Tephritidae) populations from Kenya and South Africa. ZK. 540:467–487.
- Trifinopoulos J, Nguyen L-T, von Haeseler A, Minh BQ. 2016. W-IQ-TREE: a fast online phylogenetic tool for maximum likelihood analysis. Nucleic Acids Res. 44 (W1):W232–W235.
- Virgilio M, Delatte H, Quilici S, Backeljau T, De Meyer M. 2013. Cryptic diversity and gene flow among three African agricultural pests: *Ceratitis rosa, Ceratitis fasciventris* and *Ceratitis anonae* (Diptera, Tephritidae). Mol Ecol. 22(9):2526–2539.
- White IM, De Meyer M, Stonehouse J. 2001. A review of native and introduced fruit flies (Diptera, Tephritidae) in the Indian Ocean islands of Mauritius, Réunion, Rodrigues and Seychelles. Paper presented at the Indian Ocean Commission, Regional Fruit Fly Symposium;June 5–9; Flic en Flac, Mauritius.