

**GENETIC CHARACTERIZATION OF HONEYBEE (*Apis mellifera*) IN COMOROS ISLANDS USING MITOCHONDRIAL AND MICROSATELLITE MARKERS**

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**June, 2019**

**DECLARATION**

This thesis is my original work and has not been presented for a degree in any other University or other award.

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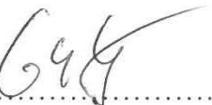
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**DEDICATION**

Dear Dad and Mum,

Remember the evenings you narrated folk tales? Yes! You broke them down in such a way that, young as I was, I always got the point. Those early lessons shaped my view of education as a concept of storytelling! Today, I dedicate my story to you. Broken down in a way that old as you may consider yourself, you will get the message. Thank you for having laid a strong foundation and motivating me to achieve the higher heights of education. It was not easy, but with your support, I moved on.

“*Mutio muno*” ~ Thank you!

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## TABLE OF CONTENTS

DECLARATION .....	Error! Bookmark not defined.
DEDICATION .....	ii
TABLE OF CONTENTS.....	v
LIST OF TABLES .....	viii
LIST OF FIGURES .....	ix
ABREVIATIONS .....	x
ABSTRACT.....	xi
CHAPTER ONE .....	1
INTRODUCTION .....	1
1.1 Background information .....	1
1.2 Statement of the problem .....	5
1.3 Justification .....	5
1.4 Hypothesis.....	7
1.5 Objectives.....	7
1.5.1 General objective .....	7
1.5.2 Specific objectives.....	7
CHAPTER TWO .....	8
LITERATURE REVIEW .....	8
2.1 Classification of honeybees.....	8
2.2 History and the origin of honeybees .....	9
2.3 Genetic diversity in honeybees and its importance.....	12
2.4 Characterization of honeybees based on mitochondrial DNA .....	14
2.5 Characterization of honeybees based on microsatellite DNA.....	15
2.7 Hardy Weinberg Equilibrium (HWE).....	19
2.8 Advantage of using $G_{ST}$ to determine population differentiation .....	20
CHAPTER THREE .....	22
MATERIALS AND METHODS.....	22
3.1 Study site .....	22
3.2 Sampling.....	22
3.3 DNA Extraction from honeybees.....	25

3.4 Mitotypes of honeybee colonizing the Comoros islands .....	25
3.4.1 PCR amplification of COI-COII intergenic region .....	25
3.4.2 Agarose gel electrophoresis and sample purification .....	26
3.4.3 Sequencing of the COI-COII intergenic region.....	27
3.4.4 COI-COII intergenic region sequence analyses .....	28
3.5 Genetic diversity, population differentiation and population structure.....	29
3.5.1 Microsatellite DNA amplification .....	29
3.5.2 Fragment analysis and allele scoring.....	31
3.5.3 Determination of queen mating frequency .....	32
3.5.4 Prediction of queens using drones .....	33
3.5.5 Genetic diversity and linkage disequilibrium.....	33
3.5.6 Population differentiation and population structure: .....	34
3.5.6.1 Population differentiation .....	34
3.5.6.2 Analysis of Molecular Variance (AMOVA).....	34
3.5.6.3 Population structure .....	34
CHAPTER FOUR.....	36
RESULTS .....	36
4.1 Mitotypes of honeybee colonizing the Comoros islands .....	36
4.1.1 Amplification of the COI-COII Intergenic Region .....	36
4.1.2 COI-COII intergenic region sequence analyses .....	37
4.2 Genetic diversity, population differentiation and population structure.....	43
4.2.1 Microsatellite DNA amplification .....	43
4.2.2 Fragment analysis and allele scoring.....	43
4.2.3 Determination of queen mating frequency .....	44
4.2.4 Prediction of queens using drones .....	44
4.2.4.1 Genetic diversity and linkage disequilibrium .....	45
4.2.4.2 Population differentiation and population structure .....	51
4.2.4.2.1 Population differentiation and Analysis of Molecular Variance .....	51
4.2.4.2.2 Population structure .....	51
CHAPTER FIVE .....	55
DISCUSSION, CONCLUSIONS AND RECOMMENDATIONS.....	55
5.1 Discussion .....	55
5.1.1 Mitochondrial DNA haplotypes in Comoros Islands .....	55

5.1.2 Genetic diversity, population differentiation and population structure .....	58
5.2 Conclusions .....	62
5.3 Recommendations .....	62
5.4 Future studies .....	62
REFERENCES .....	63
APPENDICES .....	72
APPENDIX I: Sampled colonies in the three main islands of Comoros .....	72
APPENDIX II: Primers used for microsatellite DNA analyses .....	75
APPENDIX III: ClustalO multiple sequence alignment.....	79
APPENDIX IV: GeneAlex file for predicted queen genotypes data .....	95
APPENDIX V: Table showing the deviations from HWE equilibrium.....	98
APPENDIX VI: AMOVA results .....	99

**LIST OF TABLES**

<b>Table 3.1:</b> Components of the 15 $\mu$ l reaction mix used for the amplification of mitochondrial DNA.....	26
<b>Table 3.2 :</b> List of microsatellite markers used for multiplex PCR reaction. ....	30
<b>Table 3.3:</b> The primer mix present in each reaction as co-loaded for fragment analysis.	31
<b>Table 4.1:</b> Pairwise genetic distances between groups. .....	41
<b>Table 4.2:</b> Genetic distances within groups. ....	41
<b>Table 4.3:</b> Table showing the observed and estimated drone genotypes. ....	44
<b>Table 4.4:</b> Table indicating various indices of diversity.....	46
<b>Table 4.5:</b> Pairwise Nei's G <sub>ST</sub> Statistics.....	50

## LIST OF FIGURES

<b>Figure 3.1:</b> Map showing sampling sites.....	24
<b>Figure 4.1:</b> Gel image for mitochondrial DNA COI-COII intergenicregion.....	36
<b>Figure 4.2:</b> Recombinant mtDNA (L2), <i>A. mellifera</i> and L1 haplotypes.....	37
<b>Figure 4.3:</b> Maximum likelihood phylogenetic tree for Comoros samples.....	39
<b>Figure 4.4:</b> Distribution of the three mtDNA haplotypes across the Comoros.....	41
<b>Figure 4.5:</b> Representative multiplex PCR gel image.....	42
<b>Figure 4.6:</b> Allele distribution at locus A113.....	45
<b>Figure 4.7:</b> Allele distribution at locus AP23.....	45
<b>Figure 4.8:</b> Heatmap showing deviations from the Hardy Weinberg Equilibrium.....	47
<b>Figure 4.9:</b> Heatmap showing pairwise linkage disequilibrium in honeybees collected in Anjouan.....	48
<b>Figure 4.10:</b> Heatmap showing pairwise linkage disequilibrium in honeybees collected in Ngazidja.....	49
<b>Figure 4.11:</b> Heatmap showing pairwise linkage disequilibrium in honeybees collected in Moheli.....	49
<b>Figure 4.12:</b> Discriminant Analysis Principal Component .....	51
<b>Figure 4.13:</b> A neighbor-joining tree structured population in Comoros islands .....	52
<b>Figure 4.14:</b> Population structure of honeybees in Comoros islands.....	53

**ABREVIATIONS**

AMOVA	Analysis of Molecular Variance
ANJ	Anjouan
CaCl	Calcium chloride
CCD	Colony Collapse Disorder
COI	Cytochrome Oxidase I
COII	Cytochrome Oxidase II
DCA	Drone Congregation Areas
DNA	Deoxyribonucleic acid
EtBr	Ethidium Bromide
ICIPE	International center of insect physiology and ecology
USDA	United States Department of Agriculture
U.V	Ultra Violet
KCl	Potassium Chloride
NaCl	Sodium Chloride
NCBI	National Centre for Biotechnology Information
NGA	Ngazidja
MLG	Multilocus genotype
MOH	Moheli
PCR	Polymerase Chain Reaction
SWIO	South-West of Indian Ocean

## ABSTRACT

Honeybees, *Apis mellifera*, are key contributors to the global economy, nutrition, food security and ecological biodiversity. Despite their robust benefits, declines in honeybee populations have been reported owing to a number of factors. Such include pests and diseases, climate change, poor management practices, change in land usage and low genetic diversity. High intra-colonial genetic diversity is associated with resilience and resistance to pest and diseases as well as increased colony level fitness. The use of morphological characters to characterize honeybees is limited by the fact that they are subject to manipulation by environmental selection pressures and introgression resulting from hybridization between sub-species. Hence, mitochondrial and microsatellite markers which are more powerful and neutral have been used. Honeybees native to the Comoros islands have not been characterized despite the fact that islands often possess endemic species which serve as distinct genetic reservoirs with unusual adaptations. This study aimed at generating extensive knowledge on the genetic diversity and population structure of honeybees in Comoros islands using mitochondrial and microsatellite markers. In the study, 160 wild and managed colonies were sampled across the Comoros archipelagos and characterized. Mitochondrial analyses involved PCR amplification and sequencing of the intergenic region spanning between COI- COII genes followed by phylogenetic analyses. In addition, 16 colonies distributed across the three islands were analyzed using 19 microsatellite markers. The mtDNA sequences analyses revealed the existence of three haplotypes in Comoros Islands where A1 haplotype, with a distribution of 51%, 80% and 91% in Anjouan, Moheli and Ngazidja respectively was predominant. Two other haplotypes (L1 and L2) were newly described in this study. They had a cumulative distribution of 48% in Anjouan, 20% in Moheli and 9% in Ngazidja. L1 and L2 showed high levels of variability as compared to A1 lineage. L2 arose from rare mtDNA recombination between L1 and A1 lineages. Discriminant Analysis of Principal Components, Provesti's neighbor joining tree and Bayesian clustering in STRUCTURE clustered the samples according to geography suggesting a restricted gene flow between islands. The population is strongly differentiated ( $G_{ST}=0.41(\pm 0.068)$ ) with a much stronger differentiation between Ngazidja and Moheli ( $G_{ST} = 0.53$ ) than Anjouan and Moheli ( $G_{ST} = 0.37$ ) or Anjouan and Ngazidja ( $G_{ST} = 0.38$ ). High levels of genetic diversity evidenced by high number of alleles per locus (5.92 ( $\pm 0.05$ )), and the high expected heterozygosity (0.66 ( $\pm 0.03$ )). Private alleles (216 and 218) were detected at A113 locus. In conclusion, mtDNA haplotypes can coexist and undergo rare mtDNA recombination as well as hybridization at the nuclear genome. There is need to investigate mechanisms that enable mtDNA recombination in honeybees.

## CHAPTER ONE

### INTRODUCTION

#### 1.1 Background information

Pollinators are major contributors to the world's economy, nutrition, food security as well as ecological biodiversity (Klein *et al.*, 2007). The global economic value of pollination services by both the wild and domesticated pollinators is estimated at USD 162 billion per annum (Gallai *et al.*, 2009). Vegetables, nuts and fruit producing crops, which are key sources of nutrients, are highly dependent on pollinators which makes them important in relation to micronutrient deficiencies in the developing world (Eilers *et al.*, 2011). Consumer surplus losses attributed to total loss of animal pollination services are estimated to range between USD 202 to 329 billion per annum (Gallai *et al.*, 2009). Apart from pollination services, the honeybee contributes to income generation through the sale of hive products such as honey, bee wax and royal jelly.

Insect pollination services are responsible for one-third of human meals. These pollination services are also vital to the ecosystem (Wallberg *et al.*, 2014). Honeybees contributes up to two thirds of the pollination services with estimates indicating that up to 35% of food consumed by human beings directly depends on insect-based pollination services (Koch, 2008). However, there has been an observed decline in the honeybees' populations over the past years (McMenamin and Genersch, 2015). In the United States (U.S) major winter losses average stands at 30% since 2006 while annual losses are were steadily approaching 50% by the year 2012 (CCD 2012). In the year 2006, United Kingdom

(U.K) experienced a 62% decrease in the honeybee population (Rac, 2006). According to a review by Pirk *et al.* (2016) on the status of honeybee health in Africa, Africa is yet to report major cases of honeybee colony losses.

The elevated honeybee losses in the U.S recorded from 2006 to 2008 was linked to a syndrome known as ‘Colony Collapse Disorder’ (CCD) (Johnson *et al.*, 2010; vanEngelsdorp *et al.*, 2009) (CCD Steering Committee, 2012a). Colony Collapse disorder is characterized by (i) a rapid adult worker bee losses in the affected hives resulting in either dead or weak colonies with a characteristic high brood population compared to the adults (ii) a noticeable absence of dead workers both within and around the affected colonies (iii) a delay in the invasion of the hives by pests as well as delayed cleptoparasitism by neighboring colonies and (iv) a noticeable absence of *Nosema* spp loads and *Varroa destructor* loads at levels known to cause economic damage.

Apart from CCD, several other factors have also been postulated as possible drivers of the observed colony declines. These include low genetic diversity (Tarpay, 2003), change in land usage, use of pesticides (Johnson *et al.*, 2010), parasites and pathogens (VanEngelsdorp *et al.*, 2009), climate change, poor nutrition and poor colony management strategies (Watanabe, 2008)

A report by the United States Department of Agriculture (USDA) shows that most colonies exhibiting poor health conditions as well as symptoms of CCD have a characteristic

increase in the viral and/or bacterial loads as well as pesticide residues, in comparison to their non-CCD counterparts (CCD Steering Committee, 2012b). Studies have also linked low colony fitness to a reduction in the intra-colonial genetic diversity making the colonies more susceptible to pests and diseases (Tarpay, 2003). Understanding the honeybee intracolonial diversity is important as it informs the levels of colony fitness. Genetically diverse colonies exhibit diverse sub-families of workers that result in colony behavioral diversity as well as increased colony efficiency in relation to fitness, productivity and survival (Mattila and Seeley, 2007).

The first evolutionary history of honeybee, *Apis mellifera* (Linnaeus, 1758) was defined using morphometric parameters (Ruttner *et al.*, 1978). Since then numerous approaches have gained usage in characterizing honeybees successfully classifying them into approximately 30 sub-species and four major groups (Péntek-Zakar *et al.*, 2015). These approaches have incorporated both the traditional morphometric parameters (Ruttner, Tassencourt and Louveaux, 1978) as well as the use of molecular markers such as microsatellite and mitochondrial markers (Garney *et al.*, 1993; Solignac *et al.*, 2003a).

Various microsatellite markers have been characterized (Estoup *et al.*, 1993; Solignac *et al.*, 2003a) and since gained wide application in the determination of nuclear genetic differences that exist within various honeybee sub-species in different studies ( Franck *et al.*, 2001; Desai and Currie, 2015; Kraus *et al.*, 2007; Loucif-Ayad *et al* 2015; Mattila and Seeley, 2007b; Palmer and Oldroyd, 2003; Péntek-Zakar *et al.*, 2015; Rasoloforivao

*et al.*, 2015; Techer *et al.*, 2015a; Techer *et al.*, 2015b; Techer *et al.*, 2015c; Techer *et al.*, 2017a; Techer *et al.*, 2017b)

Extensive studies exist on the genetic diversity of honeybees in most of the islands to the SWIO. However, limited knowledge exist on the genetic diversity and population structure of honeybees native to the Comoros islands where most colonies exist as wild populations. Attempts to study the honeybees of Comoros relied on sampling foragers on flowers at a distance of 5 km (Techer *et al* 2017a, 2017b) which might not offer a true representation of the colonies present in the islands as there is a high likelihood of over-sampling individuals from the same colony. In addition, given the fact that Comoros islands is majorly populated by wild honeybee populations, it was of much interest to determine the level of genetic diversity and population structure exhibited by the honeybees population under minimal human interference. This study employed extensive sampling to determine the nuclear and mitochondrial genetic diversity as well as the population structure of honeybees across three main islands of the Comoros archipelagos (Ngazidja, Anjouan and Moheli).

## 1.2 Statement of the problem

Honeybees offer important pollination services vital for food production, ecological biodiversity, world economy, nutrition, and food security (Klein *et al.*, 2007). Gradual declines in honeybee populations have been reported majorly in the northern hemisphere (McMenamin and Genersch, 2015). Consumer surplus losses associated to this decline were last estimated to range between USD 202 to 329 billion per annum (Gallai *et al.*, 2009) and the figure is likely to have increased over time.

Decline in honeybee population is majorly associated with three broad factors namely pests and pathogens, environmental stressors and genetic diversity and vitality (Potts *et al.*, 2010). All these factors work as either individual entities or in synergy to impact the honeybee health (Quarles, 2008; Watanabe, 2008; VanEngelsdorp *et al.*, 2009; Johnson *et al.*, 2010). Most studies on the genetic diversity and population structure of honeybees have relied on managed honeybee populations. As such, there is limited knowledge on the genetic diversity and population structure of wild honeybee populations existing within a fragmented landscape under minimal human interference.

## 1.3 Justification

Genetically diverse colonies exhibit diverse worker sub-families which confers behavioral diversity to the colony, leading to an increase in colony level efficiency and fitness in terms of acquiring food, and response to pests and diseases (Mattila and Seeley, 2007; Oldroyd *et al.*, 1992; Tarpy *et al.*, 2013). The impact of low genetic diversity on most managed colonies has been studied and the results indicate that a reduced level of genetic diversity at the intra-colonial level affects the fitness of the colony making the colonies

more susceptible to pests and diseases (Tarpy *et al.*, 2013) as well as less efficiency in food acquisition, which results into less productive colonies. Although the use of morphological characteristics is still considered a significant approach, it lacks the power to characterize honeybees into distinct subspecies (Franck *et al.*, 2000). Furthermore, it does not elucidate the phylogenetic relationship among them. This can be attributed to the fact that they are subject to manipulation by environmental selection pressures and introgression resulting from hybridization between sub-species (Franck *et al.*, 2000). Specific molecular markers such as the mitochondrial and neutral nuclear genome markers have given new insights into the successful classification of honeybees into subspecies. Sequencing of the COI-COII intergenic region (Garney *et al.*, 1993) has successfully classified honeybees in accordance with the Ruttner's classification which was based on morphological characters. Furthermore, new sub-species have been identified and added to the Ruttner's list.

Although a recent study on the genetic diversity of honeybees native to the South-West Indian Ocean islands describes both the nuclear and mitochondrial diversity in Comoros islands, the study lacks power owing to low samples collected from Comoros archipelago between the year 2013 and 2015. For instance due to the small sample size, it was not possible to compute pairwise  $F_{ST}$  values. Though, the population portrayed differentiation, the Moheli samples did not significantly differentiate with the authors citing low sample size coupled with high null allele number frequencies as the possible causes (Techer *et al.*, 2017a; 2017b). Very few samples per colony were used which limited the ability of the study to determine the diversity at the colony level.

Understanding the genetic diversity and population structure of honeybees is of significance as the knowledge generated informs conservation strategies for endemic honeybee species within their natural habitats and protect them from human interference. It also enables the identification of beneficial genotypes within a population that can be used for breeding purposes. Breeding is key to sustainable beekeeping program

#### **1.4 Hypothesis**

Honeybees of Comoros islands of Ngazidja, Anjouan and Moheli exist as a large homogeneous population with a single mitotype, low genetic diversity, non-structured and undifferentiated

#### **1.5 Objectives**

##### **1.5.1 General objective**

- i. To determine the mitotypes, genetic diversity, population structure and genetic differentiation of honeybees in Comoros islands using mitochondrial and microsatellite markers

##### **1.5.2 Specific objectives**

- i. To identify the mitotypes of honeybee colonizing the Comoros islands through COI-COII intergenic region sequence analysis
- ii. To identify the genetic diversity, population differentiation and population structure using microsatellite DNA markers

## CHAPTER TWO

### LITERATURE REVIEW

#### **2.1 Classification of honeybees**

Honeybees, *Apis mellifera*, are classified under the Kingdom, Animalia: Phylum, Arthropoda: Class Insecta: Order, Hymenoptera: Family, Apidae: Genus, *Apis*: and Species, *mellifera* (Ruttner, 1988). They are also classified in the Aculeata section of hymenopterans - hymenopterans whose females possess stings. The Aculeata section include bees, wasps and ants. Bees exhibit similarities to the sphecoid group of wasps but generally more unlike the rest of Aculeata, some bees are more hairy and robust when compared to the wasps. In contrast to sphecoid wasps, honeybees do not capture other insects and spiders as their protein source but they rather depend on flowers as the sole source of proteins needed for the development of their larvae and ovaries. The sphecoid wasps and honeybees together constitute the Apoidea superfamily which is further subdivided into two distinct groups i.e. the spheciiformers and the apiformers. Honeybees are grouped among the apiformers group of the Apoidea superfamily (Michener, 2000).

Apart from *Apis mellifera*, which is a cavity nesting honeybee, other honeybee species also exist. These include, *Apis koschevnikovi*; *Apis nuluensis*; *Apis nigrocincta* and *Apis cerana* classified among the cavity nesting bees; *Apis florea* and *Apis andreniformis* categorized among the dwarf honeybees, and *Apis dorsata* and *Apis laboriosa* grouped among the giant honeybees i.e. (Michener, 2000). Apart from *Apis mellifera* all the other species are endemic to Asia.

## 2.2 History and the origin of honeybees

The honeybee is one of the most successful species in the Kingdom Animalia due to its ease of adapting to new environmental conditions. It is widely thought that honeybees evolved in the tropics during the mid-tertiary period of the Pleistocene epoch and rapidly evolved and spread fast to various parts of the world (Ruttner, 1988)

The evolutionary origin of *Apis mellifera* was proposed based on three main scenarios. The first scenario as proposed by Ruttner *et al.* (1978) postulates that *Apis mellifera* originated from the Middle East and Northern Africa before colonizing Europe following two main routes namely (i) the western route through North Africa and (ii) through the Iberian Peninsula. This hypothesis was based on morphological analysis and suggested continuity between the African lineage (A) and the M lineage and an ancestral form of *Apis mellifera syriaca* from Israel, Lebanon and Jordan. The second hypothesis by Cornuet and Garnery (1991) and Garnery *et al.*, (1992) also proposes *Apis mellifera* to have originated from Middle East but it does not suggest colonization via the Western route. This second scenario is strongly based on phylogenetic analysis in which the A lineage and the C lineage are grouped together instead of the M, contradicting the migration via the strait of Gibraltar. A third hypothesis suggests an African origin. This hypothesis capitalizes on the ability of domesticated *Apis mellifera* to form winter clusters representing a derived adaptation to temperate climates (Wilson, 1971). The hypothesis is based on the observation that *Apis mellifera* does not colonize the tropical Asia, it is more likely the proposed ancestral form had an African origin.

Biodiversity of honeybees was originally determined using morphometric analysis that was entirely based on extensive collection of samples followed by multivariate analysis of morphometrical characters (Ruttner *et al.*, 1978). This identification technique was long considered to be definitive of the honeybees. Through this classification approach, the North-Eastern Africa and the Middle East were established as the area of origin for *Apis mellifera*. From their centre of origin, the species are thought to have invaded Africa and Europe following three distinct patterns that informed their classification. These are the South and Central Africa Branch (A), North African and North European Branch (M) and the North Mediterranean branch (C). The classification was further improved by the addition of the fourth evolutionary branch known as the Oriental (O) branch, which invaded the Near and Middle Eastern sub-species (Ruttner, 1988). Most recently two more branches have been described as the Y lineage from Ethiopia and the Z lineage from Syria (Franck *et al.*, 2001; Ruttner, 1988; Alburaki *et al.*, 2013).

Despite the fact that honeybee, *Apis mellifera*, naturally inhabits Europe, Africa and the Middle East up to nine other *Apis* species are exclusively found in Asia hence , *Apis mellifera* is widely assumed to have originated from Asia. However, some hypothesis based on phylogenetic analysis using genetic markers, particularly the analysis of over 1000 single nucleotide polymorphisms (SNPs), placed the root tree of *Apis mellifera* subspecies to samples originating from Africa suggesting that *Apis mellifera* bares its origin from Africa (Whitfield *et al.*, 2006). Re-analysis of the same data by Fan *et al.* (2012), failed to support the initial findings which had suggested an out of Africa origin and moved on to strongly link its origin to Asia. The latter findings are corroborated by the

studies by Wallberg *et al.*, (2014) who suggested an out of Asia origin citing the fact that all other honeybee species are almost exclusively found in Asia.

The lineage *Apis mellifera* is thought to have split from other cavity-nesting bees and later diversified into the present subspecies eventually colonizing their present habitat. Morphometric and molecular analysis have successfully grouped the honeybees into five evolutionary lineages and further into over 29 subspecies ( Ruttner, 1988; Franck *et al.*, 2001). Studies based on the mtDNA and nuclear DNA genetic diversity suggest the first split of honeybees to have occurred about 6-9 million years ago ( Cornuet and Garnery, 1991; Arias and Sheppard, 1996). Contrary to this, genetic variations observed among the extant subspecies show some degree of similarity and highly suggest that they have not experienced long isolation periods. Based on genetic dating, the four major lineages (A, M, O and C) are thought to have diverged about 0.7-1.3 million years ago ( Garnery *et al.*, 1992; Arias and Sheppard, 1996). More recently, Wallberg *et al.* (2014) used whole genome sequencing to give insights into the evolutionary history of honeybees. In their work, the A, C and M lineages are shown to have diverged about 300,000 years ago. Furthermore, they show that the O and C lineages separated more recently at a period of about 38,000 years from the present. However, they suggest the given dates as the minimum divergence times citing the possibility of continued gene flow among the honeybees despite longer divergence periods. Little data is available to infer the geographical ranges inhabited by the honeybees from the time they split from the other cavity nesting bees to their present habitat. Morphological and genetic relationships among the subspecies can however give a clue to the timing and location of their common origin (Fan, *et al.*, 2012).

### 2.3 Genetic diversity in honeybees and its importance

Hymenopterans exhibit a haplodiploid system of sex determination where males are monogamous while females are polygamous. In this system, fertilized eggs develop into females while the unfertilized eggs develop into males. Sex is determined by constitution of alleles at the sex determining locus. In the case of heterozygosity within the sex determining region, females will develop, while males develop from homozygous eggs. However, diploid eggs which are homozygous at the sex determining locus results in diploid male offspring. Such offspring are reproductively useless as they tend to produce few sperm cells hence most of them are killed at the larval stage of development among the *Apis mellifera*. Those that survive to maturity produce triploid offspring that lack reproductive potential.

The honeybee mating strategy is of increasing interest. During the mating seasons, drones fly out of the hive and congregate high in the air at distinct locations (Ruttner, 1966; Vallet and Coles, 1993). These zones of congregation are termed as Drone Congregation Areas (DCAs) and are comprised of drones that originate from different colonies (Baudry *et al.*, 1998). Generally, drones arising from apiaries that are within an interval of 5 km apart visit the same DCA. As such, each DCA has a mixed population of drones that ultimately contribute to higher diversity. Most recently, drones originating from up to 238 colonies have been detected in the same DCA (Baudry *et al.*, 1998). After emergence, the young queen flies out for mating. The mating of the virgin queens usually takes place high in the air at the DCA during a mating or nuptial flight. This occurs at the age of 5-10 days from emergence of the queen. During the mating flight virgin queens perform one or

two orientation flights next to the hive, which last for about 2 minutes before finally flying to the DCA, where they copulate with several drones, usually 10-20 drones (Schlüns *et al.*, 2005), that in turn deposit their male gametes in the queen's lateral oviduct. The mating flight generally lasts for 10 -30 minutes, and upon return to the hive, 5% of the sperm stored in the oviduct is transferred to the spermatheca of the queen in a filling process which takes up to 24 hours (Woyke, 1962). The queen then uses the sperm to fertilize eggs as it lays them. The fertilized eggs becomes females and the unfertilized eggs develop into drones.

Genetically diverse colonies arising from the polyandry mating of the queen, at the DCA, exhibit diverse worker sub-families which confers to the colony behavioral diversity and this increases the colony level efficiency in terms of fitness, survival and productivity (Oldroyd *et al.*, 1992; Mattila and Seeley, 2007). Workers from colonies with low genetic diversity on the other hand portray a narrow range of behavioral threshold. Studies have indicated that a high level of genetic diversity at the intra-colony level may lower the chances of infestation with parasites and pathogens (Schmid-Hempel, 1995; Sherman *et al.*, 1988; Tarpy, 2003). It is also postulated that close kinship among colonies increases the chances of acquiring a parasitic infection among the colonies as compared to the genetically diversified colonies (Shykoff and Schmid-Hempel, 1991). Colonies with no or very low genetic diversity are advantageous in the perspective of the parasites as they are more susceptible to parasitic colonization compared to colonies with a high genetic diversity. Colonies originating from a single paternity have also been shown to have a charac-

teristic lower production of drone brood, lower honey and pollen storage, as well as a fewer numbers of queen cells (Fuchs and Schade, 1994).

A recent study has shown that genetically diverse colonies show a greater mite mortality rate as compared to genetically similar colonies, an observation that is specific to a certain season, winter, of the year (Desai and Currie, 2015). Desai and Curies (2015) also reported that for some pathogens, the pathogenic load tend to be higher in genetically similar colonies as compared to the genetically diverse colonies. Intra-colonial genetic diversity therefore benefits the colony by conferring some advantageous phenotypic characteristics that includes hygienic behavior, grooming behavior as well as other mechanisms which reduce the transmission of pathogens and parasites.

#### **2.4 Characterization of honeybees based on mitochondrial DNA**

The honeybees' mtDNA is a small circular molecule ranging between 16 500 to 17, 000 base pairs long. It is uniparentally transmitted by the queen to her offspring (drones and workers) thus expected to be non-recombinant. As a result, a single honeybee is sufficient to determine the mito-haplotype of all honeybees in a colony. When using mtDNA to characterize honeybees, it is ideal to select a sample from an emerging adult or a pupa in order to eliminate the risk of picking and analyzing adult workers that may have drifted between colonies (Evans *et al.*, 2013).

The different mitochondrial lineages are inferred from the observation that mitochondrial DNA of honeybees contains a COI-COII intergenic region that exhibits an enormous

amount of length polymorphism. The region is also characterized by sequence variations that, in addition to the different fragments produced by endonuclease cleavage, are used to extensively distinguish among honeybee lineages as well as discriminate among subspecies (Garnery *et al.*, 1992; Franck *et al.*, 2000; Sheppard and Smith, 2000). The COI-COII intergenic region exhibits at least seven distinct length variants that can be explained by a unique combination of three distinct sequences i.e. Po (67 bp), P (54 bp) and Q (192-197 bp). Different honeybee haplotypes contain different combinations of these sequences which are; PoQ, PoQQ, PoQQQ, PQ, PQQ, PQQQ and Q. The variability in length combined with availability of restriction site polymorphism has been utilized to develop a simple and rapid test that is used to characterize honeybee haplotypes based on mitochondrial DNA (*Dra*I test) (Garnery *et al.*, 1993).

## **2.5 Characterization of honeybees based on microsatellite DNA**

Microsatellites are short tandem repeats found in most species genomes and are used as informative markers in the field of population genetics, conservation biology and evolutionary biology. They are generally defined as short tandem repeats (STRs) or simple sequence repeats (SSRs) comprising 2-10 base pair units and present themselves as either perfect or imperfect repeats within an organism. Microsatellites exhibit high variability in terms of the number of repeats found in every microsatellite region. Current knowledge considers microsatellites as the most useful genetic markers in both population and quantitative genetics. The markers are biparentally inherited thus making them more useful tools in parental determination and genetic polymorphic analyses (Estoup *et al.*, 1995). A polymorphic microsatellite has more than one potential allele at a given locus. Given the fact that they are codominant, Mendelian inherited and neutral markers, microsatellites

are easily typed. In addition, they have a high distinctive power among closely related individuals (Abdul-Muneer, 2014) ultimately making them suitable candidates for determining the population structure.

Microsatellites have previously been used to characterize honeybee populations belonging to the African and European origin (Estoup *et al.*, 1995). Estoup *et al.* (1995) scored alleles for seven (7) microsatellite loci among various subspecies and reported a high degree of genetic variation within the honeybee samples. The variation ranged between seven (7) to thirty (30) alleles per locus. The African subspecies were also shown to have a higher genetic variation as compared to those of the European origin. Also, of importance to note is that super sisters i.e. honeybees from a similar patriline, clustered together as opposed to half-sisters. This would not be the case using mtDNA given its maternal inheritance nature which would result in all super-sisters and half-sisters arising from same queen clustering together.

The microsatellite results, therefore, makes it possible to determine heterozygosity observed within a colony. Microsatellite based studies reveal moderate genetic diversity among honeybees (*Apis mellifera unicolor*) in Madagascar characterized by low allele number per locus in comparison to South Africa (Rasoloforivao *et al.*, 2015); detect no distinct genetic structure in Rodrigues island but shows the genetic diversity in the island to be closely related to continental populations (Techer *et al.*, 2015). Lastly, they reveal a highly structured population among the honeybees in Seychelles (Techer *et al.*, 2015).

Seychelles is an Archipelago composed of three main islands (Mahe, Praslin and La Digue) - a situation similar to Comoros islands (Ngazidja, Moheli and Anjouan). Based on microsatellite data, the honeybee population in Seychelles exhibit a significant nuclear similarity to *A.m unicolor* from Madagascar.

## **2.6 Genetic diversity of honeybees to the South West of Indian Ocean Islands**

Current studies on the genetic diversity of the honeybees native to the South West of the Indian Ocean Islands (SWIO) give mixed results ( Techer *et al.*, 2014; Rasolofaarivao *et al.*, 2015; Techer *et al.*, 2015a; Techer *et al.*, 2017a; 2017b). Techer *et al.* (2014) reported the absence of the African lineage in Rodrigues. This study also identified the existence of three haplotypes belonging to the C lineage. The C1 haplotype (*Apis mellifera ligustica*) was the most prevalent (81.3%). The C2 (*Apis mellifera carnica*) haplotype was reported in 18.0% of the study population. The study also revealed the existence of a new haplotype which was named C1-Rod.

However, the study did not detect any new genetic structure. In addition, no difference was noted in the genetic diversity in comparison to the continental populations. Further analyses of these suggested the possibility of the occurrence of a bottleneck scenario in the island. These findings are different from those in Seychelles islands ( Techer *et al.*, 2015) where all the mtDNA sequences belonged to the African lineage with 96.7% of the study population belonging to the A1 (*Apis mellifera unicolor*) sub-lineage. In addition, sub-lineage Z was uniquely identified in two islands (Praslin and La Digue) that comprise part of the Seychelles archipelagos. Seychelles was also shown to have a high and well-

structured nuclear genetic diversity which signifies a restricted gene flow between the islands. A more notable observation was the high nuclear similarities between the honeybee populations in Seychelles and *Apis mellifera unicolor* that is native to Madagascar (Techer *et al.*, 2015). The study by Rasoloforivao *et al.* (2015) showed that the honeybees of Madagascar are majorly comprised of A1 sub-species (99.5%) and 0.4% of A4 haplotype. Madagascar, also, has a unique A lineage haplotype which comprised of 0.2% of the study population. Microsatellite DNA analysis showed presence of moderate genetic diversity among the Madagascar's honeybees populations.

Most recently, large scale mitochondrial DNA sequencing has revealed the existence of a new private A1 haplotype that is shared between Madagascar and all the islands forming the SWIO archipelagos (Techer *et al.*, 2017a; 2017b). Presence of private African haplotypes in each of the archipelagos was used to suggest diversity radiation in each of the archipelagos. In addition, Comoros archipelagos were established as the possible contact point between the continental African and the insular populations ( Techer *et al.*, 2017a, Techer *et al.*, 2017b). These studies showed that all the SWIO ocean islands except Rodriguez are populated by the African lineage of the honeybee sub-species. The study by Techer *et al.*, (2017a) suggested complete dominance of the A lineage in Madagascar, Seychelles and Comoros islands. La Reunion and Mauritius had 95.5% and 56.1% of the A lineage respectively. The European lineages C and M were uniquely reported in the Mascarene archipelagos. A striking difference in their distribution was noted with La reunion having 4.6% whereas Mauritius had 44% (Techer *et al* 2017a).

## 2.7 Hardy Weinberg Equilibrium (HWE)

The Hardy Weinberg equilibrium describes a population that is not evolving (ideal population) where evolution refers to changes observed in the allele frequencies in a population or a gene pool. It is based on the Hardy Weinberg principle which assumes absence of external disturbances to a population thus inferring constant genetic variations to the population. This principle is based on seven assumptions, which include: (i) No selection (ii) Absence of mutation (iii) Absence of gene flow (iv) Presence of an infinite population (v) Random mating (vi) Each individual producing similar number of offspring and (vii) No migration occurring into and out of the population.

Mathematically a population that is in conformity to the HWE equilibrium assumes the equation

$$p^2 + 2pq + q^2 = 1$$

Where  $p^2$  = probability of having the dominant homozygous alleles (**AA**),  $q^2$  = probability of having dominant recessive alleles (**aa**) while  $2pq$  = the probability of having heterozygosity in the allele composition (**Aa**)

This equation can be further simplified to show that  $p$  represents all individuals that are homozygous dominant (AA) and half the individual who are heterozygous (Aa) while  $q$  represents individuals who are homozygous recessive and half of those who are heterozygous (aA).

In such, the equation is rewritten as

$$p = AA + \frac{1}{2}Aa$$

**While**

$$q = aa + \frac{1}{2}aA$$

**Therefore**

$$p + q = 1$$

A population is assumed to be in a HWE if the frequency of A ( $p$ ) and a ( $q$ ) are both maintained at 50 % over generations if that was the case represented in the parental genotype. Deviations from this equilibrium is an indicator that the population is evolving.

## 2.8 Advantage of using $G_{ST}$ to determine population differentiation

$G_{ST}$  is an improvement to the Wright's  $F_{ST}$  statistics that was originally developed with the aim of analyzing biallelic data.  $G_{ST}$  has a higher capacity to analyze multiallelic data than  $F_{ST}$ . Based on this measure of diversity  $G_{ST} = 0$  indicates that the population is not differentiated and the more the  $G_{ST}$  approaches 1 the more differentiated the population. A population with a  $G_{ST}$  value of 1 indicates the fixation of that given allele in the population.

$G_{ST}$  is mathematically represented as (Nei, 1987);

$$G_{ST} = (H_T - H_S) / H_T$$

In the equation,  $H_T$  represents the total genetic diversity, while  $H_S$  represents the diversity within the population (this is the equivalent of expected population's heterozygosity in the case of diploids). From the equation above it is clear that  $G_{ST}$  value arises from calcu-

lation of genetic diversity (expected heterozygosity) within and among the study populations. Hedrick's  $G_{ST}$  is an improvement to the initial Nei's  $G_{ST}$  (Nei, 1972) with corrections for the latter's weaknesses (Hedrick, 2005).

## **2.9 The Principle of Discriminant Analysis of Principle Components**

Discriminant Analysis of Principal Components (DAPC) is a multivariate STRUCTURE-like statistical approach of inferring the population structure among genetically related individuals (Jombart, Devillard and Balloux, 2010). The method works based on an assumption that all markers are in linkage disequilibrium and that a panmictic population exists (Pritchard *et al.*, 2000) i.e. all individuals in the populations are randomly mating.

The approach divides the sample variance into within-group and between-groups components as a way of maximizing discrimination between groups. The method first transforms data using principal component analysis (PCA) and then employs discriminant analysis (DA) to identify subsequent clusters.

## CHAPTER THREE

### MATERIALS AND METHODS

#### **3.1 Study site**

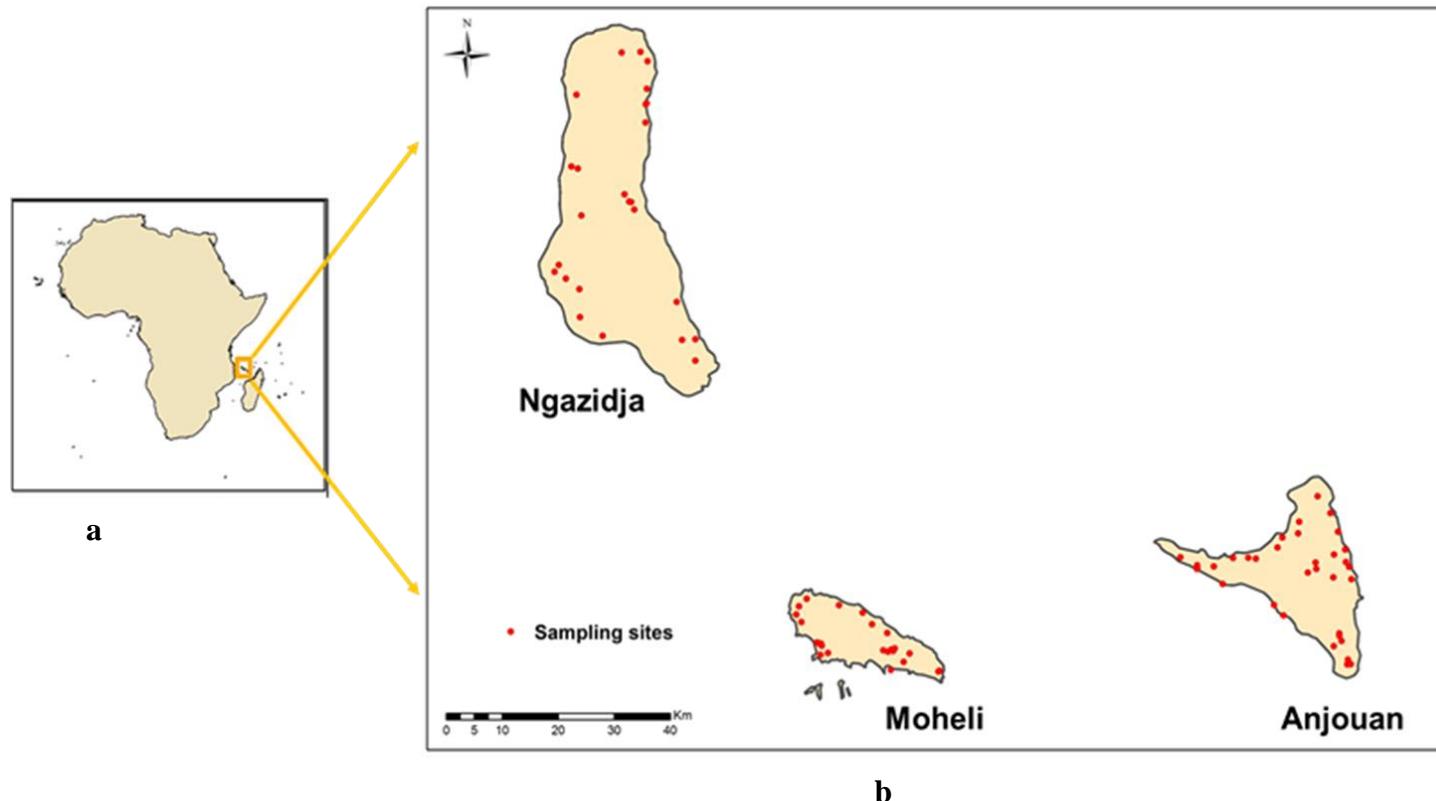
Comoros, an archipelago made up of volcanic islands, is located off the South-East coast of Africa to the eastern side of Mozambique and North-West of Madagascar ( $12^{\circ}10'S$   $44^{\circ}15'E$ ). It is made up of three main islands namely Grande Comore or Ngazidja ( $1,025\text{ km}^2$ ), Anjouan or Nzwani ( $424\text{ km}^2$ ) and Moheli or Mwali ( $211\text{ km}^2$ ). The sampling sites on the three main islands are represented below (Figure 3.1)

#### **3.2 Sampling**

One hundred and sixty (160) honeybee colonies were sampled across the different geographic locations in the Comoros archipelagos in June 2017. These included forty-eight (48) colonies from Ngazidja, fifty two (52) colonies from Anjouan and sixty colonies (60) from Moheli. The sampling was carried out on both managed and feral colonies. Where available, apiaries were sampled and for an apiary to be considered for sampling, it had to be five kilometers away from the previously sampled one since honeybees are known to have a flight range of 5 km and sampling within that range increases the chances of sampling close relatives. Three (3) colonies were randomly sampled from each apiary, and in cases where apiaries had three or less colonies, all colonies were sampled. Feral colonies were also sampled at an approximate distance of 5 km apart and where the feral colonies were within a close proximity they were all treated as an apiary.

One emerging adult per colony was sampled for mitochondrial DNA analysis. The choice of an emerging adult per colony reduced chances of sampling a kleptomaniac from neighboring colonies as would be the case if workers at the entrance were to be considered. Since mtDNA is maternally inherited, a single individual per colony was sufficient to infer the maternal haplotype.

For microsatellite DNA analyses 288 workers were sampled from 12 colonies across the three islands. During sampling 24 workers per colony were randomly sampled from four colonies in each island. The colonies were located in four geographically distinct zones on every island. These included Iconi ( $n=24$ ), Bweni ( $n=24$ ), Ipvwani ( $n=24$ ) and Tsini-moichongo ( $n =24$ ) in Ngazidja ( $N=96$ ); Jimilime ( $n=24$ ), Kambalahari ( $n=24$ ), Nyombeni ( $n=24$ ), and Bougweni ( $n=24$ ) in Anjouan ( $N=96$ ) and Siryziroudrani ( $n=24$ ), Badralaju ( $n=24$ ), Domoni ( $n=24$ ) and Dahoni ( $n=24$ ) in Moheli ( $N=96$ ). (Appendix I). All honeybee samples were immediately stored in 96 % ethanol and placed in cool boxes in the field and transported to the African Reference Laboratory for Bee Health at the International Center of Insect Physiology and Ecology (ICIPE), Nairobi, Kenya where they were stored at -20 °C prior to molecular analyses.



**Figure 3.1:** Sampling sites on the three main islands of Comoros islands. Letter **a** represents location of Comoros on the map of Africa while **b** is an enlargement of the three islands indicating the exact sampling locations. (Source: Emily, icipe GIS Unit)

### **3.3 DNA Extraction from honeybees**

The honeybee samples were placed in a laminar flow chamber which was turned on for 30 minutes to dry off the ethanol before DNA extraction. For mtDNA analysis DNA was extracted from the honeybee thoraces while for microsatellite DNA analyses, DNA was extracted from 24 individual honeybees per colony using Qiagen DNA extraction kit (Qiagen Inc. Germany) following the manufacturer's instructions with slight modifications. Briefly, the thoracic muscle tissue of individual honeybee or the entire bee for mtDNA and microsatellite DNA analyses, respectively, were placed in a 2µl microcentrifuge tube containing 150µl of pre-chilled Phosphate Buffered Saline (PBS) (pH=7.4) (Kakumanu *et al.*, 2016 ) and agitated using a TissueLyser II (Qiagen Inc. Germany). DNA was extracted from 100µl of the homogenate following all the manufacturer's instructions. The integrity of the extracted DNA was checked through gel electrophoresis prior to PCR amplification. The concentration of the extracted DNA was checked using a nanodrop and standardized to 50ng/µl.

### **3.4 Mitotypes of honeybee colonizing the Comoros islands**

#### **3.4.1 PCR amplification of COI-COII intergenic region**

Total DNA was subjected to PCR amplification using the E2 (forward) (5'-GGCAGAATAAGTGCATTG-3') and H2 (reverse) (5'-CAATATCATTGATGACC-3') primers described in Garnery *et al.* (1993). The primers targeted the mitochondrial COI-COII intergenic region that has been extensively used in determining the subspecies and evolutionary lineages among honeybees (Garnery *et al.*, 1993; Magnus *et al.*, 2011; Rasofoarivao *et al.*, 2015; Techer *et al.*, 2015a, 2015b).

A 15 $\mu$ l PCR reaction containing 1 x Phusion High Fidelity Taq polymerase with HF buffer, 0.4  $\mu$ M of each of the primers, 0.5 $\mu$ l DNA at 50ng/  $\mu$ l was set up (Table 3.1). DNA from a previously amplified COI-COII intergenic region was used as a positive control nuclease free water was served as template control in the negative control reaction. The reaction profile was set up with an initial denaturation step of 98°C for 30 seconds followed by 40 cycles of 98°C for 10 seconds, 48°C for 45 seconds 62°C for 30 seconds and a final extension step at 72°C for 10 minutes in a ProFlex PCR machine (Applied Biosystems).

**Table 3.1:** Components of the 15 $\mu$ l reaction mix used for mitochondrial DNA amplifications

Component	Initial Concentration	Volume per reaction ( $\mu$ l)	Final Concentration
Phusion High Fidelity	2 x	7.5	1 x
Taq polymerase with HF buffer			
Forward Primer (H2)	10 picomoles	0.6	0.4 picomoles
Reverse Primer (E2)	10 picomoles	0.6	0.4 picomoles
DMSO	100%	0.45	3%
Nuclease free water	-	5.35	-
Template	50ng/ $\mu$ l	0.5	1.67 ng/ $\mu$ l

### 3.4.2 Agarose gel electrophoresis and sample purification

Two percent (2%) agarose gel was prepared by adding 2 grams of agarose powder into 100ml of Tris Acetate Ethylenediaminetetraacetic acid (TAE) buffer in a glass conical flask. The mixture was heated in a microwave for 3 minutes to dissolve the agarose powder. The solution was allowed to cool to about 60°C before 2 $\mu$ l of Ethidium Bromide

(EtBr) DNA staining dye was added. It was then swirled and poured on a preset gel casting tray where it was allowed to cool for 20 minutes before being transferred into a gel electrophoresis tank pre-filled with TAE buffer and the combs removed.

The first lane of the gel was loaded with 5 $\mu$ l of 100 bp DNA ladder premixed with 1  $\mu$ l of a 6 x loading dye (New England Biolabs). One microliter of the 6 x loading dye was also mixed with 5 $\mu$ l of the sample on a parafilm before the mixture was loaded into the wells. The gel was run for one hour at 80 volts and later visualized under a UVP GelDoc-It<sup>TS2</sup> Imager (Avantor). The 600bp and 380bp positive samples were selected for purification and subsequent sequencing.

Purification was done using the ExoSapIT purification kit (New Englands Biolabs) following the manufacturer's instructions. Briefly, a 13 $\mu$ l microliter reaction was prepared by adding 3  $\mu$ l of exonuclease I enzyme and Alkaline phosphatase premixed in the ratio of 2:1, respectively into 10 $\mu$ l of the PCR amplicon. The samples were then incubated at 37°C for 15 minutes followed by 80°C for 30 minutes to inactivate unused enzymes.

### **3.4.3 Sequencing of the COI-COII intergenic region**

Ten microliters (10 $\mu$ l) of the purified samples was packaged in a 1.5 $\mu$ l microcentrifuge tube and sent to Macrogen Inc. (Amsterdam, Netherlands) where bidirectional sequencing was done using the Sanger chain termination technology.

### 3.4.4 COI-COII intergenic region sequence analyses

The resultant sequences were edited and consensus sequences for the forward and reverse strands generated using BioEdit version 7.2.5 (Hall, 1999). COI-COII sequences belonging to different species of honeybees *Apis mellifera* (AY587542.1), *Apis dorsata* (AY588415.1), *Apis Koschevnikovi* (AY587546.1), *Apis laboriosa* (AY587548.1), *Apis cerana* (AY587544.1), *Apis nigrocincta* (AY587545.1), *Apis nuluensis* (AY587543.1), *Apis florea* (AY588416.1) and *Apis andreniformis* (AY588417.1) were mined from the GenBank database in NCBI and included to the consensus sequences for phylogenetic analyses.

In addition, different *Apis mellifera* subspecies i.e. A1\_MAD3 (KT828418), A1\_MYT2 (KT828439), A1\_SEY1 (KT828433), Z2\_SEY1 (KT828478), A1\_MAD15 (KT828430), A1\_GCO1 (KT828435), A1\_GCO2 (KT828436), A1\_ANJ1 (KT828437), A1\_MAD12 (KT828427), Z7\_EGY1 (KT828447), A4\_SEN1 (KT828455), A4\_SEN2 (KT828456), A4\_STP1 (KT828457), A4\_a (KT828448), A66\_STP1 (KT828475), C2\_b (KT828496), C1\_a (KT828493), M7'\_a (KT828492), C1\_Rod1 (KT828494), A4\_ZAF5 (KT828469), M4\_a (KT828487), A6\_TCD1 (KT828474), A1\_MAD15 (KT828430), A1\_MAD12 (KT828427), A11\_PRT1 (KT828479), M7\_a (KT828490) were mined from the GeneBank database and included among the consensus sequences for multiple sequence comparison

These sequences were subsequently aligned using Muscle (Edgar, 2004) on MEGA 7.0.26 platform (Kumar *et al.*, 2012) and a phylogenetic tree constructed with *A. florea* as the outgroup. Prior to constructing the phylogenetic tree, the best nucleotide substitu-

tion model was predicted based on maximum likelihood statistics and a neighbor-joining tree. Gaps/missing data in the sequence alignment were subjected to partial deletion set at a threshold of 95% site coverage. Through this approach the model with the lowest Bayesian Information Criterion (BIC), Tamura 3-parameter, was considered to best describe the nucleotides substitution model (Kumar *et al.*, 2012). The sequences from dwarf honeybees (*Apis florea* and *Apis andreniformis*), which are the most ancient honeybee species (Ruttner, 1988) were used as outgroups. The sequences were grouped according to the various clusters obtained from the phylogenetic tree, and the genetic distances within and between groups calculated in MEGA.

### **3.5 Genetic diversity, population differentiation and population structure**

#### **3.5.1 Microsatellite DNA amplification**

Nineteen previously published honeybees' polymorphic microsatellite loci were scored. These included A113, A24, AC306, AP81, A88, UN351,A56, SEX1, AP273, UN16603, UN4987, UN370, A28, AP289, A124, A35, A8, AP33, and A43 (Estoup, 1994; Estoup *et al.*, 1995; Pierre Franck, Garnery *et al.*, 1998; Shaibi *et al.*, 2008; Solignac *et al.*, 2003a). Forward primers were tagged using fluorescent dyes (FAM, HEX and TET) that absorb and emit light at different wavelengths (Appendix I).

The amplification reaction set up was distributed into six multiplex PCR reactions (Table 3.2). Selection of the plexes was based on the fluorescent dyes and differences in the product size. A fifteen microliter (15 $\mu$ l) multiplex PCR reactions was ran using MyTaq<sup>TM</sup> HS DNA Polymerase (BIOLINE). The final reaction volume contained 1 x My Taq Polymerase reaction buffer, 0.067 units per microliter of MyTaq Hot start Polymerase,

0.2 $\mu$ m of each of the primers, 1 $\mu$ l of DNA at a concentration of 50ng/ $\mu$ l. The reaction was set up with an initial denaturation step of 95°C for 1 minute, followed by 40 cycles of 95°C for 30s, 52°C for 15 seconds, 72°C for 30 seconds and a final elongation step of 72°C for 10 minutes.

**Table 3. 2:** List of microsatellite markers used for multiplex PCR reaction showing how they were combined in the different plexes based on fragment size and the fluorochrome.

Marker	Product range (bp)	Fluorochrome	Multiplex
A113	202-224	FAM	1
AP81	124-136	TET	1
UN351	147-166	FAM	1
A56	270-300	TET	2
AP289	174-288	HEX	2
A88	136-149	HEX	2
AC306	165-181	FAM	3
A28	128-134	FAM	3
A24	93-116	TET	3
A8	165-181	TET	4
AP33	225-247	HEX	4
A35	94-123	HEX	4
SEX1	142-187	HEX	4
UN467.16603	261-282	FAM	5
UN462.4987	168-178	FAM	5
AP273	106-110	FAM	5
A43	124-154	TET	6
B124	216-232	HEX	6
UN467.370	174-192	HEX	6

### 3.5.2 Fragment analysis and allele scoring

The resultant PCR products from different plexes were co-loaded into three different mixes in 96-well plates. Mixes were first selected based on labeling dye, where products labelled with different dyes were distributed across the plates (Table 3.3). In the event that two products labelled using similar dye had to be co-loaded on one plate, then they had to be of different sizes so as to avoid overlapping of peaks during the genotyping process. The mixes were packaged in a 96 well plate and sent to the Institute of Biology, Martin Luther University Halle-Wittenberg in Germany where capillary sequencing was done using MegaBace automatic sequencer.

**Table 3.3:** The primer mix present in each reaction as co-loaded for fragment analysis in the MegaBace automatic sequencer.

MIX 1	MIX 2	MIX 3
A113	AC306	UN467-16603
UN351	A28	UN462-4987
A56	A8	AP273
AP81	A24	A43
AP289	AP33	B124
A88	A35	UN467-370
	SEX1	

MegaBace Fragment Profiler Version 1.2 was used to score the allele sizes at the various loci. Briefly, a peak filter algorithm was generated using expected products ranges shown in Table 3.2. Three filters were generated in accordance with the primer information. Using the algorithm, alleles at various loci were scored. Additionally, alleles were verified by eye and scoring of alleles sizes was manually double-checked and finally exported to an excel sheet. Micro-Checker 2.2.3 (Van Oosterhout *et al.*, 2014) was used to check for genotyping errors including null alleles, stuttering and large allele drop outs. Queen alleles for each locus per colony were manually scored. Scoring was based on an assumption that all colonies were homozygous at each locus.

tion that every member of a colony arises from a single queen hence queen alleles are expected to be represented in each and every worker. Homozygosity and heterozygosity among the workers were inferred based on the methods previously described (Neumann *et al.*, 1999). A homozygous queen was scored when an allele was present across all sampled workers whilst heterozygous queens was considered in the event that different workers in a colony bared either one of the two alleles present in the heterozygous queen.

After scoring the queen allele, the remaining allele was scored as the drone alleles. For homozygous workers, one of the homozygotes was automatically scored as the drone allele while for heterozygous workers the allele different from either of the queen's allele was scored as the drone allele. Determining drone alleles in heterozygous workers portraying both the heterozygous queen alleles involved a multiloci genotyping in order to score drone allele. When multilocus genotyping failed to separate the alleles, the remaining alleles were divided into half and scored as different drones.

### **3.5.3 Determination of queen mating frequency**

Multilocus genotyping approach was used to determine the number of drones in each sample population. Drones were considered super siblings when they portrayed identical alleles at all loci scored while those exhibiting variations at different loci were considered half-siblings. Following this criteria the number of drones that mate with each queen in the different colonies was determined. Due to finite sample size, the observed drones were used to calculate estimate actual number of drones that mate with the queen by fitting in a Poisson distributions while using drones data to calculate the queens mating frequency super siblings were only represented once in order to avoid overestimation of the

mating frequency. Variations in the queen mating frequency between islands was determined using Kruskal-Wallis Test (MacDonald, 2009).

### **3.5.4 Prediction of queens using drones**

The observed drones per island were also used to infer their maternal lineage and sibship assignment in Colony version 2.0.6.4 (Jones and Wang, 2010). For super siblings, only one was used in to avoid overrepresentation of a particular drone genotype. Following this process the queen genotypes in each subpopulation (Anjouan, Moheli and Ngazidja) were scored. Manually scored queen genotypes were included among the predicted queen genotypes parenting more than two drones and used for downstream analyses.

### **3.5.5 Genetic diversity and linkage disequilibrium**

Using *poppr* package in R, observed individuals (N), observed multilocus genotypes (MLG), expected multilocus genotypes (eMLG) given the smallest sample size ( $N \geq 10$ ) based on rarefaction, Shannon-waiver index of MLG diversity (Shannon, 2001), Stoddart and Taylor's Index of MLG diversity (Stoddart and Taylor, 1988), Simpson's index (Simpson, 1949), Evenness, E5 (Grünwald *et al.*, 2003), Nei's unbiased gene diversity (Nei, 1978), the index of association,  $I_A$  (Brown *et al.*, 1980; Smith *et al.*, 1993) and the standardized index of association were determined. Using *poppr* and *magrittr* packages (Kamvar *et al.*, 2014), alleles per locus, allele mean numbers ( $Na$ ) and unbiased expected heterozygosity ( $H_{exp}$ ) were estimated based on Nei's method (Nei, 1978).

To test for deviation from the HWE, *pegas* package was used to compute chi-square ( $\chi^2$ ) statistics and the p-values for the entire dataset (Comoros) as well as for every subpopulation (Anjouan, Moheli and Ngazidja). Using *lattice* package a heatmap showing

deviations from the HWE ( $p<0.05$ ) for every locus in each sub-population was computed. Linkage disequilibrium was determined using *poppr* and *magrittr* packages. The test assumed that all alleles observed at the different loci were linked hence they do not freely recombine during sexual reproduction. The index of association ( $I_A$ ) (Brown *et al.*, 1980) was computed assuming 999 permutations test the hypothesis. Pairwise linkage disequilibrium was further calculated to determine alleles that were in strong linkage.

### **3.5.6 Population differentiation and population structure:**

#### **3.5.6.1 Population differentiation**

Population differentiation was calculated using *mmod* package where Hedrick's standardized measure of genetic differentiation ( $G_{ST}$ ) (Hedrick, 2005) was calculated. The  $G_{ST}$  per locus as well as the global  $G_{ST}$  value per subpopulation were determined. Also calculated was a pairwise  $G_{ST}$  between the subpopulations (Anjouan, Moheli and Ngazidja).

#### **3.5.6.2 Analysis of Molecular Variance (AMOVA)**

Using *poppr* package in R, AMOVA test (Excoffier, Smouse and Quattro, 1992) was performed to detect the level of population differentiation within samples, within and between populations. Test for significance was carried out using the *ade4* package where both the Monte-Carlo tests (Excoffier, Smouse and Quattro, 1992) were performed with 999 permutations.

#### **3.5.6.3 Population structure**

Queen and genotypes were used determine the population structure. Using *poppr* package a neighbor joining tree based on Provesti's distance with 1000 permutations was calculated and visualized using *ape* package. Besides, the population structure was determined using DAPC in R (Jombart, Devillard and Balloux, 2010). Additionally, the population

structure was determined using Bayesian clustering approach implemented in STRUCTURE 2.3.3 (Pritchard, Stephens and Donnelly, 2000)

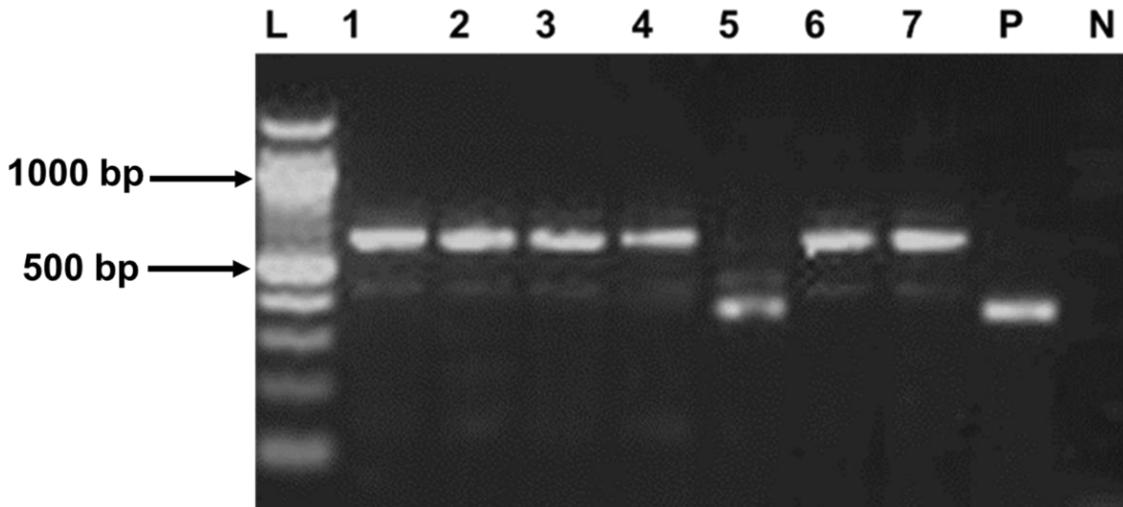
## CHAPTER FOUR

### RESULTS

#### **4.1 Mitotypes of honeybee colonizing the Comoros islands**

##### **4.1.1 Amplification of the COI-COII Intergenic Region**

One hundred and fourteen (114) samples from Ngazidja ( $n=40$ ), Anjouan ( $n=35$ ) and Moheli ( $n=39$ ) were successfully amplified and sequenced. Two band populations were observed across the three islands (4.1.1). One was 380 bp ( $n=28$ ) while the other was 600 bp ( $n=86$ ). The 380 bp mitotypes were distributed such that 16 were in Anjouan, 7 in Moheli and 5 in Ngazidja while the 600 bp mitotypes were distributed in the frequency of 19 in Anjouan, 32 in Moheli and 35 in Ngazidja. A total of 45 samples, though PCR amplified, were not successfully sequenced. These included 17 from Anjouan, 21 from Moheli, and 8 from Ngazidja.



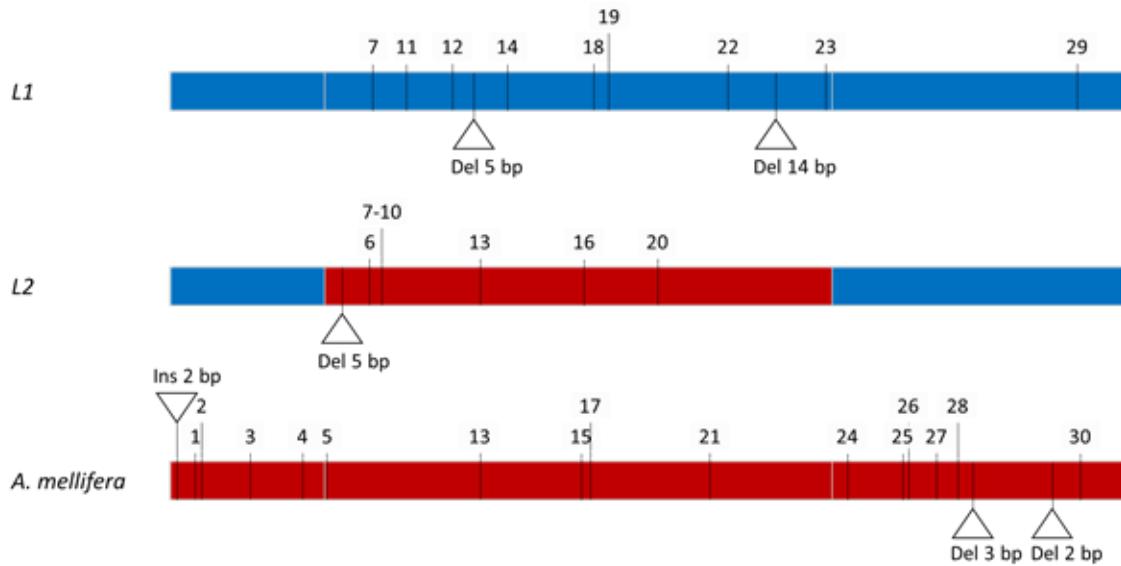
**Figure 4.1:** Gel image showing variations in mitochondrial DNA COI-COII intergenic region of honeybee samples from Comoros islands. **L** represents 100 bp molecular marker, **1-7** represents the samples from different sites, **P** is the positive control and **N** is the negative control.

#### 4.1.2 COI-COII intergenic region sequence analyses

Two sequence populations 600bp and 380bp, respectively were obtained. Sequence alignment revealed the presence of three major differences among these sequences. Based on these differences the sequences were grouped into L1, L2 and A1 haplotypes (Figures 3). L1 and L2 were newly described while A1 corresponded to *A. m. unicolor* haplotypes common to the SWIO. L1 had a distinctive TAGGT indel starting from position 3697-3702 and a second major AAATCATAATATTG indel at position 3791-3805. L2 was characterized by a TCAAAT indel starting from position 3654-3659. It also had a distinctive TTCA insert from position 3669 - 3672. Both L2 and L1 shared common inserts and an indel among themselves. These included a CTT insertion at positions 3861-3863; an AA insertion at position 3886-3887 and an AATTA indel at position 3613-3617. A1 portrayed similarity to the reference sequence. Apart from the described insertions and indels, the sequences expressed distinct single nucleotide polymorphisms (SNPs) among them. The L2 uniquely contained T3666A, T3701A, T3744C and A3759G SNPs. The L1 haplotype exclusively had SNPs T3665C, T3692G, T3738C and G3814A. Both the L2 and L1 haplotypes uniquely had SNPs that distinguished them which were absent in the A1 haplotype. These included C366T, T3643C, G3651C, T669C, T3734A, A3737T, T3776C, T3821A, A3839T, T3841C, G3850C, A3857T, T3897C, T3904A, T3907C, A3918T and A3922T (Appendix III).

The three haplotypes shared regions of high conservation among themselves. Briefly, both L1 and A1, unlike L2, portrayed high conservation of the TCAAAT ancestral sequence at position 3791-3805. L2 and A1 on the other hand, unlike L1, showed high con-

servation of the ancestral TAG located at position 3697-3699. Furthermore, they had a conservation of AAATCATAATATTG at position 3791-3805. The L2 haplotype appeared to have arisen from a rare mtDNA recombination between L1 and the *A. mellifera* haplotype (4.2).



**Figure 4.2:** Recombinant mtDNA haplotype (L2) as indicated by the shared unique indels and single nucleotide polymorphisms between *A. mellifera* haplotype and L1. The different numerical numbers indicates either unique single nucleotide polymorphisms (SNPs) where the number are unique to each segment or shared SNPs where the numbers are similar between two or more segments. Ins means insertion, while Del means deletion.

A maximum likelihood phylogenetic tree rooted at *A. florea* strongly (bootstrap = 97%) separated the giant honeybees (*A. dorsata*, *A. laboriosa* and *A. dorsata bingami*) from the rest. The next separation (bootstrap = 75%) distinguished L1, L2, *A. koschevnikovi* and *A. mellifera* from other cavity nesting bees namely *A. nigrocincta*, *A. cerana* and *A. nuluensis*. *A. mellifera* group was partially separated (bootstrap = 23%) from *A. koschevnikovi*, L1 and L2. *A. koschevnikovi* was partially (bootstrap=31%) separated from L1 and L2. Notably, there was 100% separation between L1 and L2. (Figure 4.3). L1 and L2 clades exceptionally contained sequences from Comoros islands. The L1 and L2 clades corre-

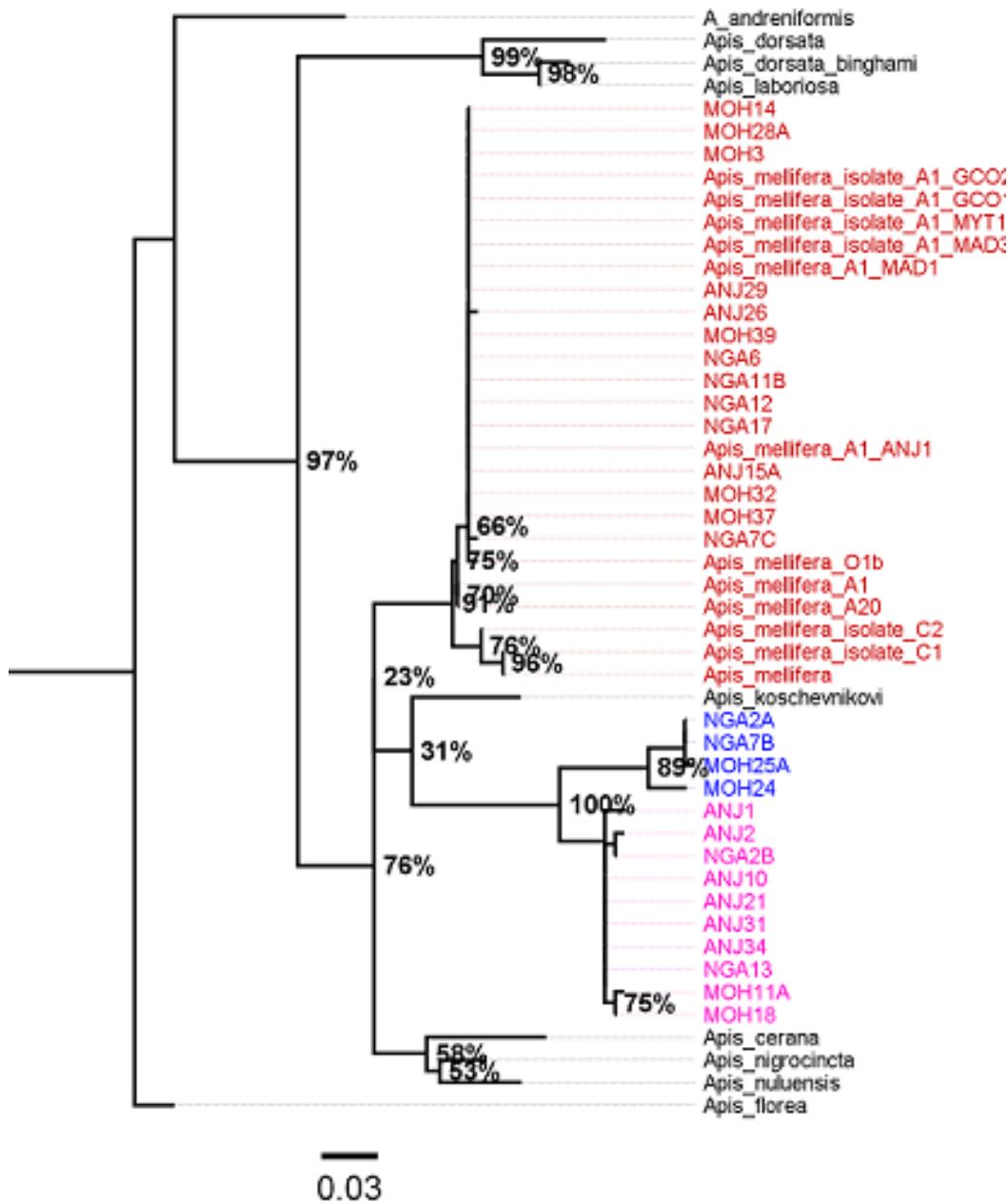
sponded to the L1 and L2 haplotypes respectively whilst the *A. mellifera* group had sequences from Comoros and other *A. mellifera* sequences belonging to the A, C and O lineages.

The sequences from other honeybee species were included in the analysis because preliminary blast search of sequences belonging to L1 and L2 haplotypes revealed close similarity to *A. koschevnikovi*. It was thus important to understand how they cluster among all other honeybee species.

Pairwise genetic distances between groups (Table 4.1) showed that L1 and L2 were closely related but distant from the other groups (*Apis mellifera*, cavity nesting bees, giant honeybees and dwarf honeybees). The genetic distance within groups (Table 4.2) revealed high genetic variability within L2 and L1 compared to *Apis mellifera* represented by samples from A, O and C lineage.

Pairwise genetic distance between groups showed that the distance between L2 and L1 is closer (0.077) than the distance between either L1 (0.107) or L2 (0.116) and *Apis mellifera*. *Apis mellifera* was also shown to be genetically close to the other cavity nesting bees (0.073) as compared to the distance between either of the L2 (0.139) and L1 (0.123) honeybees (Table 2.1). Computing the genetic distance within groups indicated high variability within both L1 (0.0070) and L2 (0.0193) as compared to the variability exhibited in the *Apis mellifera* group (0.0058) (Table 4.2). The variation within the group is how-

ever lower as compared to cavity nesting bees (0.0809), giant honeybees (0.0534) and dwarf honeybees (0.0912) (Table 4.2).



**Figure 1.3:** Maximum likelihood phylogenetic tree showing clustering of samples from Comoros L1 (Pink), L2 (Blue) and *A. mellifera* (Red) sub-species among other honeybee species. The evolutionary history was inferred by using the Maximum Likelihood method and Tamura 3-parameter model. The tree with the highest log likelihood (-970.63) is

shown. The percentage of trees in which the associated taxa clustered together is shown next to the branches. Initial tree(s) for the heuristic search were obtained automatically by applying Neighbor-Join and BioNJ algorithms to a matrix of pairwise distances estimated using the Maximum Composite Likelihood (MCL) approach, and then selecting the topology with superior log likelihood value. A discrete Gamma distribution was used to model evolutionary rate differences among sites (5 categories (+G, parameter = 0.5414)). The rate variation model allowed for some sites to be evolutionarily invariable ([+I], 9.19% sites). The tree is drawn to scale, with branch lengths measured in the number of substitutions per site. This analysis involved 49 nucleotide sequences. All positions with less than 95% site coverage were eliminated, i.e., fewer than 5% alignment gaps, missing data, and ambiguous bases were allowed at any position (partial deletion option). There were a total of 217 positions in the final dataset. Evolutionary analyses were conducted in MEGA X.

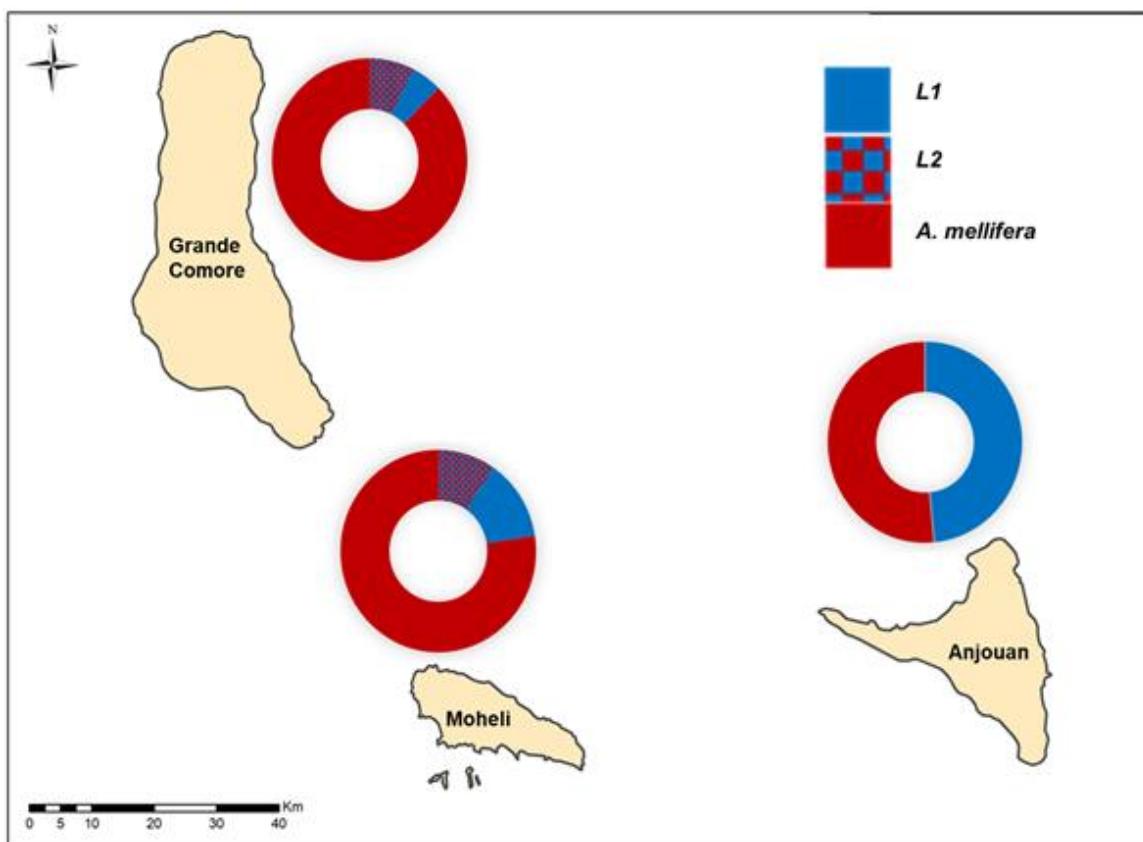
**Table 4.1:** Pairwise genetic distances between groups showing a close relationship between L1 and L2 as opposed to the other groups.

	1	2	3	4	5	6
1. L2						
2. L1		0.077				
3. <i>Apis mellifera</i>	0.116	0.107				
4. Cavity nesting bees	0.139	0.123	0.073			
5. Giant honeybees	0.180	0.184	0.140	0.180		
6. Dwarf honeybees	0.208	0.188	0.140	0.150	0.150	

**Table 4.2:** Genetic distances within groups indicating high genetic variability within L1 and L2 compared to *Apis mellifera* represented by samples from A, C and O lineages.

Genetic distance		
1. L1		0.0070
2. L2		0.0193
3. <i>Apis mellifera</i>		0.0058
4. Cavity nesting bees		0.0809
5. Giant honeybees		0.0534
6. Dwarf honeybees		0.0912

The various haplotypes were distributed in the three islands with L1 and the *A. mellifera* haplotype occurring in Anjouan in an almost equal frequency (4.4) The haplotype (s) were distributed such that Ngazidja had 88% (95% CI = 0.80-0.97) of the *A. mellifera* A1 haplotype; 5% (95% CI = 0.0-0.15) of L1 haplotype and of 7% (95% CI = 0.0-0.17) L2 haplotype. Moheli was characterised by 77% (95% CI = 0.68-0.93) of the normal A1 haplotype, 13% (95% CI = 0.03-0.29) of the L1 haplotype and 10% (95% CI = 0.0-0.26) of the L2 haplotype. Anjouan had 52% (95% CI = 0.36-0.70) of the normal *A. mellifera* haplotype, 48% (95% CI = 0.33 – 0. 67) of the L1 haplotype and 0% (95% CI = 0.00 - 0.10) of L2 haplotype.

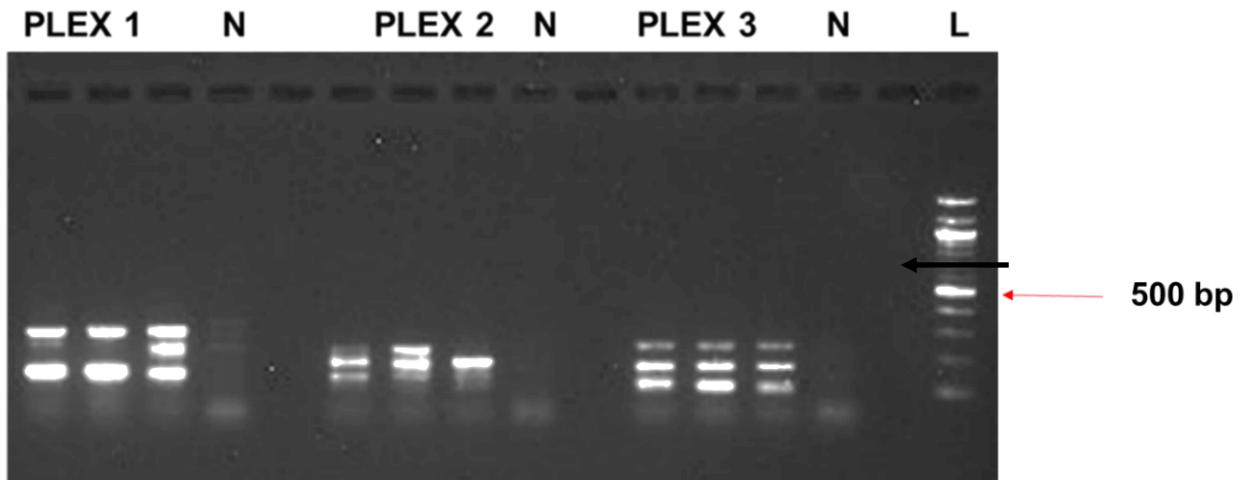


**Figure 4.4:** Distribution of the three mtDNA haplotypes across the Comoro islands.  
(Map Credit: Emily Kimathi, icipe Geo-informatics unit)

## 4.2 Genetic diversity, population differentiation and population structure

### 4.2.1 Microsatellite DNA amplification

Several microsatellite loci were successfully amplified in a multiplex PCR reaction (Figure 4.5).



**Figure 4.5:** Representative multiplex PCR gel image showing the different sizes of amplified fragments using microsatellite markers. N represents the negative control while L represents a 100 base pairs molecular marker (New England Biolabs)

### 4.2.2 Fragment analysis and allele scoring

All the sixteen amplified colonies, four from each island, were successfully genotyped.

Upon fragment analysis, twelve (12) out of the nineteen (19) loci were successfully scored and alleles generated. From the alleles obtained, four queen genotypes per island were manually scored. A minimum of nineteen (19) and a maximum of twenty four (24) drone genotypes in the various sampled colony were recorded (Table 4.3).

**Table 4.3:** Table showing the observed and estimated drone genotypes from the different colonies sampled in individual islands.

Colony	Island	Observed drones	Estimated Drones
ANJ16	Anjouan	22	34
ANJ32	Anjouan	22	34
ANJ5	Anjouan	19	27
ANJ8	Anjouan	22	34
MOH12	Moheli	21	31
MOH18	Moheli	22	34
MOH5	Moheli	21	31
MOH23	Moheli	24	38
NGA15	Ngazidja	20	30
NGA23	Ngazidja	21	31
NGA4	Ngazidja	23	36
NGA7B	Ngazidja	19	27

#### 4.2.3 Determination of queen mating frequency

The observed queen mating frequency ranged between 19 and 24 drones (Table 4.3). Due to finite samples size, estimated queen mating frequency was calculated by fitting in a Poisson distribution. The estimated queen mating frequency ranged between 27 and 38 drones. Kruskal-wallis test found no significant difference in the mating frequencies among the three islands ( $p=0.395$ ).

#### 4.2.4 Prediction of queens using drones

Following the input of drones data into Colony software and manual prediction of queens from workers dataset, 145 queens were retrieved with 59 arising from Anjouan, 38 from

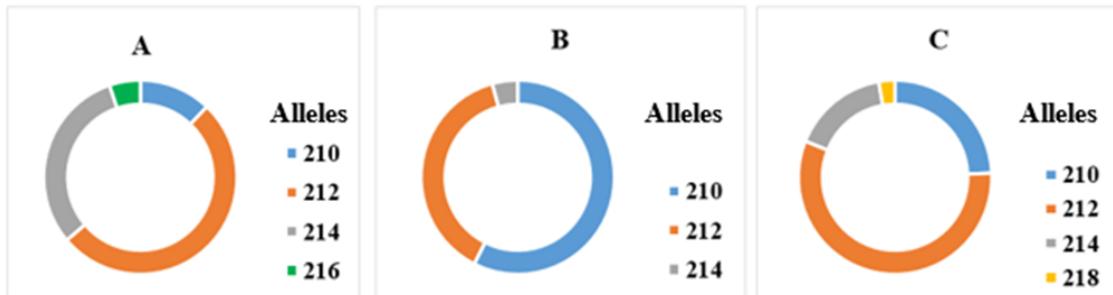
Ngazidja and 48 form Moheli. From these data set, 61 queens parented more than two drones, i.e. 17 from Anjouan, 24 from Moheli and 20 from Ngazidja (Appendix II).

#### **4.2.4.1 Genetic diversity and linkage disequilibrium**

Considering Comoros as a single population, the least number of alleles was recorded at the AP23 locus (2) whilst AP33 locus had the highest number of alleles (12) (Table 9). The mean number of alleles detected was 5.92 ( $\pm 0.05$ ). Simpson's diversity index per locus ranged between 0.29 (AP23) and 0.84 (A306) with a mean of 0.65 ( $\pm 0.05$ ) (Table 9). Expected heterozygosity ( $H_{exp}$ ) per locus ranged between 0.29 (AP23) and 0.85 (AC306).

Comparable genetic diversity was detected between sites. In Moheli the mean number of alleles detected was 3.92 ( $\pm 0.04$ ) whilst Anjouan and Ngazidja had a mean number of alleles of 3.75 ( $\pm 0.05$ ) and 3.67 ( $\pm 0.05$ ), respectively. Simpson's diversity index showed that Ngazidja ( $0.65\pm 0.08$ ) had the highest mean index of diversity followed by Moheli ( $0.59\pm 0.04$ ) with Anjouan  $0.55\pm 0.08$  having the least. Using Nei's 1978 diversity Moheli had the highest expected heterozygosity ( $0.61\pm 0.03$ ), followed by Anjouan ( $0.57\pm 0.07$ ) and Ngazidja ( $0.50\pm 0.10$ ).

Private alleles were recorded at A113 locus where allele 216 was private to Anjouan whilst allele 218 was private to Ngazidja (Figure 4.6). Though not private, allele 108 on AP23 locus was notably absence in Ngazidja (Figure 4.7). Most of the sampled loci showed significant deviations from HWE (Figure 8)



**Figure 4.6:** Allele distribution at locus A113 in Anjouan (A), Moheli (B) and Ngazidja (C) showing the presence of private alleles 216 and 218 in Anjouan and Ngazidja respectively

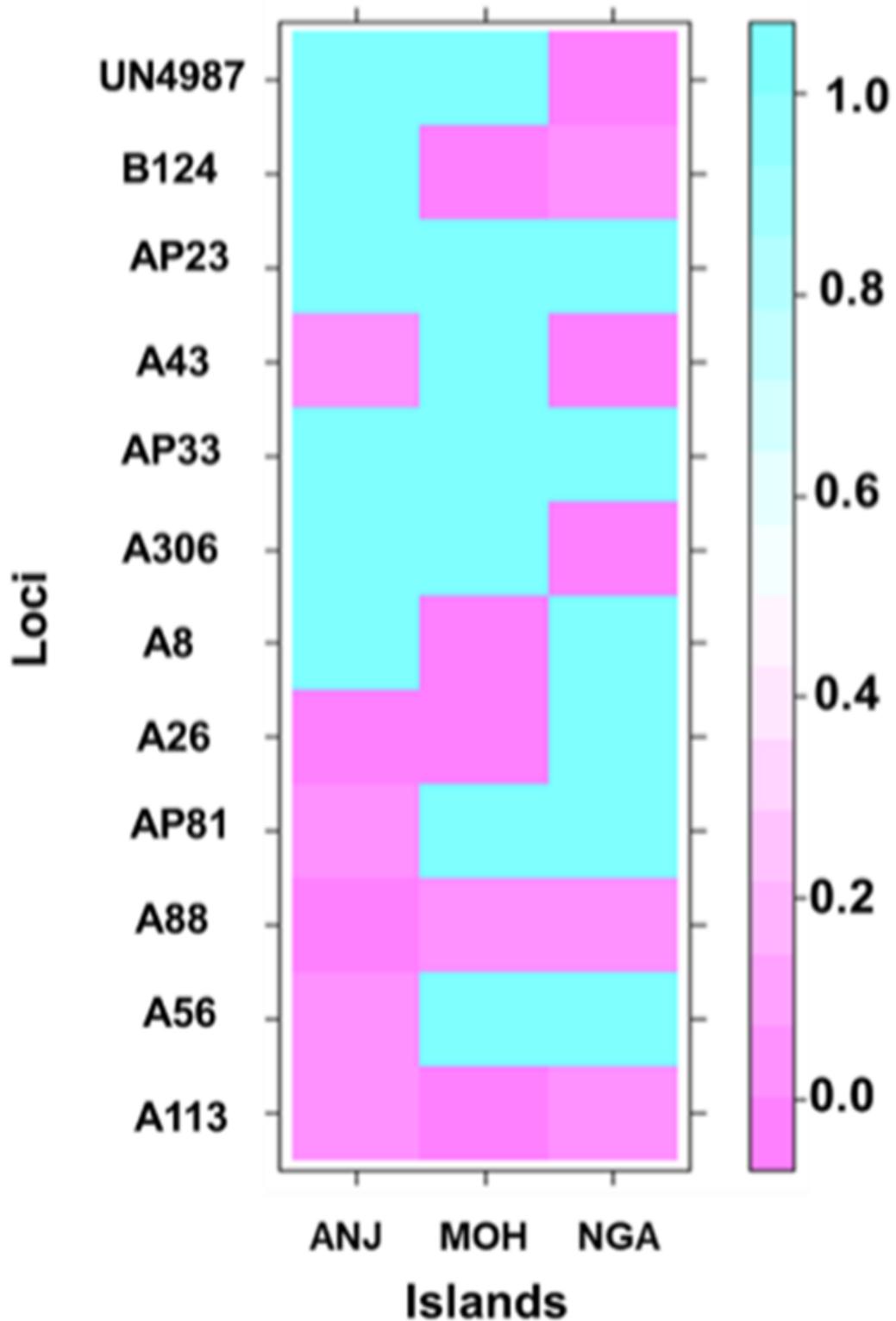


**Figure 4.7:** Allele distribution at locus AP23 in Anjouan (A), Moheli (B) and Ngazidja (C) showing the absence of allele 108 in Ngazidja.

**Table 4.4:** Table indicating various indices of diversity within the entire Comoros population and within individual islands.

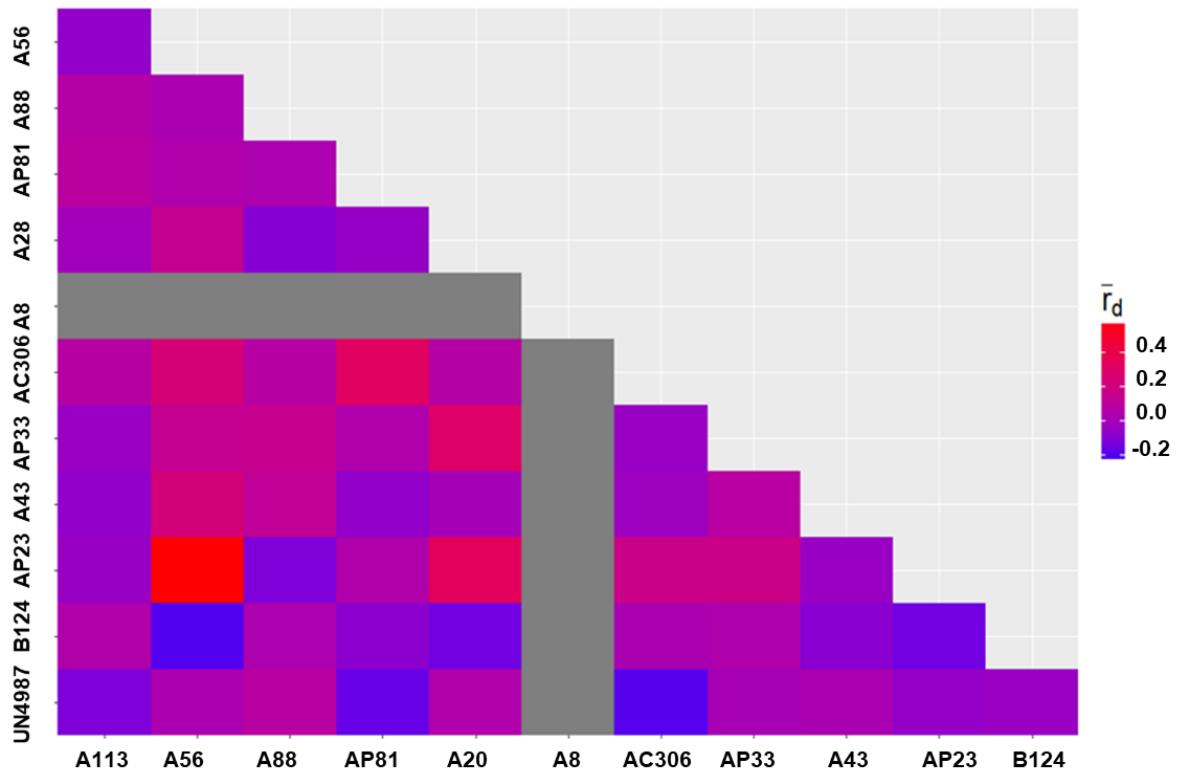
Locus	Entire Comoros Population					Ngazidja				Anjouan				Moheli			
	$N_a$	Simpson's diversity Index	$H_{exp}$	Eve nness	$G_{ST}$	$N_a$	Simpson 's diversi ty Index	$H_{exp}$	Even ness	$N_a$	Simpson 's divers ity Inde x	$H_{exp}$	Eve nness	$N_a$	Simpso n's dive rsity In dex	$H_{exp}$	Eve nness
A113	3	0.63	0.64	0.93	0.14	3	0.62	0.64	0.91	3	0.61	0.63	0.89	3	0.55	0.57	0.88
A56	4	0.64	0.64	0.90	0.68	4	0.67	0.69	0.91	2	0.36	0.37	0.77	2	0.30	0.31	0.71
A88	6	0.69	0.70	0.80	0.28	3	0.5	0.51	0.74	6	0.78	0.81	0.86	5	0.67	0.69	0.81
AP81	8	0.78	0.79	0.76	0.19	7	0.71	0.73	0.70	6	0.76	0.79	0.80	5	0.77	0.80	0.93
A28	5	0.66	0.67	0.76	0.27	0	1	0.00	0.00	3	0.54	0.56	0.83	5	0.67	0.70	0.78
A8	3	0.32	0.32	0.60	0.36	0	1	0	0.00	1	0.00	0.00	0.00	3	0.49	0.50	0.76
AC306	10	0.84	0.85	0.88	0.76	6	0.67	0.69	0.74	5	0.67	0.70	0.75	4	0.71	0.73	0.91
AP33	12	0.77	0.78	0.61	0.76	5	0.69	0.71	0.82	5	0.61	0.64	0.67	4	0.36	0.38	0.56
A43	5	0.68	0.69	0.81	0.53	4	0.51	0.53	0.72	2	0.45	0.47	0.91	4	0.67	0.69	0.90
AP23	2	0.29	0.29	0.69	0.31	1	0	0	0	2	0.26	0.27	0.66	2	0.48	0.50	0.97
B124	8	0.80	0.81	0.84	0.40	6	0.75	0.77	0.82	5	0.72	0.75	0.86	5	0.74	0.76	0.85
UN498	5	0.74	0.74	0.84	0.19	5	0.70	0.72	0.81	5	0.78	0.81	0.94	5	0.62	0.64	0.68
7	Mean± SEM	5.92 ±0.05	0.65 ±0.05	0.66 ±0.03	0.79 ±0.06	0.41±0.068	3.67±0.05	0.65±0.08	0.50±0.10	0.60±0.53	0.55±0.08	0.57±0.07	0.75±0.34	3.92±0.04	0.59±0.04	0.61±0.03	0.81±0.03

$N_a$ , number of alleles;  $H_{exp}$ , expected unbiased heterozygosity;  $G_{ST}$ , Hendrick's standardised  $G_{ST}$  SEM, standard error of Mean

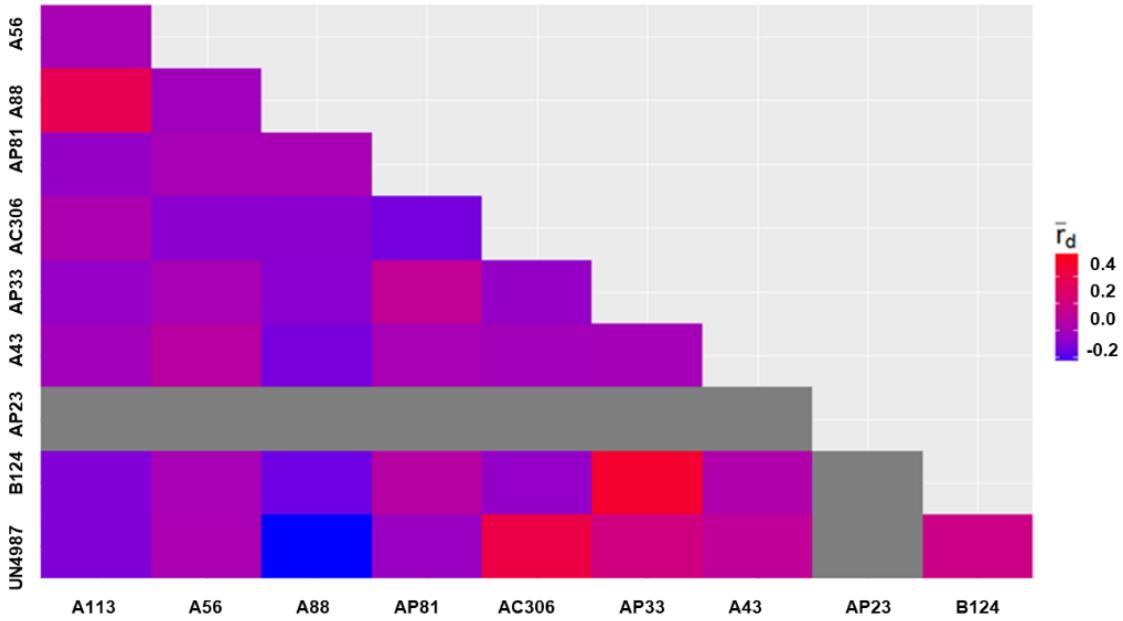


**Figure 4.8:** Heatmap showing deviations from the HWE. Loci in pink are those suspected of not being in HWE while all the loci in blue conform to the HWE.

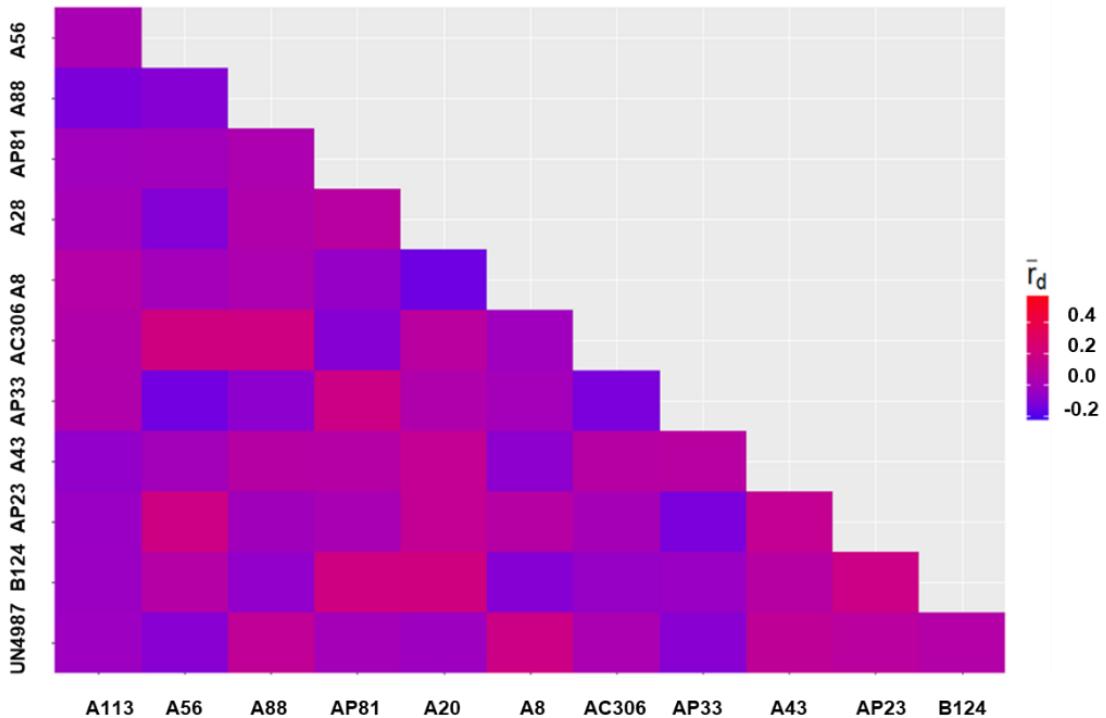
Most of the loci were in a linkage disequilibrium. However, markers AP23 and A56 were strongly linked in Anjouan honeybee populations (Figure 4.9). In Ngazidja, markers AP33 and B124; A113 and A88 and AC306 and UN4987 were strongly linkage (Figure 4.10). In Moheli, strong linkage was not detected in any of the loci (Figure 4.11).



**Figure 4.9:** Heatmap showing pairwise linkage disequilibrium pattern in honeybees collected in Anjouan. Blue indicates no linkage, red indicates strong linkage between markers whilst grey indicates missing data.



**Figure 4.10:** Heatmap showing Pairwise linkage disequilibrium pattern in honeybees collected in Ngazidja. Blue indicates no linkage, red indicates strong linkage between markers whilst grey is missing data.



**Figure 4.11** Heatmap showing Pairwise linkage disequilibrium pattern in honeybees collected in Moheli. Blue indicates no linkage while red indicates strong linkage between markers

#### **4.2.4.2 Population differentiation and population structure**

##### **4.2.4.2.1 Population differentiation and Analysis of Molecular Variance**

Using  $G_{ST}$  to infer entire population differentiation revealed a strong differentiation ( $G_{ST}=0.41(\pm 0.068)$ ) (Table 4.4) for the Comoros population. A113 ( $G_{ST}=0.14$ ) was the least differentiated whilst locus AP33 was strongly differentiated ( $G_{ST}=0.76$ ) (Table 4.4). Pairwise  $G_{ST}$  comparisons between islands showed strong differentiation between all the islands ( $G_{ST}>0.2$ ) with a much stronger differentiation occurring between Moheli and Ngazidja (0.52) (Table 4.5). Pairwise genetic distance based on Hedrick's  $G_{ST}$  showed that Ngazidja and Anjouan were much closer than Ngazidja and Moheli while was almost equidistant from both Ngazidja and Anjouan (Table 4.5). AMOVA test revealed significant variations within and between samples as well as between populations ( $p = 0.001$ ) (Appendix VI).

**Table 4.5:** Pairwise Nei's  $G_{ST}$  Statistics revealing stronger differentiation between Ngazidja and Moheli as compared to the differentiation between Moheli and Anjouan or Anjouan and Ngazidja.

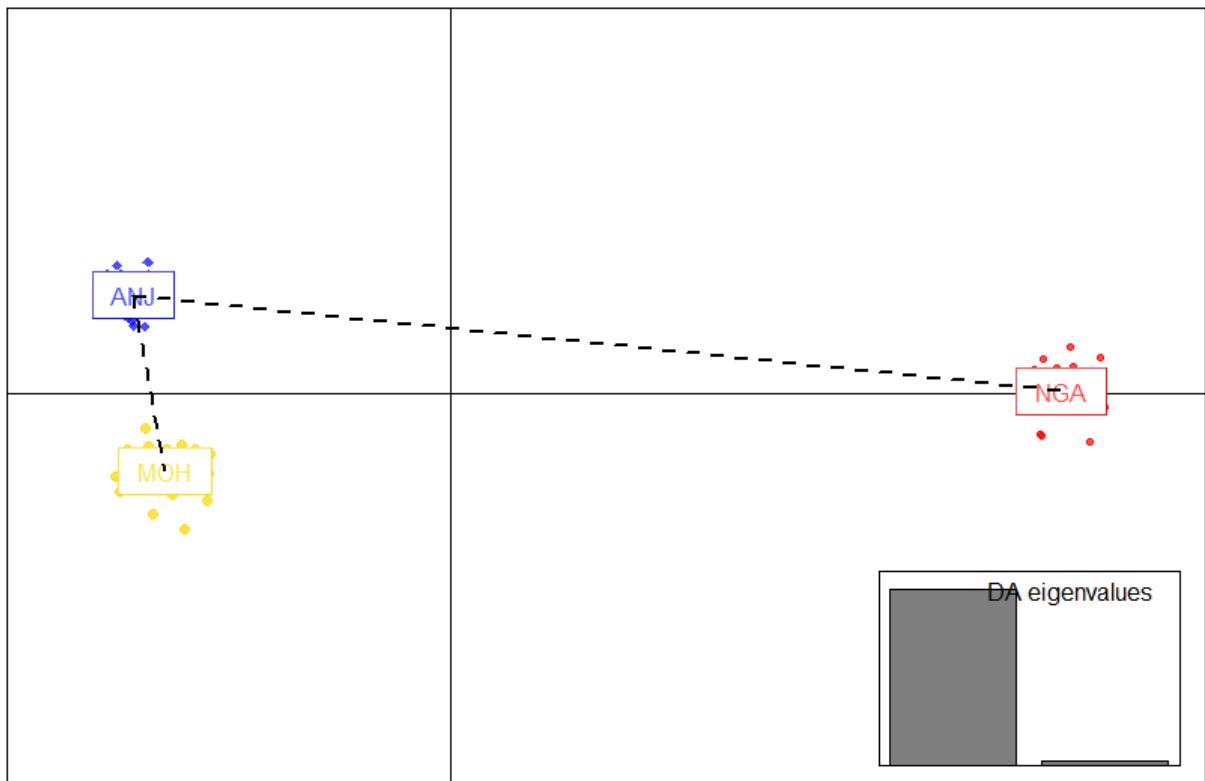
	ANJ	MOH
MOH	0.368	
NGA	0.380	0.525

Several loci in the different populations showed significant deviations from the HWE (Figure 4.8). (Appendix V).

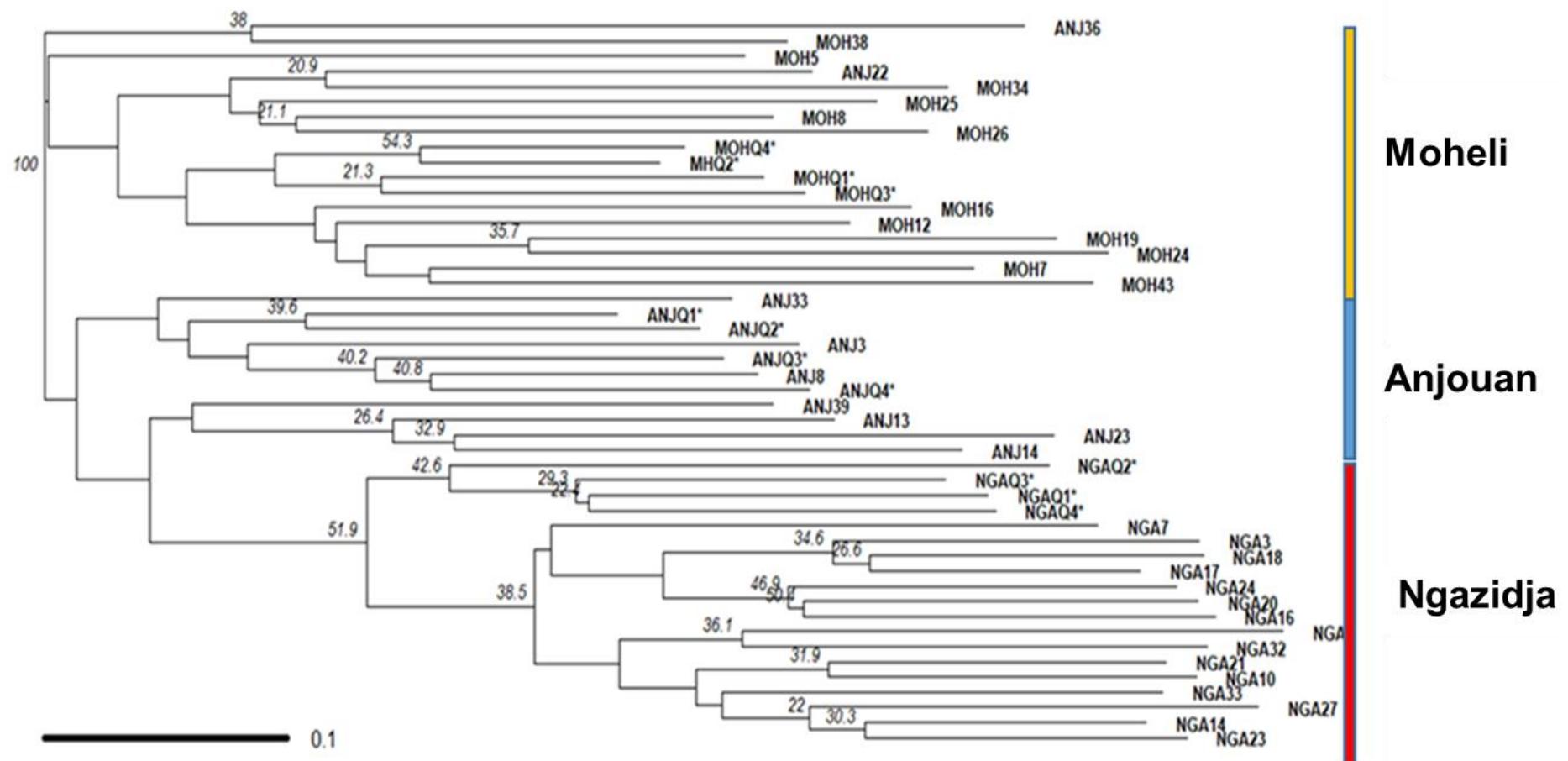
##### **4.2.4.2.2 Population structure**

A neighbor joining tree constructed revealed a population structured according to geography (Figure 4.13). The first level of separation strongly (bootstrap = 100%) separated samples from Anjouan and Ngazidja from Moheli. Samples from Anjouan were partially separated from the Ngazidja. In the DAPC analysis (Figure 4.12), PC 1 separated An-

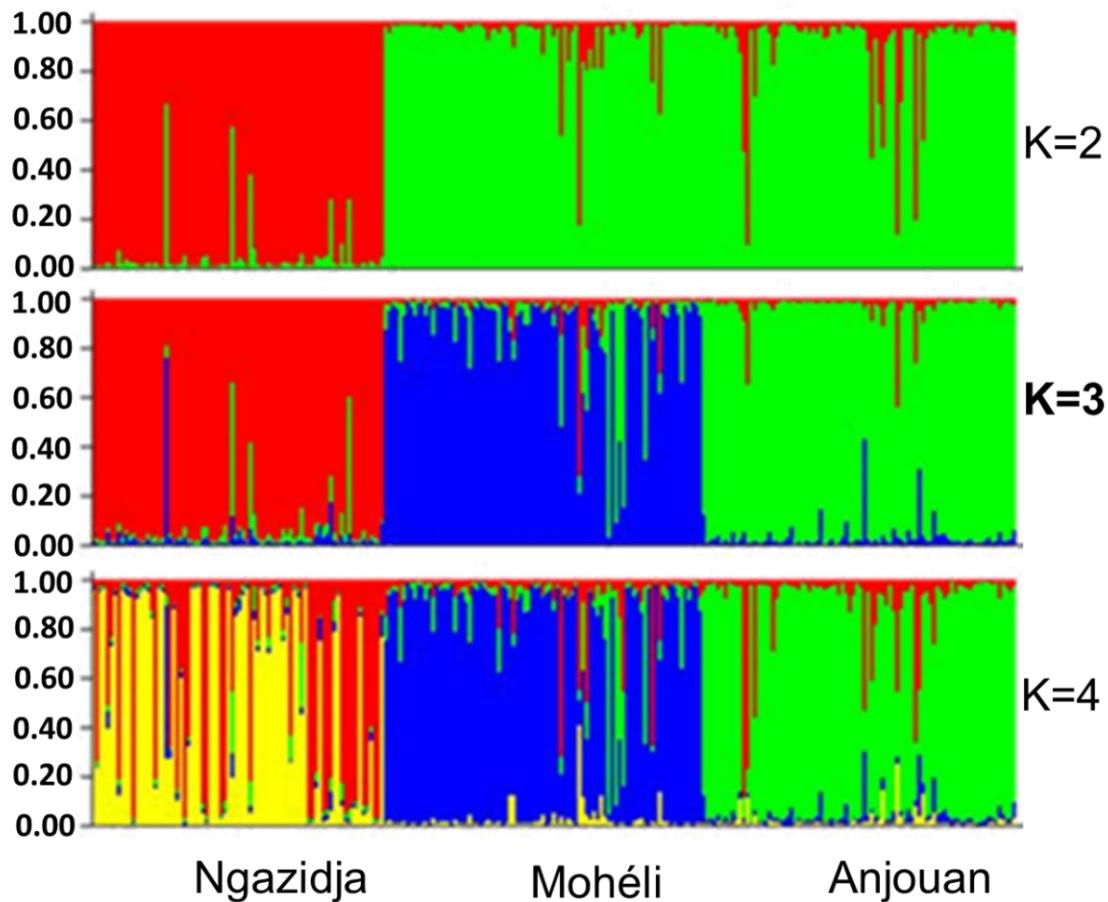
jouan and Moheli from the Ngazidja, whilst PC 2 separated Anjouan and Moheli populations. The most significant STRUCTURE results ( $K = 3$ ) also clustered the samples in accordance to the islands of origin (Figure 4.14). Notably, Ngazidja population with 51.9% bootstrap support divided into two sub-populations in the neighbor joining tree. This separation was also seen in the PC 2 of DAPC analysis (Figure 4.12) and STRUCTURE analysis at  $K=4$  (Figure 4.14).



**Figure 4.12:** Discriminant Analysis of Principal Component (DAPC) image showing how samples from the three islands in Comoros interact in space. ANJ=Anjouan; NGA=Ngazidja; MOH=Moheli.



**Figure 4.13:** A neighbor-joining tree calculated based on 12 microsatellite loci and 1000 bootstraps using Provesti's distance showing a structured population in Comoros islands.



**Figure 4.14:** Population structure of honeybees in Comoros islands. In the image, the best population structure is achieved at  $K=3$  which indicates structuring according to the island of origin namely, Ngazidja (red), Moheli (blue) and Anjouan (green).  $K=2$  clusters both Moheli and together whilst  $K=4$  shows partial separation in Ngazidja (red and yellow) but does not achieve significant separation in Anjouan.

## CHAPTER FIVE

### DISCUSSION, CONCLUSIONS AND RECOMMENDATIONS

#### **5.1 Discussion**

##### **5.1.1 Mitochondrial DNA haplotypes in Comoros Islands**

Mitochondrial DNA sequence analysis showed that the Comoros Islands are colonized by the A lineage of honeybees that are 100 % similar to *A. mellifera unicolor* native to Madagascar (Rasolofoarivao *et al.*, 2015). This results support Techer *et al.* (2017a, 2017b) findings which reported the exclusive presence of A1 lineage in most of the islands to the SWIO except in the Rodrigues islands where the European C lineage was predominant (Techer *et al.*, 2017a). Contrary to Techer *et al.* (2017a, 2017b), these study revealed existence of a honeybee subspecies (L haplotype(s)) that closely clustered alongside the *A. koschevnikovi*, a honeybee subspecies native to Southeast Asia. Close relationship between L1 and L2 with *A. koschevnikovi* needs an explanation.

Human movement from Southeast Asia to Madagascar and Comoros is well documented with Archaeobotanical evidence showing the introduction of Asian crops to the islands during the period (Boivin *et al.*, 2016). It is likely that Southeast Asia honeybees were also introduced in the Comoros islands during that time and have since colonized the island to the present time. The A1 haplotype is distinguishable from both L1 and L2 based on the length of the COI-COII amplified fragment (Figure 4.1). Whereas the A1 fragment is 600 bp the other two are 380 bp in size.

Recombination, the principle mechanism underlying nuclear reorganization is a rare phenomenon in the mtDNA genomes. This study shows that L2 arose from a rare mtDNA recombination between A1 and L1 (Figure 4.2). Cases of recombining mtDNA genomes are well documented in plants, fungi (Barroso and Labarère, 1997) and animals (Ladoukakis and Zouros, 2001; Ciborowski *et al.*, 2007; Good *et al.*, 2008). Heteroplasmy, occurrence of more than one mtDNA or plastid DNA within a cell, has been suggested by previous studies to play a central role in ensuring recombination in the mtDNA genome (Baptista-Ferreira, Economou and Casselton, 1983; Barroso and Labarère, 1997; Saville, Kohli and Anderson, 1998). Though the sperm tail, which harbors the mitochondria, in many animals is excluded from penetrating the egg, there exists evidence of a breach to this barrier in some animals, mostly insects. This result in the incorporation of the male mitochondria into the egg (Hildereth and Lucchesi, 1963). In addition, cases of more than one sperm penetrating the egg among insects are well documented, a scenario known as polyspermy.

Honeybees are known to be characterized by both mechanisms i.e. complete penetration and polyspermy ultimately leading to a more paternal contribution of mtDNA (Meusel and Moritz, 1993) which could then lead to heteroplasmy. Furthermore, the possible leakage of considerable amounts of paternal mtDNA into the egg among the honeybees has been shown (Meusel and Moritz, 1993). Direct evidence of mitochondrial DNA recombination in fungi, plants and protists (Gray, 1989) exists. Besides, fusion of the mtDNA has been shown in *Drosophila* (Yaffe, 1999), Crickets (Rand and Harrison, 1986), scallops(Snyder *et al.*, 1987) as well as sturgeons (Buroker *et al.*, 1990) with non-

homologous recombination being put forward as the probable recombination mechanism involved.

The study also showed, for the first time, the coexistence of two honeybee haplotypes without preferential replacement of one haplotype as reported in different studies (Hall and Muralidharan, 1989; Wragg *et al.*, 2018). The African honeybee *A. mellifera scutellata* was introduced in South America in the early 1950s and has since spread to colonize most countries in the South America slowly replacing the European *A. mellifera mellifera* haplotype. The same scenario was recently observed in La Reunion where European honeybee species of the C lineage were introduced 30 years ago. In the study, the A lineage haplotype was shown to have replaced the C lineage (Wragg *et al.*, 2018).

The coexistence of two haplotypes in Comoros islands suggest that they both exist as part of mitonuclear coadaptation complexes (Hill, 2015) necessary for survival in their habitat. Mitonuclear coadaptation refers to cases where specific nuclear genes are adapted to survive alongside specific mtDNA genes and this necessitates the need for cotransmission of genes involved in the complex formation (Rand, Haney and Fry, 2004; Rogell *et al.*, 2014; Hill, 2015). Nuclear gene conveying key survival adaptations could be linked to the different haplotypes thus the need for their coexistence. Maintenance of two haplotypes in a near equilibrium as observed in Anjouan (4.) could arise from a negative frequency-dependend selection (NFDS) mechanism (Kazancioğlu and Arnqvist, 2014). In this type of selection mechanism, the mitotype with the lowest population frequency increases whilst the mitotype with the highest population frequency decreases until a bal-

ance is achieved (Hill, 2015). An experimental study involving seed beetles (Kazancioğlu and Arnqvist, 2014) as well as few observation in drosophila (MacRae and Anderson, 1988; Oliver *et al.*, 2005; Andrianov *et al.*, 2008) associates NFDS to the coexistence of two mitochondrial haplotypes in a near stable equilibrium.

### **5.1.2 Genetic diversity, population differentiation and population structure**

The Comoros honeybees exhibit high genetic diversity evidenced by high average expected heterozygosity ( $0.66 (\pm 0.03)$ ), alleles per locus ( $5.92 (\pm 0.05)$ ) and Simpson's diversity index ( $0.65 (\pm 0.05)$ ). The average allele per locus for individual populations ranging between  $3.67 (\pm 0.05)$  and  $3.92 (\pm 0.04)$  is much higher than in Madagascar, where an average of 2.47 and 3.18 alleles per locus within populations were reported (Rasolofoarivao *et al.*, 2015). However, the average alleles per locus for the entire population ( $5.92 (\pm 0.05)$ ) is below the average in the study in Madagascar (7.76) (Rasolofoarivao *et al.*, 2015).

High diversity within the Comoros honeybee populations could be the result of high queen mating frequency (27 to 38 drones). Increased polyandry mating results in increased offspring heterozygosity (Taylor, Price and Wedell, 2014). The high mating frequency increases the number of reproductively successful males that give rise to increased effective population size (Trontti *et al.*, 2007). On average, honeybees mate with up to 16 drones (Schlüns *et al.*, 2005). The high queen mating frequency recorded in Comoros could arise from the need to reduce the chances of inbreeding and its associated cost. Generally, Islands are inhabited by small populations and given the division of labor nature of Hymenopterans, the census population does not translate to the effective popu-

lation. In order to maintain the small population, it is important to keep the inbreeding rate as low as possible and a higher queen mating frequency would be one check mechanism (Nomura, 2018). The haplodiploid sex determination mechanism in social insects renders inbreeding costly as homozygosity at the sex determining locus results in sterile diploid males which can neither contribute to the workforce nor reproduction (Trontti *et al.*, 2007).

Higher average alleles per locus in Moheli could explain the high diversity ( $H_{exp} = 0.61 (\pm 0.03)$ ) in the population as compared to Anjouan ( $H_{exp} = 0.57 (\pm 0.07)$ ) and Ngazidja ( $H_{exp} = 0.50 (\pm 0.10)$ ) both of which have low average alleles per locus. One possible explanation for the high variability is the small human population in Moheli (50,854) and slow urbanization (Andjib, 2015) which exert minimal fragmentation on the honeybee natural habitats. Anthropogenic disturbance resulting in fragmentation of natural habitats is associated with reduction in species abundance and diversity (Hung, Ascher and Holway, 2017). During sampling, wild honeybee colonies were found nesting on house roofs and walls, old vehicle and around human settlement which suggest the lack of appropriate nesting sites following human interference.

Broadly, high variability at the microsatellite loci corroborates previous studies which showed high polymorphism levels at different microsatellite loci among the African honeybees (Estoup *et al.*, 1995; Franck *et al.*, 1998). The high polymorphism can be explained based on the suggestion by Frank *et al.* (1998), who linked high levels of polymorphism among the African honeybees to their pronounced swarming tendency and

migratory behavior. In addition, it could be as a result of climate change (Frank *et al.*, 2001) or the presence of a large effective population size that gives room for the maintenance of more alleles in a population (Estoup *et al.*, 1995). Large effective population could result from the high queen mating frequency. The high genetic diversity within populations, suggests that the population is panmictic characterised by random mating within breeding populations. Increased polymorphism among the microsatellite loci are a result of random mating (Loucif-Ayad *et al.*, 2015). High nuclear diversity coupled to restricted gene flow within population was reported among honeybees of Seychelles archipelagos whose geographical organization is almost similar to the Comoros Archipelagos (Techer *et al.*, 2015). Findings in honeybees from Comoros thus corroborates Techer *et al.* (2015) suggestions that high variations might arise due to existence of a permanent geographic barrier-in this case the ocean water.

The presence of private alleles 216 and 218 at the A113 locus in Anjouan and Moheli is of interest to note (Figure 4.6). Though present, their limited frequency cannot be used to give a conclusive argument regarding their occurrence. The complete absence of allele 108 and dominance of allele 104 at the AP23 locus in Ngazidja needs an explanation. Genetic drift which is usually associated with random fluctuations in the number of alleles at specific loci is the most likely explanation. In genetic drift, alleles either decrease or increase over time leading to complete loss and fixation of a specific alleles (Sarma *et al.*, 2019). It is also expected that Ngazidja with the lowest genetic diversity is likely to experience a genetic drift among the infrequently occurring alleles as compared on the other islands. The absence of allele 104 in Ngazidja is an indicator of restricted gene flow be-

tween either Anjouan or Moheli and Ngazidja. Significant deviation from the HWE within individual populations suggests that the population is undergoing evolution and that there is inbreeding within the islands.

The geographic structuring, strong pairwise genetic differentiation as well as the significant variance between islands based on AMOVA suggests a restricted gene flow between islands. Isolation by distance of over 50 km is the most possible explanation since honeybees cannot fly over a large water mass separated by such long distances (Techer *et al.*, 2016). Poorly developed beekeeping practices in the islands where most colonies exist as wild colonies deter human aided movement of bees between islands.

The study results suggest that hybridization gene flow occurs in the nuclear genome within islands since no sub-structuring is observed within islands. Sub-structuring of the Ngazidja is non-significant based on STRUCTURE analysis was shown to be non-significant and cannot be used to describe any population structure in Ngazidja. Although, the same sub-structuring was detected in PC 2 of DAPC analysis and neighbor joining tree, the two analyses were based on inferred queen genotypes which have been shown to have negative impact when used to infer population structure (Lepais *et al.*, 2010). Besides, the STRUCTURE result was based on true drone genotypes thus more powerful. These findings are similar to what is reported in Seychelles archipelagos where the population is structured according to the islands but no structure is noted within individual islands (Techer *et al.*, 2015).

## 5.2 Conclusions

This work has shown that fragmentation to a certain scale can result in population structure as the nuclear genome of the honeybees in Comoros is structured according to the islands. Through the work, it is clear that honeybees have the ability to undergo recombination and resulting in unique mitotypes. Through this work it is shown, for the first time, the possibility of existence of a new haplotype of honeybees in the Comoros islands which could have separated from the other cavity nesting bees before *A. mellifera*.

## 5.3 Recommendations

Since the current study has shown the existence of unique mitotypes Comoros islands, there is need to ensure that the honeybee are adequately conserved in order to retain the endemic species.

## 5.4 Future studies

Future studies need to investigate the presence of honeybee pathogens in the Comoros islands and compare he rate of infection, and pathogen load among the described haplotypes. Future studies should also investigate the mechanism that govern mtDNA recombination in honeybees. Lastly, there is need to carry out a phenotypic study on the Comoros honeybees and correlate it to the genotypic data presented in this study.

## REFERENCES

- Abdul-Muneer, P. M. (2014)** ‘Application of Microsatellite Markers in Conservation Genetics and Fisheries Management: Recent Advances in Population Structure Analysis and Conservation Strategies’, *Genetics Research International*. Hindawi Publishing Corporation, 2014.: 1–11. doi: 10.1155/2014/691759.
- Alburaki, M., Bertrand, B., Legout, H., Moulin, S., Alburaki, A., Sheppard, W. S. and Garnery, L. (2013)** ‘A fifth major genetic group among honeybees revealed in Syria’, *BMC Genetics*, 14. doi: 10.1186/1471-2156-14-117.
- Andjib, S. A. (2015)** Vice President in charge of the Ministry of Territory Development, of Infrastructures, Urbanization and Housing. Available at: <http://habitat3.org/wp-content/uploads/National-Reports-Comoros-English.pdf>.
- Andrianov, B. V., Sorokina, S. Y., Mugue, N. S., Reznik, N. L. and Mitrofanov, V. G. (2008)** ‘Dynamics of mitochondrial polymorphism in a natural population of *Drosophila littoralis*’, *Russian Journal of Genetics*, 44(2), : 159–164. doi: 10.1007/s11177-008-2006-2.
- Arias, M. C. and Sheppard, W. S. (1996)** ‘Molecular Phylogenetics of Honey Bee Subspecies (*Apis mellifera* L.) Inferred from Mitochondrial DNA Sequence’, *Molecular Phylogenetics and Evolution*, 5(3), : 557–566. doi: 10.1006/mpev.1996.0050.
- Baptista-Ferreira, J. L. C., Economou, A. and Casselton, L. A. (1983)** ‘Mitochondrial genetics of *Coprinus*: recombination of mitochondrial genomes’, *Current genetics*, 7(5), : 405–407. doi: 10.1038/414745a.
- Barroso, G. and Labarère, J. (1997)** ‘Genetic evidence for nonrandom sorting of mitochondria in the basidiomycete *Agrocybe aegerita*’, *Applied and Environmental Microbiology*, 63(12), : 4686–4691. doi: 10.1534/genetics.112.140483.
- Baudry, E., Solignac, M., Garnery, L., Gries, M., Cornuet, J. and Koeniger, N. (1998)** ‘Relatedness among honeybees (*Apis mellifera*) of a drone congregation’, *Proceedings of the Royal Society B: Biological Sciences*, 265(1409), : 2009–2014. doi: 10.1098/rspb.1998.0533.
- Boivin, N. L., Lucas, L., Fuller, D. Q., Radimilahy, C., Walshaw, S., Pawlowicz, M., Shipton, C., Helm, R., et al. (2016)** ‘Ancient crops provide first archaeological signature of the westward Austronesian expansion’, *Proceedings of the National Academy of Sciences*, 113(24),: 6635–6640. doi: 10.1073/pnas.1522714113.
- Brown, A. H., Feldman, M. W. and Nevo, E. (1980)** ‘Multilocus Structure of Natural Populations of *Hordeum spontaneum*.’, *Genetics*, 96(2), : 523–536.
- Buroker, N. E., Brown, J. R., Gilbert, T. A., O’Hara, P. J., Beckenbach, A. T., Thomas, W. K. and Smith, M. J. (1990)** ‘Length heteroplasmy of sturgeon mitochondrial DNA: An illegitimate elongation model’, *Genetics*, 124(1), : 157–163.

- Ciborowski, K. L., Consuegra, S., García de Leániz, C., Beaumont, M. A., Wang, J. and Jordan, W. C. (2007)** ‘Rare and fleeting: an example of interspecific recombination in animal mitochondrial DNA’, *Biology Letters*, 3(5), : 554–557. doi: 10.1098/rsbl.2007.0290.
- Cornuet and Garnery (1991)** ‘Mitochondrial DNA variability in honeybees and its phylogeographic implications’, *Apidologie*, 22(6), : 627–642. doi: 10.1051/apido:19910606.
- Cornuet, J. and Garnery, L. (1991)** ‘Putative Origin and Function of the Intergenic Region Between COI and COII of *Apis mellifera* L. Mitochondrial DNA’.
- Desai, S. D. and Currie, R. W. (2015)** ‘Genetic diversity within honey bee colonies affects pathogen load and relative virus levels in honey bees, *Apis mellifera* L’, *Behavioral Ecology and Sociobiology*, 69(9), pp. 1527–1541. doi: 10.1007/s00265-015-1965-2.
- Edgar, R. C. (2004)** ‘MUSCLE: Multiple sequence alignment with high accuracy and high throughput’, *Nucleic Acids Research*, 32(5), : 1792–1797. doi: 10.1093/nar/gkh340.
- Eilers, E. J., Kremen, C., Greenleaf, S. S., Garber, A. K. and Klein, A. M. (2011)** ‘Contribution of pollinator-mediated crops to nutrients in the human food supply’, *PLoS ONE*, 6(6). doi: 10.1371/journal.pone.0021363.
- Estoup, A., Presa, P., Krieg, F., Vaiman, D. and Guyomard, R. (1993)** ‘(CT)<sub>n</sub> and (GT)<sub>n</sub> microsatellites: A new class of genetic markers for *Salmo trutta* L.(brown trout)’, *Heredity*, 71(5), : 488–496. doi: 10.1038/hdy.1993.167.
- Estoup, A., Garnery, L., Solignac, M. and Cornuet, J. M. (1995)** ‘Microsatellite variation in honey bee (*Apis mellifera* L.) populations: Hierarchical genetic structure and test of the infinite allele and stepwise mutation models’, *Genetics*, 140(2), : 679–695. doi: 10.1089/hum.2006.17.1165.
- Estoup, A., Solignac, M. and Cornuet, J.-M. (1994)** ‘Precise Assessment of the Number of Patrilines and of Genetic Relatedness in Honeybee Colonies’, *Proceedings of the Royal Society B: Biological Sciences*, 258(1351), : 1–7. doi: 10.1098/rspb.1994.0133.
- Evans, J. D., Schwarz, R. S., Chen, Y. P., Budge, G., Cornman, R. S., De la Rua, P., de Miranda, J. R., Foret, S., et al. (2013)** ‘Standard methods for molecular research in *Apis mellifera*’, *Journal of Apicultural Research*, 52(4), : 1–54. doi: 10.3896/IBRA.1.52.4.11.
- Excoffier, L., Smouse, P. E. and Quattro, J. M. (1992)** ‘Analysis of molecular variance inferred from metric distances among DNA haplotypes: application to human mitochondrial DNA restriction data.’, *Genetics*, 131(2), : 479–91. doi: 10.3354/meps198283.
- Franck, P., Garnery, L., Solignac, M. and Cornuet, J.-M. (1998)** ‘The origin of West European subspecies of honeybees (*Apis mellifera*): New insights from

- microsatellite and mitochondrial data', *Evolution*, 52(4), : 119–1134.
- Franck, P., Garnery, L., Solignac, M. and Cornuet, J. (2000)** 'Molecular confirmation of a fourth lineage in honeybees from the Near East', *Apidologie*, 31, : 167–180. doi: 10.1051/apido:2000114.
- Franck, P., Garnery, L., Loiseau, A., Oldroyd, B. P., Hepburn, H. R., Solignac, M. and Cornuet, J. M. (2001)** 'Genetic diversity of the honeybee in Africa: Microsatellite and mitochondrial data', *Heredity*, 86(4), : 420–430. doi: 10.1046/j.1365-2540.2001.00842.x.
- Fuchs, S. and SCHADE, V. (1994)** 'Lower Performance in Honeybee Colonies of Uniform Paternity', *Apidologie, Springer Verlag*, 25(2), : 155–168. doi: 10.1051/apido:19940204.
- Gallai, N., Salles, J. M., Settele, J. and Vaissière, B. E. (2009)** 'Economic valuation of the vulnerability of world agriculture confronted with pollinator decline', *Ecological Economics*. Elsevier B.V., 68(3), pp. 810–821. doi: 10.1016/j.ecolecon.2008.06.014.
- Garnery, L., Solignac, M., Celebrano, G. and Cornuet, J. (1993)** 'A simple test using restricted PCR-amplified mitochondrial DNA to study the genetic structure of *Apis mellifera* L.', (18).
- Garnery, L., Cornuet, J.-M. and Solignac, M.** (1992) 'Evolutionary history of the honey bee *Apis mellifera* inferred from mitochondrial DNA analysis', *Molecular Ecology*, 1(3), : 145–154. doi: 10.1111/j.1365-294X.1992.tb00170.x.
- Good, J. M., Hird, S., Reid, N., Demboski, J. R., Steppan, S. J., Martin-Nims, T. R. and Sullivan, J. (2008)** 'Ancient hybridization and mitochondrial capture between two species of chipmunks', *Molecular Ecology*, 17(5), : 1313–1327. doi: 10.1111/j.1365-294X.2007.03640.x.
- Gray, M. W. (1989)** 'Origin and Evolution of Mitochondrial DNA', *Annual Reviews Cell Biology*, : 25–50.
- Grünwald, N. J., Goodwin, S. B., Milgroom, M. G. and Fry, W. E. (2003)** 'Analysis of Genotypic Diversity Data for Populations of Microorganisms', *Phytopathology*, 93(6), : 738–746. doi: 10.1094/PHYTO.2003.93.6.738.
- Hall, H. G. and Muralidharan, K. (1989)** 'Evidence from mitochondrial DNA that African honey bees spread as continuous maternal lineages', *Nature*, 339(6221), : 211–213. doi: 10.1038/339211a0.
- Han, F., Wallberg, A. and Webster, M. T. (2012)** 'From where did the western honeybee (*Apis mellifera*) originate?', *Ecology and Evolution*, 2(8), pp. 1949–1957. doi: 10.1002/ece3.312.
- Hedrick, P. W. (2005)** 'A standardized genetic differentiation measure.', *Evolution; international journal of organic evolution*, 59(8), : 1633–1638. doi: Doi 10.1554/05-076.1.
- Hildereth, P. E. and Lucchesi, J. C. (1963)** 'Fertilization in *Drosophila* I. Evidence for

- the Regular Occurrence of Monospermy', *Developmental Biology*, 6, : 262–278.
- Hill, G. E. (2015)** 'Mitonuclear Ecology', *Molecular Biology and Evolution*, 32(8), :1917–1927. doi: 10.1093/molbev/msv104.
- Hung, K.-L. J., Ascher, J. S. and Holway, D. A. (2017)** 'Urbanization-induced habitat fragmentation erodes multiple components of temporal diversity in a Southern California native bee assemblage', *PLOS ONE*. Edited by N. E. Raine, 12(8), p. e0184136. doi: 10.1371/journal.pone.0184136.
- Johnson, R. M., Ellis, M. D., Mullin, C. A. and Frazier, M. (2010)** 'Pesticides and honey bee toxicity – USA', *Apidologie*, 41(3), : 312–331. doi: 10.1051/apido/2010018.
- Jombart, T., Devillard, S. and Balloux, F. (2010)** 'Discriminant analysis of principal components: a new method for the analysis of genetically structured populations', *BMC Genetics*, 11(1), p. 94. doi: 10.1186/1471-2156-11-94.
- Jones, O. R. and Wang, J. (2010)** 'COLONY: A program for parentage and sibship inference from multilocus genotype data', *Molecular Ecology Resources*, 10(3), : 551–555. doi: 10.1111/j.1755-0998.2009.02787.x.
- Kakumanu, M. L., Reeves, A. M., Anderson, T. D., Rodrigues, R. R. and Williams, M. A. (2016)** 'Honey bee gut microbiome is altered by in-hive pesticide exposures', *Frontiers in Microbiology*, 7(AUG), pp. 1–11. doi: 10.3389/fmicb.2016.01255.
- Kamvar, Z. N., Tabima, J. F. and Grünwald, N. J. (2014)** 'Poppr: an R package for genetic analysis of populations with clonal, partially clonal, and/or sexual reproduction', *PeerJ*, 2, p. e281. doi: 10.7717/peerj.281.
- Kazancioğlu, E. and Arnqvist, G. (2014)** 'The maintenance of mitochondrial genetic variation by negative frequency-dependent selection', *Ecology Letters*. Edited by D. Ebert, 17(1), : 22–27. doi: 10.1111/ele.12195.
- Klein, A.-M., Vaissiere, B. E., Cane, J. H., Steffan-Dewenter, I., Cunningham, S. A., Kremen, C. and Tscharntke, T. (2007)** 'Importance of pollinators in changing landscapes for world crops', *Proceedings of the Royal Society B: Biological Sciences*, 274(1608), : 303–313. doi: 10.1098/rspb.2006.3721.
- Delaplane, K. S. andamp; Mayer, D. F. (2000)**. Crop Pollination by Bees. - New York, Oxon (CABI Publishing). ISBN 0-85199-448-2
- Kraus, F. B., Franck, P. and Vandame, R. (2007)** 'Asymmetric introgression of African genes in honeybee populations (*Apis mellifera* L.) in Central Mexico', *Heredity*, 99(2),: 233–240. doi: 10.1038/sj.hdy.6800988.
- Kumar, S., Stecher, G., Peterson, D. and Tamura, K. (2012)** 'MEGA-CC: Computing core of molecular evolutionary genetics analysis program for automated and iterative data analysis', *Bioinformatics*, 28(20), : 2685–2686. doi: 10.1093/bioinformatics/bts507.
- Ladoukakis, E. D. and Zouros, E. (2001)** 'Direct Evidence for Homologous

- Recombination in Mussel ( *Mytilus galloprovincialis* ) Mitochondrial DNA', *Molecular Biology and Evolution*, (May), : 1168–1175.
- Lepais, O., Darvill, B., O'Connor, S., Osborne, J. L., Sanderson, R. A., Cussans, J., Goffe, L. and Goulson, D. (2010)** 'Estimation of bumblebee queen dispersal distances using sibship reconstruction method', *Molecular Ecology*, 19(4), : 819–831. doi: 10.1111/j.1365-294X.2009.04500.x.
- Loucif-Ayad, W., Achou, M., Legout, H., Alburaki, M. and Garnery, L. (2015)** 'Genetic assessment of Algerian honeybee populations by microsatellite markers', *Apidologie*, 46(3), : 392–402. doi: 10.1007/s13592-014-0331-0.
- MacDonald, J. H. (2009)** 'Kruskal-Wallis Test', *Biological Handbook of Statistics*, (1), : 165–172. doi: 10.1002/9780470479216.corpsy0491.
- MacRae, A. F. and Anderson, W. W. (1988)** 'Evidence for non-neutrality of mitochondrial DNA haplotypes in *Drosophila pseudoobscura*', *Genetics*, 120(2), : 485–494.
- Magnus, R. ., Tripodi, A. . and Szalanski, A. . (2011)** 'Mitochondrial DNA diversity of honey bees, *Apis mellifera* L. (Hymenoptera: Apidae) from queen breeders in the United States', *Journal of Apicultural Science*, 55(1), : 37–47.
- Mattila, H. R. and Seeley, T. D. (2007)** 'Genetic diversity in honey bee colonies enhances productivity and fitness', *Science*, 317(5836), pp. 362–364. doi: 10.1126/science.1143046.
- McMenamin, A. J. and Genersch, E. (2015)** 'Honey bee colony losses and associated viruses', *Current Opinion in Insect Science*. Elsevier Inc, 8, : 121–129. doi: 10.1016/j.cois.2015.01.015.
- Meusel, M. S. and Moritz, R. F. A. (1993)** 'Transfer of paternal mitochondrial DNA during fertilization of honeybee (*Apis mellifera* L.) eggs', *Current Genetics*, 24(6), : 539–543. doi: 10.1007/BF00351719.
- Michener, C. D. (2000)** *The Bees of the World*. 2715 North Charles Street Baltimore, Maryland 21218-4363: The John Hopkins University Press. Available at: www.press.jhu.edu.
- Nei, M. (1972)** 'Genetic Distance between Populations', *The American Naturalist*, 106(949), : 283–292. doi: 10.1086/282771.
- Nei, M. (1978)** 'Estimation of average heterozygosity and genetic distance from a small number of individuals', *Genetics*, 89(3), : 583–590. doi: 10.3390/ijms15010277.
- Nei, M. (1987)** *Molecular Evolutionary Genetics*, Columbia University Press.
- Neumann, P., Moritz, R. F. A., Mautz, D. and Ökologie, M. (1999)** 'Using DNA microsatellites for maternity testing in honeybees ( *Apis mellifera* L .) To cite this version : HAL Id : hal-00891644 Using DNA microsatellites for maternity testing in honeybees ( *Apis mellifera* L .) a'.

- Nomura, T. (2018)** ‘Maximum avoidance of inbreeding in haplodiploid populations’, *Mathematical Biosciences*. Elsevier, 306 (7), : 49–55. doi: 10.1016/j.mbs.2018.10.006.
- Oldroyd, B. P., Rinderer, T. E. and Buco, S. M. (1992)** ‘Intra-colonial foraging specialism by honey bees (*Apis mellifera*) (Hymenoptera: Apidae)’, *Behavioral Ecology and Sociobiology*, 30, : 291–295. doi: 10.1007/BF00170594.
- Oliver, P., Balanya, J., Ramon, M., Picornell, A., Serra, L., Moya, A. and Castro, J. A. (2005)** ‘Population dynamics of the 2 major mitochondrial DNA haplotypes in experimental populations of *Drosophila subobscura*’, *Genome*, 48(6), : 1010–1018. doi: 10.1139/g05-077.
- Palmer, K. A. and Oldroyd, B. P. (2003)** ‘Evidence for intra-colonial genetic variance in resistance to American foulbrood of honey bees (*Apis mellifera*): Further support for the parasite/pathogen hypothesis for the evolution of polyandry’, *Naturwissenschaften*, 90(6), : 265–268. doi: 10.1007/s00114-003-0418-3.
- Péntek-Zakar, E., Oleksa, A., Borowik, T. and Kusza, S. (2015)** ‘Population structure of honey bees in the Carpathian Basin (Hungary) confirms introgression from surrounding subspecies’, *Ecology and Evolution*, 5(23), : 5456–5467. doi: 10.1002/ece3.1781.
- Pirk, C. W. W., Strauss, U., Yusuf, A. A., Démares, F. and Human, H. (2016)** ‘Honeybee health in Africa—a review’, *Apidologie*, 47(3), : 276–300. doi: 10.1007/s13592-015-0406-6.
- Potts, S. G., Biesmeijer, J. C., Kremen, C., Neumann, P., Schweiger, O. and Kunin, W. E. (2010)** ‘Global pollinator declines: trends, impacts and drivers’, *Trends in Ecology and Evolution*. Elsevier Ltd, 25(6), : 345–353. doi: 10.1016/j.tree.2010.01.007.
- Pritchard, J. K., Stephens, M. and Donnelly, P. (2000)** ‘Inference of population structure using multilocus genotype data’, *Genetics*, 155, : 945–959. doi: 10.1111/j.1471-8286.2007.01758.x.
- Quarles, W. (2008)** ‘Pesticides and honey bee colony collapse disorder.’, *Ipm practitioner*, 30(9/10), : 1–10.
- Rand, D. M., Haney, R. A. and Fry, A. J. (2004)** ‘Cytonuclear coevolution: The genomics of cooperation’, *Trends in Ecology and Evolution*, 19(12), : 645–653. doi: 10.1016/j.tree.2004.10.003.
- Rand, D. M. and Harrison, R. G. (1986)** ‘Mitochondrial DNA transmission genetics in crickets.’, *Genetics*, 114(3), : 955–970.
- Rasolofoarivao, H., Clémencet, J., Techer, M. A., Ravaomanarivo, L. H. R., Reynaud, B. and Delatte, H. (2015)** ‘Genetic diversity of the endemic honeybee: *Apis mellifera unicolor* (Hymenoptera: Apidae) in Madagascar’, *Apidologie*, 46(6), : 735–747. doi: 10.1007/s13592-015-0362-1.
- Rogell, B., Dean, R., Lemos, B. and Dowling, D. K. (2014)** ‘Mito-nuclear interactions

- as drivers of gene movement on and off the X-chromosome', *BMC Genomics*, 15(1), : 1–9. doi: 10.1186/1471-2164-15-330.
- Ruttner, F. (1966)** 'The Life and Flight Activity of Drones', *Bee World*, 47(3), : 93–100. doi: 10.1080/0005772X.1966.11097111.
- Ruttner, Friedrich (1988)** *Biogeography and Taxonomy of Honeybees*. 1st edn. Berlin: Springer-Verlag Berlin Heidelberg GmbH.
- Ruttner, F., Tassencourt, L. and Louveaux, J. (1978)** 'Biometrical-statistical analysis of the geographical variability of *Apis mellifera* L.', *Apidolodie*, 9(4), : 363–381. doi: 10.1051/apido:19780408.
- Sarma, H., Pradhan, S., Mattaparthi, V. S. K. and Kaushik, S. (2019)** 'Phylogenetic Analysis: Early Evolution of Life', in *Encyclopedia of Bioinformatics and Computational Biology*. Elsevier, : 938–952. doi: 10.1016/B978-0-12-809633-8.20171-4.
- Saville, B. J., Kohli, Y. and Anderson, J. B. (1998)** 'mtDNA recombination in a natural population', *Proceedings of the National Academy of Sciences*, 95(February), : 1331–1335.
- Schlüns, H., Moritz, R. F. A., Neumann, P., Kryger, P. and Koeniger, G. (2005)** 'Multiple nuptial flights, sperm transfer and the evolution of extreme polyandry in honeybee queens', *Animal Behaviour*, 70(1), : 125–131. doi: 10.1016/j.anbehav.2004.11.005.
- Schmid-Hempel, P. (1995)** 'Parasites and social insects', 26(3), : 255–271.
- Shaibi, T., Lattorff, H. M. G. and Moritz, R. F. A. (2008)** 'A microsatellite DNA toolkit for studying population structure in *Apis mellifera*', *Molecular Ecology Resources*, 8(5), : 1034–1036. doi: 10.1111/j.1755-0998.2008.02146.x.
- Sheppard, W. S. and Smith, D. R. (2000)** 'Identification of African-Derived Bees in the Americas: A Survey of Methods', *Annals of the Entomological Society of America*, 93(2), : 159–176. doi: 10.1603/0013-8746(2000)093[0159:IOADBI]2.0.CO;2.
- Sherman, P. W., Seeley, T. D. and Reeve, H. K. (1988)** 'Parasites , Pathogens , and Polyandry in Social Hymenoptera Author ( s ): Paul W . Sherman , Thomas D . Seeley and Hudson K . Reeve Published by : The University of Chicago Press for The American Society of Naturalists Stable URL : <http://www.jstor.org/st>', 131(4), : 602–610.
- Simpson, E. H. (1949)** 'Measurement of Diversity', *Nature*, : 688–688. doi: 10.1038/163688a0.
- Smith, J. M., Smith, N. H., O'Rourke, M. and Spratt, B. G. (1993)** 'How clonal are bacteria?', *Proceedings of the National Academy of Sciences*, 90(10), : 4384–4388. doi: 10.1073/pnas.90.10.4384.
- Snyder, M., Fraser, A. R., LaRoche, J., Gartner-Kepkay, K. E. and Zouros, E. (1987)** 'Atypical mitochondrial DNA from the deep-sea scallop *Placopecten*

- magellanicus', *Proceedings of the National Academy of Sciences*, 84(21), : 7595–7599. doi: 10.1073/pnas.84.21.7595.
- Solignac, M., Vautrin, D., Loiseau, A., Mougel, F., Baudry, E., Estoup, A., Garnery, L., Haberl, M. and Cornuet, J. M. (2003)** 'Five hundred and fifty microsatellite markers for the study of the honeybee (*Apis mellifera* L.) genome', *Molecular Ecology Notes*, 3(2), : 307–311. doi: 10.1046/j.1471-8286.2003.00436.x.
- Stoddart, J. A. and Taylor, J. F. (1988)** 'Genotypic diversity: Estimation and prediction in samples', *Genetics*, 118(4), : 705–711.
- Tarpy, D. R. (2003)** 'Genetic diversity within honeybee colonies prevents severe infections and promotes colony growth', *Proceedings of the Royal Society B: Biological Sciences*, 270(1510), : 99–103. doi: 10.1098/rspb.2002.2199.
- Tarpy, D. R., Vanengelsdorp, D. and Pettis, J. S. (2013)** 'Genetic diversity affects colony survivorship in commercial honey bee colonies', *Naturwissenschaften*, 100(8), : 723–728. doi: 10.1007/s00114-013-1065-y.
- Taylor, M. L., Price, T. A. R. and Wedell, N. (2014)** 'Polyandry in nature: A global analysis', *Trends in Ecology and Evolution*. Elsevier Ltd, 29(7), : 376–383. doi: 10.1016/j.tree.2014.04.005.
- Techer, M. A., Clémencet, J., Turpin, P., Volbert, N., Reynaud, B. and Delatte, H. (2015)** 'Genetic characterization of the honeybee (*Apis mellifera*) population of Rodrigues Island, based on microsatellite and mitochondrial DNA', *Apidologie*, 46(4), : 445–454. doi: 10.1007/s13592-014-0335-9.
- Techer, M. A., Clémencet, J., Simiand, C., Portlouis, G., Reynaud, B. and Delatte, H. (2016)** 'Genetic diversity of the honeybee (*Apis mellifera* L.) populations in the Seychelles archipelago', *Insect Conservation and Diversity*. Edited by S. R. Leather and D. Roubik, 9(1), : 13–26. doi: 10.1111/icad.12138.
- Techer, M. A., Clémencet, J., Simiand, C., Turpin, P., Garney, L., Reynaud, B. and Delatte, H. (2017a)** 'Genetic diversity and differentiation among insular honey bee populations in the southwest Indian Ocean likely reflect old geographical isolation and modern introductions', *PLoS ONE*, 12(12), : 1–26. doi: 10.1371/journal.pone.0189234.
- Techer, M. A., Clémencet, J., Simiand, C., Preaduth, S., Azali, H. A., Reynaud, B. and Hélène, D. (2017b)** 'Large-scale mitochondrial DNA analysis of native honey bee *Apis mellifera* populations reveals a new African subgroup private to the South West Indian Ocean islands', *BMC Genetics*. BMC Genetics, 18(1), : 1–21. doi: 10.1186/s12863-017-0520-8.
- Trontti, K., Thurin, N., Sundstrom, L. and Aron, S. (2007)** 'Mating for convenience or genetic diversity? Mating patterns in the polygynous ant *Plagiolepis pygmaea*', *Behavioral Ecology*, 18(2), : 298–303. doi: 10.1093/beheco/arl083.
- Vallet, A. M. and Coles, J. A. (1993)** 'The perception of small objects by the drone honeybee', *Journal of Comparative Physiology A: Sensory, Neural and Behavioral*

- Physiology*, 172(2), : 183–188. doi: 10.1007/BF00189395.
- VanEngelsdorp, D., Evans, J. D., Saegeffer, C., Mullin, C., Haubruege, E., Nguyen, B. K., Frazier, M., Frazier, J., et al. (2009)** ‘Colony collapse disorder: A descriptive study’, *PLoS ONE*, 4(8). doi: 10.1371/journal.pone.0006481.
- Wallberg, A., Han, F., Wellhagen, G., Dahle, B., Kawata, M., Haddad, N., Luz, Z., Simões, P., et al. (2014)** ‘A worldwide survey of genome sequence variation provides insight into the evolutionary history of the honeybee *Apis mellifera*’, *Nature Publishing Group*. Nature Publishing Group, 46(10), : 1081–1088. doi: 10.1038/ng.3077.
- Watanabe, M. E. (2008)** ‘Colony Collapse Disorder: Many Suspects, No Smoking Gun’, *BioScience*, 58(5),: 384. doi: 10.1641/B580503.
- Whitfield, C. W., Behura, S. K., Berlocher, S. H., Clark, A. G., Johnston, J. S., Sheppard, W. S., Smith, D. R. and Suarez, A. V (2006)** ‘Thrice Out of Africa : Ancient and Recent Expansions of the Honey Bee, *Apis mellifera*’, *Science*, 314(October), : 642–645. doi: 10.1126/science.1132772.
- Wilson, E. O. (1971)** ‘The Insect Societies’, in *The Insect Societies*, p. 548.
- Woyke, J. (1962)** ‘Natural and Artificial Insemination of Queen Honeybees’, *Bee World*, 43(1), : 21–25. doi: 10.1080/0005772X.1962.11096922.
- Wragg, D., Techer, M. A., Canale-Tabet, K., Basso, B., Bidanel, J.-P., Labarthe, E., Bouchez, O., Le Conte, Y., Clémencet, J., Delatte, H. and Vignal, A. (2018)** ‘Autosomal and Mitochondrial Adaptation Following Admixture: A Case Study on the Honeybees of Reunion Island’, *Genome Biology and Evolution*, 10(1), : 220–238. doi: 10.1093/gbe/evx247.
- Yaffe, M. P. (1999)** ‘The machinery of mitochondrial inheritance and behavior’, *Science*, : 1493–1497. doi: 10.1126/science.283.5407.1493.

## APPENDICES

**APPENDIX I:** Sampled colonies in the three main islands of Comoros. Mitochondrial DNA analysis was performed on all the sampled colonies whilst the colonies in with \* ( $N=12$ ) were used for microsatellite DNA analysis.

<b>Colony</b>	<b>Site Name</b>	<b>Latitude</b>	<b>Longitude</b>	<b>Type of colony</b>	<b>Altitude (m)</b>
Ngazidja 1	-	S11° 44.176'	E43° 14.884'	Wild	63
Ngazidja 2	-	S11° 46.442'	E43° 16.868'	Apiary	307
Ngazidja 3	Bweni	S11° 45.456'	E43° 15.567'	Wild	59
Ngazidja 4*	Iconi	S11° 44.823'	E43° 14.464'	Wild	14
Ngazidja 5	Iconi	S11° 49.063'	E43° 16.925'	Wild	41
Ngazidja 6	Singani	S11° 50.817'	E43° 19.103'	Wild	140
Ngazidja 7*	Tsinimoichongo	S11° 51.137'	E43° 28.027'	Apiary	111
Ngazidja 8	Panda	S11° 52.586'	E43° 21.979'	Wild	112
Ngazidja 9	Simbusa	S11° 53.138'	E43° 28.027'	Wild	538
Ngazidja 10	Nyuma Milima	S11° 51.201'	E43° 26.758'	Wild	669
Ngazidja 11	Tsini Mapanga	S11° 47.636'	E43° 26.233'	Apiary	63
Ngazidja 12	Usipvo	S11° 34.980'	E43° 16.087'	Wild	31
Ngazidja 13	Usipvo	S11° 35.157'	E43° 16.712'	Wild	155
Ngazidja 14	Kwambani	S11° 37.576'	E43° 21.212'	Apiary	503
Ngazidja 15*	Bweni	S11° 38.252'	E43° 21.656'	Wild	482
Ngazidja 16	Sada	S11° 38.301'	E43° 21.855'	Wild	406
Ngazidja 17	-	S11° 39.009'	E43° 22.150'	Wild	370
Ngazidja 18	Dzahani	S11° 39.554	E43° 17.044'	Traditional hive	369
Ngazidja 19	Ifundihe	S11° 30.851'	E43° 23.231'	Wild	255
Ngazidja 20	Babadyani	S11° 29.038'	E43° 23.340'	Wild	253
Ngazidja 21	Babadyani	S11° 29.160'	E43° 23.246'	Wild	268
Ngazidja 22	Mwadja	S11° 27.704'	E43° 23.362'	Wild	332
Ngazidja 23*	Iuvwani	S11° 24.150'	E43° 23.959'	Wild	19
Ngazidja 24	Trepezini	S11° 25.123'	E43° 23.423'	Wild	189
Ngazidja 25	Uziyo	S11° 24.310'	E43° 20.927'	Wild	325
Ngazidja 26	Fomboni	S11° 24.242'	E43° 22.752'	Wild	226

<b>Colony</b>	<b>Site Name</b>	<b>Latitude</b>	<b>Longitude</b>	<b>Type of colo-</b>	<b>Altitude (m)</b>
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				<b>ny</b>	
Moheli 1	Mshegi	S12° 17.795'	E43° 45.052'	Wild	93
Moheli 2	Mahurani	S12° 18.615'	E43° 46.525'	Wild	36
Moheli 3	Sephadar Pomboni	S12° 16.723'	E43° 44.144'	Wild	15
Moheli 4	Batse	S12° 16.021'	E43° 41.875'	Wild	17
Moheli 5*	Domoni	S12° 15.428'	E43° 38.755'	Wild	37
Moheli 6	Djitrone	S12° 16.130'	E43° 38.018'	Wild	67
Moheli 7	Pakouni	S12° 16.884'	E43° 37.759'	Wild	138
Moheli 8	Barakani	S12° 17.588'	E43° 38.244'	Wild	122
Moheli 9	Wallah	S12° 19.508'	E43° 39.741'	Wild	18
Moheli 10	Wallah	S12° 19.561'	E43° 39.878'	Wild	24
Moheli 11	Bandani	S12° 19.606'	E43° 40.145'	Wild	15
Moheli 12*	Dahoni	S12° 19.682'	E43° 40.084'	Wild	22
Moheli 13	Wallah	S12° 19.805'	E43° 40.204'	Wild	28
Moheli 14	Wallah	S12° 20.672'	E43° 40.071'	Wild	24
Moheli 15	Ndronoroni	S12° 20.491'	E43° 40.809'	Wild	116
Moheli 16	Bandani	S12° 21.319'	E43° 48.089'	Wild	221
Moheli 17	Mwahani	S12° 22.061'	E43° 46.851'	Wild	22
Moheli 18*	Badralaju	S12° 22.218'	E43° 51.449'	Wild	39
Moheli 19	Shisiwani	S12° 22.419'	E43° 52.119'	Wild	44
Moheli 20	Badralaju	S12° 22.159'	E43° 51.549'	Wild	83
Moheli 21	Mlabada	S12° 20.524'	E43° 48.668'	Wild	279
Moheli 22	Wanani	S12° 20.294'	E43° 47.082'	Wild	275
Moheli 23*	Siryziroudrani	S12° 20.351'	E43° 46.574'	Wild	304
Moheli 24	Singana	-	-	Wild	-
Moheli 25	Crde Mibani	S12° 20.241'	E43° 46.122'	Wild	277
Moheli 26	Dangoni	S12° 20.164'	E43° 46.890'	Wild	299
Moheli 27	Uhoni	S12° 20.066'	E43° 47.230'	Wild	275
Moheli 28	Barakani	-	-	Wild	-
Moheli 29	Oungoni	-	-	Wild	-
Moheli 30	Miremani	-	-	Wild	-
Moheli 31	Moja oume	-	-	Wild	-
Moheli 32	Mlabanda	-	-	Wild	-

Moheli 33	Ndremeyani	-	-	Wild	-
Moheli 34	Salemani	-	-	Wild	-
Moheli 35	Mirngoni	-	-	Wild	-
Moheli 36	Nimachioi	-	-	Wild	-
Moheli 37	Mrabao	-	-	Wild	-
Moheli 27	Bandasalmini	-	-	Wild	-
Moheli 38	Mirngoni	-	-	Wild	-
Moheli 40	Banahari	-	-	Wild	-
Moheli 41	Mtrouni	-	-	Wild	-
Moheli 42	Dargoube	-	-	Wild	-
Moheli 43	Nkagani	-	-	Wild	-

Colony	Site Name	Latitude	Longitude	Type of colony	Altitude (m)
Anjouan 1	Patsy	S12° 09.307'	E44° 26.080'	Wild	263
Anjouan 2	Cuvette/Ntsameni	S12° 12.033'	E44° 27.765'	Wild	439
Anjouan 3	Tsembenou/Cuvette	S12° 12.620'	E44° 27.830'	Wild	468
Anjouan 4	Dindri	S12° 12.963'	E44° 27.024'	Wild	566
Anjouan 5*	Kambalahari	S12° 13. 417'	E44° 29.475'	Wild	476
Anjouan 6	Jeje	S12° 13.587'	E44° 31.218'	Wild	64
Anjouan 7	Bambao Matsanga	S12° 12.436'	E44° 30.989'	Wild	14
Anjouan 8*	Jimilime	S12° 05.828'	E44° 27.960'	Wild	284
Anjouan 9	Hajaho	S12° 07.394'	E44° 29.196'	Wild	22
Anjouan 10	Mahale	S12° 09.142'	E44° 29.929'	Wild	49
Anjouan 11	Ongoni ya Marahani	S12° 10.819'	E44° 30.632'	Wild	38
Anjouan 12	Bambao Matsanga (Hospital)	S12° 11.980'	E44° 30.683'	Wild	47
Anjouan 13	Mromvovo	S12° 11.275'	E44° 29.542'	Wild	197
Anjouan 14	Moudiriyani/Nyaboimio	S12° 21.526'	E44° 31.152'	Wild	230
Anjouan 15	Bweju	S12° 21.107'	E44° 30.888'	Wild	327
Anjouan 16*	Nyombeni	S12° 21.561'	E44° 30.807'	Wild	257

Anjouan 17	Tshoroni	-	-	Wild	-
Anjouan 18	Dagi	S12° 19.855'	E44° 29.516'	Wild	542
Anjouan 19	Baoramafouga	S12° 22.419'	E43° 52.119'	Wild	44
Anjouan 20	Liwara	S12° 19.360'	E44° 30.259'	Wild	609
Anjouan 21	Zipwepweri	S12° 18.866'	E44° 30.057'	Wild	674
Anjouan 22	Crde mremani	S12° 18.650	E44° 30.043'	Wild	-
Anjouan 23	Lavaniju	-	-	Wild	-
Anjouan 24	Kohani	-	-	Wild	-
Anjouan 25	Pomoni	S12° 16.968'	E44° 24.685'	Wild	-
Anjouan 26	Shitsacouni	S12° 15.988'	E44° 23.770'	Wild	-
Anjouan 27	Maranare	S12° 14.031'	E44° 18.824'	Wild	-
Anjouan 28	Badramji	S12° 12.619'	E44° 16.341'	Wild	-
Anjouan 29	Simia mvonyi	S12° 12.543'	E44° 16.378'	Wild	-
Anjouan 30	Nyobeni	S12° 11.523'	E44° 14.741'	Wild	129
Anjouan 31	Sima	S12° 12.275	E44° 16.367	Wild	-
Anjouan 32*	Bougweni	S12° 12.375	E44° 17.964	Wild	-
Anjouan 33	Hombo	S12° 10.627	E44° 24.108	Wild	-
Anjouan 34	Foubouni	S12° 11.597	E44° 19.806	Wild	-
Anjouan 35	Mvoure	S12° 11.573	E44° 21.294	Wild	-
Anjouan 36	Njimandra	S12° 11.660	E44° 22.004	Wild	-
Anjouan 37	Micontsy	S12° 09.678	E44° 24.569	Wild	-
Anjouan 38	Nyatraga	-	-	Wild	13
Anjouan 39	-	S12° 08.216	E44° 26.186	Wild	53

APPENDIX II: Primers used for microsatellite DNA analyses indicating both the forward and reverse primer sequences, expected product size ranges, dye name and the source.

Primer	Upper sequence (5'-3')	Lower sequence (5'-3')	(Tm in °C)	Product size	Fluorochrome	Source
Name				Range (bp)		
A29	CAACTTCAACTGAAATC	AAACAGTACATT-	52	128-175	FAM	(Solignac <i>et al.</i> , 2003)
	CG	GTGACCC				
Ap33	TTTCTTTTGTG-	AAATATGGCGAAC-	52	225-247	HEX	(Solignac <i>et al.</i> , 2003)
	GACAGCG	GTGTG				
AP289	AGCTAGGTCTTCCTAA-	TTCGACCGCAA-	52	174-288	HEX	(Solignac <i>et al.</i> , 2003)
	GAGTGTG	TAACATTC				
A56	CCCCAGATGTCGCCATT	TCATTCTTCGCGAAC	55	282	TET	(Solignac <i>et al.</i> , 2003)
	C	CG				
A35	GTACACGGTTGCAC-	CTTCGATGGTCGTT-	52	94-123	HEX	(Solignac <i>et al.</i> , 2003)
	GGTTG	GTACCC				
B124	GCAACAGGTGGGTTA-	CAGGATAGGG-	52	216-232	HEX	(Solignac <i>et al.</i> , 2003)
	GAG	TAGGTAAGCAG				
A008	CGAAGGTAAGGTAAAT	GGCGGTTAAAGTTCTGG	52	165-181	TET	(Solignac <i>et al.</i> , 2003)
	GGAAC					

A113	CTCGAATCGTGGCGTCC	CCTGTATTTGCAAC- CTCGC	55	202-234	FAM	(Shaibi, Latorff and Moritz, 2008)
AP273	GATCTT- GTGTAAACAGCCG	GATCTCTGGCAGAC- GAAGAG	52	106-110	FAM	(Solignac <i>et al.</i> , 2003)
A088	CGAATTAAACCGATT- GTCG	GATCG- CAATTATTGAAGGAG	52	136-149	HEX	(Solignac <i>et al.</i> , 2003)
AC306	GAA- TATGCCGCTGCCACC	TTTCGTT- GCATCCGAGCG	55	165-185	FAM	(Solignac <i>et al.</i> , 2003)
A28	GAAGAGCGTTGGTT- GCAGG	GCCGTTCATGGTTAC- CACG	52	128-134	FAM	(Solignac <i>et al.</i> , 2003)
AP081	GGATCGTCGAGGC GTT- GA	GAAAGTATTCCGCCGAG CA	55	124-136	TET	(Solignac <i>et al.</i> , 2003)
A43	CACCGAAACAA- GATGCAAG	CCGCTCATTAAAGA- TATCCG	52	124-154	TET	(Solignac <i>et al.</i> , 2003)
Sex 1	AGTG- CAAAATCCAATCATC	ATTCGATCACCCAAA- GAA	52	142-187	HEX	(Shaibi, Latorff and Moritz, 2008)
UN467-	TTCCACAATAGA-	AATTGGAGAACACAG-	52	261-288	FAM	(Shaibi, Latorff and

16603	TAAAACAATACG	CATC					Moritz, 2008)
UN467-	TGTAATAC-	GAAAAACTGTGCTCGTG	52	174-192	HEX	(Shaibi, Latorff and	
370	GTGATGTCAAAACA	AA					Moritz, 2008)
UN462.4	AAAATGACAAAAAC-	AATCGTTGCCAAGA-	48	168-178	FAM	(Shaibi, Latorff and	
987	GGAGAA	GAATC					Moritz, 2008)
A24	CACAAGTTCCAACAATG	CACATTGAGGATGAGCG	55	93-116	TET	(Solignac <i>et al.</i> , 2003)	
	C						
UN351	AGCATACTTCTTCAC-	TCCGTTATGCTTCATT	52	147-166	FAM	(Shaibi, Latorff and	
	CGAACACC	TCGA					Moritz, 2008)

**APPENDIX III: ClustalO multiple sequence alignment representing all the sequenced COI-COII intergenic spanning the region between positions 3441 to 3880 of the reference sequence (NC\_001566.1).**

NC_001566.1_(RefSeq)	AATTCAACCAATCATCACTTGAATGATTAAATTTTACCCCTCTAGATCATTACATT	3300
MOH30	-----	0
NGA25_(reversed)	-----	0
MOH36B	-----	0
ANJ18B	-----	0
ANJ8B	-----	0
NGA34	-----	0
MOH3	-----	0
ANJ15A	-----	0
MOH32	-----	0
ANJ26A	-----	0
ANJ22	-----	0
ANJ25	-----	0
ANJ28	-----	0
ANJ23	-----	0
ANJ19B	-----	0
NGA17	-----	0
NGA23	-----	0
MOH25B	-----	0
NGA22	-----	0
NGA26A_(reversed)	-----	0
ANJ33	-----	0
MOH24	-----	0
ANJ35A_2	-----	0
ANJ35B	-----	0
ANJ34	-----	0
ANJ31	-----	0
ANJ30	-----	0
Anj21	-----	0
MOH28A	-----	0
NGA11B	-----	0
ANJ18A	-----	0
MOH10	-----	0
NGA26B_(reversed)	-----	0
NGA16	-----	0
MOH12C	-----	0
NGA9	-----ATGCATTGAACCTAACATTAAAGTATTTAAACTTTAT	47
NGA6	-----TGCATTGAACCTAACATTAAAGTATTTAAACTTTAT	46
NGA14A	-----TGCATTGAACCTAACATTAAAGTATTTAAACTTTAT	46
NGA7A	-----TGCATTGAACCTAACATTAAAGTATTTAAACTTTAT	46
NGA11A	-----TGCATTGAACCTAACATTAAAGTATTTAAACTTTAT	46
NGA15	-----TGCATTGAACCTAACATTAAAGTATTTAAACTTTAT	46
NGA14B	-----TGCATTGAACCTAACATTAAAGTATTTAAACTTTAT	46
NGA20	-----TGCATTGAACCTAACATTAAAGTATTTAAACTTTAT	46
NGA10	-----TGCATTGAACCTAACATTAAAGTATTTAAACTTTAT	46
NGA5B	-----TGCATTGAACCTAACATTAAAGTATTTAAACTTTAT	46
ANJ35A	-----CTT--TTATTAAATTATATTATAATTATTTAAAAAT----	40
ANJ26B	-----0	
NGA7C	-----AGAATAAGTGCATTGAACCTAACATTAAAGTATTTAAACTTTAT	54
NGA31	-----TTAAGATTCAATATAAAGTATTTAAACTTTAT	36
MOH29	-----TTTGATTAAAAATAATGTTTATTATCTTTAT	35
NGA1	-----TAAGATTAAAAATAAAGTATTAAATACATTATTTAT	35
MOH15B	-----0	
MOH40	-----TTTTA	5
MOH42B	-----TTTTA	5
MOH39	-----TTTTA	5
ANJ39	-----0	
ANJ17	-----0	
ANJ37	-----0	
ANJ10	-----GC-----	2
MOH42A	-----0	
MOH14	-----0	
MOH27	-----0	
MOH7	-----0	
NGA8B	-----GAACCTAACATTAAAGTATTTAAACTTTAT	40
ANJ36	-----0	
MOH23C	-----0	
MOH8	-----0	
MOH17B	-----0	

MOH16B	-----	0
MOH11A	-----	0
NGA7B	-----	0
NGA2A	-----	0
NGA2B	-----	0
ANJ20B	-----	0
MOH25A_(reversed)	-----	0
NGA28A	--GCACTGAACT-----TATGATT-CAAAAATAAGGTTTATAAATTTTAT	45
MOH9	--GCATTGAACT-----TAAGATT-CAAATATAAAAGTATTTTAAACTTTAT	45
NGA21	--GCATTGAACT-----TAAGATT-CAAATATAAAAGTATTTTAAACTTTAT	45
NGA11C	--GCATTGAACT-----TAAGATT-CAAATATAAAAGTATTTTAAACTTTAT	45
NGA3	--GCATTGAACT-----TAAGATT-CAAATATAAAAGTATTTTAAACTTTAT	45
ANJ29	-----0	
KT828418.1_(A1_MAD3)	-----AACT-----TAAGATTCAAATATAAAAGT-ATTTTAAACTTTAT	39
KT828427.1_(A1_MAD12)	-----AACT-----TAAGATTCAAATATAAAAGT-ATTTTAAACTTTAT	39
NGA14C	-----AATTCAAATATAAAAGTATTTATAAAACTTTAT	33
NGA12	-----TCAATATAAAAGTATTTTAAACTTTAT	29
NGA5A	-----TGCATTGAACTTAAGATICAATATAAAAGTATTTTAAACTTTAT	46
NGA8A	-----ATGCATTGAACTTAAGATTCAAATATAAAAGTATTTTAAACTTTAT	47
ANJ12	-----GTGCATTGAACTTAAGATTCAAATATAAAAGTATTTTAAACTTTAT	47
MOH11B	-----GTGCATTGAACTTAAGATTCAAATATAAAAGTATTTTAAACTTTAT	47
NGA4	-----AGTCATTGAACTTAAGATTCAAATATAAAAGTATTTTAAACTTTAT	48
MOH34	-----AGTCATTGAACTTAAGATTCAAATATAAAAGTATTTTAAACTTTAT	48
NGA15B	-----AATGCATTGAATTAAAGATTCAAATATAAAAGTATTTTAAACTTTAT	48
MOH19	-----GTGCATTGAACTTAAGATTCAAATATAAAAGTATTTTAAACTTTAT	47
MOH23B	-----0	
MOH36A	-----ATTCAAATATAAAAGTATTTTAAACTTTAT	31
ANJ2	-----0	
ANJ27	-----CATTGAATTAAAGCTTCAA	19
MOH37	-----TTTCTTTTAT	10
MOH26	-----TTTTAATTTCCTTTAAT	19
AJ20C	-----0	
MOH18	-----0	
ANJ06	-----0	
ANJ5	-----0	
ANJ7B	-----0	
ANJ1	-----0	
MOH4	-----0	
NGA19	----CCATGTCAGTCATTGAACTTAAGATICAATATAAAAGTATTTTAAACTTTAT	55
MOH15A	----CAGAATAAGTCATTGAACTTAAGATTCAAATATAAAAGTATTTTAAACTTTAT	55
NGA13	-----0	
NC_001566.1_(RefSeq)	TAGAAATTCCATTATTAATTAAAAATTAAATTAAATCAATTAAATTAAATTAAATTAA	3360
MOH30	-----AAAAAAGGGTTT-----TTTAATTTTCT	25
NGA25_(reversed)	-----CTTAAGATTCAAATATAAAAGTATT-----TTAAACTTTAT	37
MOH36B	-----TTTTAACTTTA	12
ANJ18B	-----ACTTTA	7
ANJ8B	-----CTTTAT	7
NGA34	-----CTTTAT	7
MOH3	-----CTTTAT	7
ANJ15A	-----CTTTAT	7
MOH32	-----CTTTAT	7
ANJ26A	-----CTTTAT	7
ANJ22	-----CTTTAT	7
ANJ25	-----CTTTAT	7
ANJ28	-----CTTTAT	7
ANJ23	-----0	
ANJ19B	-----CTTTA	6
NGA17	-----CTTTA	6
NGA23	-----CTTTA	6
MOH25B	-----CTTTA	6
NGA22	-----CTTTA	6
NGA26A_(reversed)	-----TTTTAT	6
ANJ33	-----TTTTAT	6
MOH24	-----0	
ANJ35A_2	-----0	
ANJ35B	-----0	
ANJ34	-----0	
ANJ31	-----0	
ANJ30	-----0	
Anj21	-----0	
MOH28A	-----ATTGAACTTAAGATTCAAATATA-----AAGTATTTTAAACTTTAT	43
NGA11B	-----ATTGAACTTAAGATTCAAATATA-----AAGTATTTTAAACTTTAT	43
ANJ18A	-----ATTGAACTTAAGATTCAAATATA-----AAGTATTTTAAACTTTAT	43
MOH10	-----ATTGAACTTAAGATTCAAATATA-----AAGTATTTTAAACTTTAT	43
NGA26B_(reversed)	-----CATTGAACTTAAGATTCAAATATA-----AAGTATTTTAAACTTTAT	44
NGA16	-----CATCGAACCTTAAGATTCAAATATA-----AAGTATTTTAAACTTTAT	44
MOH12C	-----0	

NGA9 TAAAATTAATAAATTATA-TAAATAAAAACAAAATATA-ACAAAATATATTATTTAAAAA

105

NGA6	TAAAATTAAATAATTAATATAAATAAAAACAAAATATA-ACAAAATATATTATTAAAA	105
NGA14A	TAAAATTAAATAATTAATATAAATAAAAACAAAATATA-ACAAAATATATTATTAAAA	105
NGA7A	TAAAATTAAATAATTAATATAAATAAAAACAAAATATA-ACAAAATATATTATTAAAA	105
NGA11A	TAAAATTAAATAATTAATATAAATAAAAACAAAATATA-ACAAAATATATTATTAAAA	105
NGA15	TAAAATTAAATAATTAATATAAATAAAAACAAAATATA-ACAAAATATATTATTAAAA	105
NGA14B	TAAAATTAAATAATTAATATAAATAAAAACAAAATATA-ACAAAATATATTATTAAAA	105
NGA20	TAAAATTAAATAATTAATATAAATAAAAACAAAATATA-ACAAAATATATTATTAAAA	105
NGA10	TAAAATTAAATAATTAATATAAATAAAAACAAAATATA-ACAAAATATATTATTAAAA	105
NGA5B	TAAAATTAAATAATTAATATAAATAAAAACAAAATATA-ACAAAATATATTATTAAAA	105
ANJ35A	-----40-----	
ANJ26B	-----AATATA-ACGAAATATTTTATTAAAA 26	
NGA7C	TAAAATTAAATAATTAATATAAATAAAAACAAAATATA-ACAAAATATATTATTAAAA	113
NGA31	TAAAATTAAATAATTAATATAAATAAAAACAAAATATA-ACAAAATATATTATTAAAA	95
MOH29	TAAAATTAAATAATTAATATAAATAAAAACAAAATATA-ACAAAATATATTATTAAAA	94
NGA1	TAAAATTAAATAATTAATATAAATAAAAACAAAATATA-ACAAAATATATTATTAAAA	94
MOH15B	-----TAAAATAAAACAAAATATA-ACAAAATATATTATTAAAA 40	
MOH40	TTAAAATTAAATAATTAATATAAATAAAAACAAAATATA-ACAAAATATATTATTAAAA	64
MOH42B	TTAAAATTAAATAATTAATATAAATAAAAACAAAATATA-ACAAAATATATTATTAAAA	64
MOH39	TTAAAATTAAATAATTAATATAAATAAAAACAAAATATA-ACAAAATATATTATTAAAA	64
ANJ39	-----0-----	
ANJ17	-----0-----	
ANJ37	-----0-----	
ANJ10	-----2-----	
MOH42A	-TAAATTAAATAATTAATAAAATAAAACAAAATATA-ACAAAATATATTATTAAAA	58
MOH14	-----TAAATTATATAAATAAAACAAAATATA-ACAAAATATATTATTATTAAAA	49
MOH27	----TAAATAATTAAATATAAATAAAACAAAATATA-ACAAAATATATTATTATTAAAA	54
MOH7	----TAAATAATTAAATATAAATAAAACAAAATATA-ACAAAATATATTATTATTAAAA	54
NGA8B	TAAAATTAAATAATTAATAAAATAAAACAAAATATA-ACAAAATATATTATTATTAAAA	99
ANJ36	-----ACAAAATATA-ACAAAATATATTATTATTAAAA 30	
MOH23C	-----TGGCAGAA-----TAAGT 13	
M0H8	-----TAACAAAATATATTATTATTAAAA 22	
MOH17B	-----TGGCAGAATAGTGCATTGAATTAA 25	
MOH16B	-----AAATATATTATTATAATTAAAT--- 23	
MOH11A	-----0-----	
NGA7B	-----0-----	
NGA2A	-----0-----	
NGA2B	-----0-----	
ANJ20B	-----CAAATATATTATTATTAAAA 18	
MOH25A_(reversed)	-----0-----	
NGA28A	TAAAATTAAATAATTAAT-ATAAAATAAAACAAAATATAACAAAATATATTATTAAAA	104
MOH9	TAAAATTAAATAATTAAT-ATAAAATAAAACAAAATATAACAAAATATATTATTAAAA	104
NGA21	TAAAATTAAATAATTAAT-ATAAAATAAAACAAAATATAACAAAATATATTATTAAAA	104
NGA11C	TAAAATTAAATAATTAAT-ATAAAATAAAACAAAATATAACAAAATATATTATTAAAA	104
NGA3	TAAAATTAAATAATTAAT-ATAAAATAAAACAAAATATAACAAAATATATTATTAAAA	104
ANJ29	-----CAAATATAACAAAATATATTATTAAAA 29	
KT828418.1_(A1_MAD3)	TAAAATTAAATAATTAAT-ATAAAATAAAACAAAATATAACAAAATATATTATTATTAAAA	98
KT828427.1_(A1_MAD12)	TAAAATTAAATAATTAAT-ATAAAATAAAACAAAATATAACAAAATATATTATTATTAAAA	98
NGA14C	TAAAATTAAATAATTAAT-ATAAAATAAAACAAAATATAACAAAATATATTATTATTAAAA	92
NGA12	TAAAATTAAATAATTAAT-ATAAAATAAAACAAAATATAACAAAATATATTATTATTAAAA	88
NGA5A	TAAAATTAAATAATTAATATAAAGAACAAAATATAACAAAATATATTATTATTAAAA	106
NGA8A	TAAAATTAAATAATTAAT-AAAATAAAACAAAATATAACAAAATATATTATTATTAAAA	106
ANJ12	TAAAATTAAATAATTAAT-AAAATAAAACAAAATATAACAAAATATATTATTATTAAAA	106
MOH11B	-----TAAAATTAAATAATTAAT-AAAATAAAACAAAATATAACAAAATATATTATTATTAAAA	106
NGA4	TAAAATTAAATAATTAATATAAATAAAACAAAATATAACAAAATATATTATTATTAAAA	108
MOH34	TAAAATTAAATAATTAATATAAATAAAACAAAATATAACAAAATATATTATTATTAAAA	108
NGA15B	TAAAATTAAATAATTAATATAAATAAAACAAAATATAACAAAATATATTATTATTAAAA	108
MOH19	TAAAATTAAATAATTAATATAAATAAAACAAAATATAACAAAATATATTATTATTAAAA	107
MOH23B	-----0-----	
MOH36A	TAAAT-TA-ATAAATTAAATATAAATAAAACAAAATATAACAAAATATATTATTATTAAAA	89
ANJ2	-----0-----	
ANJ27	ATATA-AA-GTGATAAACTTTTATTAAAATATTAA----- 59	
MOH37	TAAAT-TA-ATAAATTAAATATAAATAAAACAAAATATAACAAAATATATTATTATTAAAA	68
MOH26	TAAAT-TA-ATAAATTAAATATAAATAAAACAAAATATAACAAAATATATTATTATTAAAA	77
AJ20C	-----0-----	
MOH18	-----0-----	
ANJ06	-----0-----	
ANJ5	-----0-----	
ANJ7B	-----0-----	
ANJ1	-----0-----	
MOH4	-----ATGAGTATGAAATAGAAACAAAATATA-ACAAAATATATTATTATTAAAA 47	
NGA19	TAAAATTAAATAATTAATATAAATAAAACAAAATATA-ACAAAATATATTATTATTAAAA	114
MOH15A	TAAAATTAAATAATTAATATAAATAAAACAAAATATA-ACAAAATATATTATTATTAAAA	114
NGA13	-----0-----	
NC_001566.1_(RefSeq)	ATGGCAGAATAAGTCATTGAC-----TTAAGATT-CAAATATAAA-GTATTTTTA 3410	
MOH30	TTAAATTAAATAATTAATATAAATAAAACAAAATATA-ACAAAATATATTATTATTAAAA	84
NGA25_(reversed)	TAAAATTAAATAATTAATATAAATAAAACAAAATATA-ACAAAATATATTATTATTAAAA	96
MOH36B	TTAAATTAAATAATTAATATAAATAAAACAAAATATA-ACAAAATATATTATTATTAAAA	71
ANJ18B	TTAAATTAAATAATTAATATAAATAAAACAAAATATA-ACAAAATATATTATTATTAAAA	66
ANJ8B	TAAAATTAAATAATTAATATAAATAAAACAAAATATA-ACAAAATATATTATTATTAAAA	66
NGA34	TAAAATTAAATAATTAATATAAATAAAACAAAATATA-ACAAAATATATTATTATTAAAA	66

MOH3	TAAAATTAAATAAATTAAATATAAAAACAAAATATA-ACAAAATATATTATTAAAAA	66
ANJ15A	TAAAATTAAATAAATTAAATATAAAAACAAAATATA-ACAAAATATATTATTAAAAA	66
MOH32	TAAAATTAAATAAATTAAATATAAAAACAAAATATA-ACAAAATATATTATTAAAAA	66
ANJ26A	TAAAATTAAATAAATTAAATATAAAAACAAAATATA-ACAAAATATATTATTAAAAA	66
ANJ22	TAAAATTAAATAAATTAAATATAAAAACAAAATATA-ACAAAATATATTATTAAAAA	66
ANJ25	TAAAATTAAATAAATTAAATATAAAAACAAAATATA-ACAAAATATATTATTAAAAA	66
ANJ28	TAAAATTAAATAAATTAAATATAAAAACAAAATATA-ACAAAATATATTATTAAAAA	66
ANJ23	-----0-----0	66
ANJ19B	TTAAAATTAAATAAATTAAATATAAAAACAAAATATA-ACAAAATATATTATTAAAAA	65
NGA17	TTAAAATTAAATAAATTAAATATAAAAACAAAATATA-ACAAAATATATTATTAAAAA	65
NGA23	TTAAAATTAAATAAATTAAATATAAAAACAAAATATA-ACAAAATATATTATTAAAAA	65
MOH25B	TTAAAATTAAATAAATTAAATATAAAAACAAAATATA-ACAAAATATATTATTAAAAA	65
NGA22	TTAAAATTAAATAAATTAAATATAAAAACAAAATATA-ACAAAATATATTATTAAAAA	65
NGA26A_(reversed)	TAAAATTAAATAAATTAAATATAAAAACAAAATATA-ACAAAATATATTATTAAAAA	65
ANJ33	TAAAATTAAATAAATTAAATATAAAAACAAAATATA-ACAAAATATATTATTAAAAA	65
MOH24	-----0-----0	65
ANJ35A_2	-----0-----0	65
ANJ35B	-----0-----0	65
ANJ34	-----0-----0	65
ANJ31	-----0-----0	65
ANJ30	-----0-----0	65
Anj21	-----0-----0	65
MOH28A	TAAAATTAAATAAATTAAATAGAATAAAAACAAAATATA-ACAAAATATATTATTAAAA	102
NGA11B	TAAAATTAAATAAATTAAATATAAAAACAAAATATA-ACAAAATATATTATTAAAA	102
ANJ18A	TAAAATTAAATAAATTAAATATAAAAACAAAATATA-ACAAAATATATTATTAAAA	102
MOH10	TAAAATTAAATAAATTAAATATAAAAACAAAATATA-ACAAAATATATTATTAAAA	102
NGA26B_(reversed)	TAAAATTAAATAAATTAAATATAAAAACAAAATATA-ACAAAATATATTATTAAAA	103
NGA16	TAAAATTAAATAAATTAAATATAAAAACAAAATATA-ACAAAATATATTATTAAAA	103
MOH12C	-----AAGAACAAAATATA-ACAAAATATATTATTAAAA-----34	103
NGA9	TTTAAATTATTAAAATTCCC---CACTTAATTCATATTA-ATTTAAAATAAATTAA	160
NGA6	TTTAAATTATTAAAATTCCC---CACTTAATTCATATTA-ATTTAAAATAAATTAA	160
NGA14A	TTTAAATTATTAAAATTCCC---CACTTAATTCATATTA-ATTTAAAATAAATTAA	160
NGA7A	TTTAAATTATTAAAATTCCC---CACTTAATTCATATTA-ATTTAAAATAAATTAA	160
NGA11A	TTTAAATTATTAAAATTCCC---CACTTAATTCATATTA-ATTTAAAATAAATTAA	160
NGA15	TTTAAATTATTAAAATTCCC---CACTTAATTCATATTA-ATTTAAAATAAATTAA	160
NGA14B	TTTAAATTATTAAAATTCCC---CACTTAATTCATATTA-ATTTAAAATAAATTAA	160
NGA20	TTTAAATTATTAAAATTCCC---CACTTAATTCATATTA-ATTTAAAATAAATTAA	160
NGA10	TTTAAATTATTAAAATTCCC---CACTTAATTCATATTA-ATTTAAAATAAATTAA	160
NGA5B	TTTAAATTATTAAAATTCCC---CACTTAATTCATATTA-ATTTAAAATAAATTAA	160
ANJ35A	-----40-----40	160
ANJ26B	TTTAAATTATTAAAATTCCC---CACTTAATTCATATTA-ATTTAAAATAAATTAA	81
NGA7C	TTTAAATTATTAAAATTCCC---CACTTAATTCATATTA-ATTTAAAATAAATTAA	168
NGA31	TTTAAATTATTAAAATTCCC---CACTTAATTCATATTA-ATTTAAAATAAATTAA	150
MOH29	TTTAAATTATTAAAATTCCC---CACTTAATTCATATTA-ATTTAAAATAAATTAA	149
NGA1	TTTAAATTATTAAAATTCCC---CACTTAATTCATATTA-ATTTAAAATAAATTAA	149
MOH15B	TTTAAATTATTAAAATTCCC---CACTTAATTCATATTA-ATTTAAAATAAATTAA	95
MOH40	TTTAAATTATTAAAATTCCC---CACTTAATTCATATTA-ATTTAAAATAAATTAA	119
MOH42B	TTTAAATTATTAAAATTCCC---CACTTAATTCATATTA-ATTTAAAATAAATTAA	119
MOH39	TTTAAATTATTAAAATTCCC---CACTTAATTCATATTA-ATTTAAAATAAATTAA	119
ANJ39	-----0-----0	119
ANJ17	-----0-----0	119
ANJ37	-----0-----0	119
ANJ10	-----2-----2	119
MOH42A	TITAATTITATTAAAATTCCC---CACTTAATTCATATTA-ATTTAAAATAAATTAA	113
MOH14	TITAATTITATTAAAATTCCC---CACGTAATTCATATTA-ATTTAAAATAAATTAA	104
MOH27	TITAATTITATTAAAATTCCC---CACTTAATTCATATTA-ATTTAAAATAAATTAA	109
MOH7	TITAATTITATTAAAATTCCC---CACTTAATTCATATTA-ATTTAAAATAAATTAA	109
NGA8B	TITAATTITATTAAAATTCCC---CACTTAATTCATATTA-ATTTAAAATAAATTAA	154
ANJ36	TITAATTITATTAAAATTCCC---CACTTAATTCATATTA-ATTTAAAATAAATTAA	85
MOH23C	GCATAAAATTAAAGCATAAAATATAAAGTTAATAAAAATTAAAT-TAAAATTATATAA	72
M0H8	TITAATTITATTAAAATTCCCACTGAAT---TCATATTAAATT-TAAAATAAATTAA	77
MOH17B	GCTTCAAATATAAAGTTGAT---AACCTTTAT---T-AAAATTAAATTATAA	72
MOH16B	-----CTATTAATTAAAATTCCC---CACTTAATTCATATTAATT-TAAAATAAATTAA	72
MOH11A	-----0-----0	72
NGA7B	-----0-----0	72
NGA2A	-----0-----0	72
NGA2B	-----0-----0	72
ANJ20B	TITAATTITATTAAAATTCCC---CACGTAATTCATATTAATT-TAAAATAAATTAA	73
MOH25A_(reversed)	-----0-----0	73
NGA28A	TITAATTITATTAAAATTCCC---CACTTAATTCATATTAATT-TAAAATAAATTAA	159
MOH9	TITAATTITATTAAAATTCCC---CACTTAATTCATATGAATT-TAAAATAAATTAA	159
NGA21	TITAATTITATTAAAATTCCC---CACTTAATTCATATTAATT-TAAAATAAATTAA	159
NGA11C	TITAATTITATTAAAATTCCC---CACTTAATTCATATTAATT-TAAAATAAATTAA	159
NGA3	TITAATTITATTAAAATTCCC---CACTTAATTCATATTAATT-TAAAATAAATTAA	159
ANJ29	TITAATTITATTAAAATTCCC---CACTTAATTCATATTAATT-TAAAATAAATTAA	84
KT828418.1_(A1_MAD3)	TITAATTITATTAAAATTCCC---CACTTAATTCATATTAATT-TAAAATAAATTAA	153
KT828427.1_(A1_MAD12)	TITAATTITATTAAAATTCCC---CACTTAATTCATATTAATT-TAAAATAAATTAA	153
NGA14C	TITAATTITATTAAAATTCCC---CACTTAATTCATATTAATT-TAAAATAAATTAA	147

NGA12	TTTAATTATTAAAATCCC---CACTTAATTATTAATTAAATTAAATAAA	143
NGA5A	TTTAATTATTAAAATCCC---CACTTAATTATTAATTAAATTAAATAAA	161
NGA8A	TTTAATTATTAAAATCCC---CACTTAATTATTAATTAAATTAAATAAA	161
ANJ12	TTTAATTATTAAAATCCC---CACTTAATTATTAATTAAATTAAATAAA	161
MOH11B	TTTAATTATTAAAATCCC---CACTTAATTATTAATTAAATTAAATAAA	161
NGA4	T-TAATTATTAAAATCCC---CACTTAATTATTAATTAAATTAAATAAA	162
MOH34	T-TAATTATTAAAATCCC---CACTTAATTATTAATTAAATTAAATAAA	162
NGA15B	T-TAATTATTAAAATCCC---CACTTAATTATTAATTAAATTAAATAAA	162
MOH19	T-TAATTATTAAAATCCC---CACTTAATTATTAATTAAATTAAATAAA	162
MOH23B	-----0-----	
MOH36A	TTAATTATTAAAATCCC---CACTTAATTATTAATTAAATTAAATAAA	144
ANJ12	-----0-----	
ANJ27	-----59-----	
MOH37	TTAATTATTAAAATCCC---CACTTAATTATTAATTAAATTAAATAAA	123
MOH26	TTAATTATTAAAATCCC---CACTTAATTATTAATTAAATTAAATAAA	132
AJ20C	-----0-----	
MOH18	-----0-----	
ANJ06	-----0-----	
ANJ5	-----0-----	
ANJ7B	-----0-----	
ANJ1	-----0-----	
MOH4	TTAATTATTAAAATCCC---CACTGA--ATTGGATTAATTAAAATAATTAAATAAA	101
NGA19	TTAATTATTAAAATCCC---CACTTAATTATTAATTAAATTAAATAAA	170
MOH15A	TTAATTATTAAAATCCC---CACTTAATTATTAATTAAATTAAATAAA	169
NGA13	-----0-----	
NC_001566.1_(RefSeq)	AACTTTTATTAAAATTCCC---CACTTAATTATTAATTAAAAATAATTAAATAAA	3466
MOH30	TTAATTATTAAAATCCC---CACTTAATTATTAATTAAAATAATTAAATAAA	139
NGA25_(reversed)	TTAATTATTAAAATCCC---CACTTAATTATTAATTAAAATAATTAAATAAA	151
MOH36B	TTAATTATTAAAATCCC---CACTTAATTATTAATTAAAATAATTAAATAAA	126
ANJ18B	TTAATTATTAAAATCCC---CACTTAATTATTAATTAAAATAATTAAATAAA	121
ANJ8B	TTAATTATTAAAATCCC---CACTTAATTATTAATTAAAATAATTAAATAAA	121
NGA34	TTAATTATTAAAATCCC---CACTTAATTATTAATTAAAATAATTAAATAAA	121
MOH3	TTAATTATTAAAATCCC---CACTTAATTATTAATTAAAATAATTAAATAAA	121
ANJ15A	TTAATTATTAAAATCCC---CACTTAATTATTAATTAAAATAATTAAATAAA	121
MOH32	TTAATTATTAAAATCCC---CACTTAATTATTAATTAAAATAATTAAATAAA	121
ANJ26A	TTAATTATTAAAATCCC---CACTTAATTATTAATTAAAATAATTAAATAAA	121
ANJ22	TTAATTATTAAAATCCC---CACTTAATTATTAATTAAAATAATTAAATAAA	121
ANJ25	TTAATTATTAAAATCCC---CACTTAATTATTAATTAAAATAATTAAATAAA	121
ANJ28	TTAATTATTAAAATCCC---CACTTAATTATTAATTAAAATAATTAAATAAA	121
ANJ23	-----CATCCCC---CTTGAAATTATTAATTAAAATAATTAAATAAA-----42	
ANJ19B	TTAATTATTAAAATCCC---CACTTAATTATTAATTAAAATAATTAAATAAA	120
NGA17	TTAATTATTAAAATCCC---CACTTAATTATTAATTAAAATAATTAAATAAA	120
NGA23	TTAATTATTAAAATCCC---CACTTAATTATTAATTAAAATAATTAAATAAA	120
MOH25B	TTAATTATTAAAATCCC---CACTTAATTATTAATTAAAATAATTAAATAAA	120
NGA22	TTAATTATTAAAATCCC---CACTTAATTATTAATTAAAATAATTAAATAAA	120
NGA26A_(reversed)	TTAATTATTAAAATCCC---CACTTAATTATTAATTAAAATAATTAAATAAA	120
ANJ33	TTAATTATTAAAATCCC---CACTTAATTATTAATTAAAATAATTAAATAAA	120
MOH24	-----0-----	
ANJ35A_2	-----0-----	
ANJ35B	-----0-----	
ANJ34	-----0-----	
ANJ31	-----0-----	
ANJ30	-----0-----	
Anj21	-----0-----	
MOH28A	TTAATTATTAAAATCCC---CACTTAACTCATATT-AATTAAAATAATTAAATAAA	157
NGA11B	TTAATTATTAAAATCCC---CACTTAATTATTAATT-AATTAAAATAATTAAATAAA	157
ANJ18A	TTAATTATTAAAATCCC---CACTTAATTATTAATT-AATTAAAATAATTAAATAAA	157
MOH10	TTAATTATTAAAATCCC---CACTGAATTATTAATT-AATTAAAATAATTAAATAAA	157
NGA26B_(reversed)	TTAATTATTAAAATCCC---CACTTAATTATTAATT-AATTAAAATAATTAAATAAA	158
NGA16	TTAATTATTAAAATCCC---CACTTAATTATTAATT-AATTAAAATAATTAAATAAA	158
MOH12C	TTAATTATTAAAATCCC---CACTTAATTATTAATT-AATTAAAATAATTAAATAAA	89
NGA9	CAATTTTAATAAAAATAATTAAATTTTATTGAAATTTTAATTCAATCT-	219
NGA6	CAATTTTAATAAAAATAATTAAATTATTTATTGAAATTTTAATTCAATCT-	219
NGA14A	CAATTTTAATAAAAATAATTAAATTATTTATTGAAATTTTAATTCAATCT-	219
NGA7A	CAATTTTAATAAAAATAATTAAATTATTTATTGAAATTTTAATTCAATCT-	219
NGA11A	CAATTTTAATAAAAATAATTAAATTATTTATTGAAATTTTAATTCAATCT-	219
NGA15	CAATTTTAATAAAAATAATTAAATTATTTATTGAAATTTTAATTCAATCT-	219
NGA14B	CAATTTTAATAAAAATAATTAAATTATTTATTGAAATTTTAATTCAATCT-	219
NGA20	CAATTTTAATAAAAATAATTAAATTATTTATTGAAATTTTAATTCAATCT-	219
NGA10	CAATTTTAATAAAAATAATTAAATTATTTATTGAAATTTTAATTCAATCT-	219
NGA5B	CAATTTTAATAAAAATAATTAAATTATTTATTGAAATTTTAATTCAATCT-	219
ANJ35A	-----40-----	
ANJ26B	CAATTTTAATAAAAATAATTAAATTATTTATTGAAATTTTAATTCAATCT-	227
NGA7C	CAATTTTAATAAAAATAATTAAATTATTTATTGAAATTTTAATTCAATCT-	209
NGA31	CAATTTTAATAAAAATAATTAAATTATTTATTGAAATTTTAATTCAATCT-	208
MOH29	CAATTTTAATAAAAATAATTAAATTATTTATTGAAATTTTAATTCAATCT-	208
NGA1	CAATTTTAATAAAAATAATTAAATTATTTATTGAAATTTTAATTCAATCT-	154
MOH15B	CAATTTTAATAAAAATAATTAAATTATTTATTGAAATTTTAATTCAATCT-	

MOH40	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	178
MOH42B	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	178
MOH39	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	178
ANJ39	-----0	
ANJ17	-----0	
ANJ37	-----0	
ANJ10	-----2	
MOH42A	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	172
MOH14	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	163
MOH27	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	168
MOH7	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	168
NGA8B	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	213
ANJ36	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	144
MOH23C	TATTTTATTAAAATTCTACATGATTATATTCAACAAATCAAATCATACTA	132
MOH8	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	136
MOH17B	A-----73	
MOH16B	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	131
MOH11A	-----0	
NGA7B	-----0	
NGA2A	-----0	
NGA2B	-----0	
ANJ20B	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	132
MOH25A_(reversed)	-----GGGCAGAAT-----9	
NGA28A	CAATTTTAATAAAAATAATTAAATTATTTATATTGAATTTTAATTCAATCT-	218
MOH9	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	218
NGA21	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	218
NGA11C	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	218
NGA3	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	218
ANJ29	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	143
KT828418.1_(A1_MAD3)	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	212
KT828427.1_(A1_MAD12)	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	212
NGA14C	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	206
NGA12	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	202
NGA5A	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	220
NGA8A	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	220
ANJ12	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	220
MOH11B	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	220
NGA4	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	221
MOH34	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	221
NGA15B	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	221
MOH19	CAATTTTAATAAAAATAATTAAATTATTTATATTGAATTTTAATTCAATCT-	221
MOH23B	-----0	
MOH36A	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	203
ANJ2	-----0	
ANJ27	-----59	
MOH37	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	182
MOH26	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	191
AJ20C	-----0	
MOH18	-----0	
ANJ06	-----0	
ANJ5	-----0	
ANJ7B	-----0	
ANJ1	-----0	
MOH4	CGATTTTAATAAAAAGAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	160
NGA19	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	229
MOH15A	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	228
NGA13	-----0	
NC_001566.1_(RefSeq)	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	3525
MOH30	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	198
NGA25_(reversed)	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	210
MOH36B	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	185
ANJ18B	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	180
ANJ8B	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	180
NGA34	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	180
MOH3	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	180
ANJ15A	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	180
MOH32	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	180
ANJ26A	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	180
ANJ22	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	180
ANJ25	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	180
ANJ28	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	180
ANJ23	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	101
ANJ19B	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	179
NGA17	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	179
NGA23	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	179
MOH25B	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	179
NGA22	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	179
NGA26A_(reversed)	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	179
ANJ33	CAATTTTAATAAAAATAATTAAATTATTTATTTTATATTGAATTTTAATTCAATCT-	179
MOH24	-----0	

ANJ35A_2	-----	0
ANJ35B	-----	0
ANJ34	-----	0
ANJ31	-----	0
ANJ30	-----	0
Anj21	-----	0
MOH28A	CAATTTCTAATAAAAATAATTAAATTATTTTATATTGAATTTTAATTCAATCT-	216
NGA11B	CAATTTTAATAAAAATAATTAAATTATTTTATATTGAATTTTAATTCAATCT-	216
ANJ18A	CAATTTTAATAAAAATAATTAAATTATTTTATATTGAATTTTAATTCAATCT-	216
MOH10	CAATTTTAATAAAAATAATTAAATTATTTTATATTGAATTTTAATTCAATCT-	216
NGA26B_(reversed)	CAATTTTAATAAAAATAATTAAATTATTTTATATTGAATTTTAATTCAATCT-	217
NGA16	CAATTTTAATAAAAATAATTAAATTATTTTATATTGAATTTTAATTCAATCT-	217
MOH12C	CAATTTTAATAAAAATAATTAAATTATTTTATATTGAATTTTAATTCAATCT-	148
NGA9	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATTAAATAAAAACAAA-ATA	275
NGA6	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATTAAATAAAAACAAA-ATA	275
NGA14A	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATTAAATAAAAACAAA-ATA	275
NGA7A	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATTAAATAAAAACAAA-ATA	275
NGA11A	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATTAAATAAAAACAAA-ATA	275
NGA15	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATTAAATAAAAACAAA-ATA	275
NGA14B	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATTAAATAAAAACAAA-ATA	275
NGA20	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATTAAATAAAAACAAA-ATA	275
NGA10	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATTAAATAAAAACAAA-ATA	275
NGA5B	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAACAAA-ATA	275
ANJ35A	----- 40	
ANJ26B	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAACAAAATAT	197
NGA7C	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAACAAAATAT	284
NGA31	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAACAAAATAT	266
MOH29	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAACAAAATAT	265
NGA1	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAACAAAATAT	265
MOH15B	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAACAAAATAT	211
MOH40	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAACAAAATAT	235
MOH42B	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAACAAAATAT	235
MOH39	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAACAAAATAT	235
ANJ39	----- ATGCATTGAAAGCTTCAAATATAAAGTTGATAAACTTTT 43	
ANJ17	----- GTGCATTGAAAGCTTCAAATATAAAGTTGATAAACTTTT 43	
ANJ37	----- ATGCATTGAAAGCTTCAAATATAAAGTTGATAAACTTTT 43	
ANJ10	----- ATIGATTAAAGCTTCAAATATAAAGTGGATAAATATT 41	
MOH42A	----- TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAAT-----AAT 220	
MOH14	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAACAAAATAT	220
MOH27	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAACAAAATAT	225
MOH7	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAACAAAATAT	225
NGA8B	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAACAAAATAT	270
ANJ36	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAACAAAATAT	201
MOH23C	TGCTGATAATTAAATTGCAATTCCAGAATAATAAATT---ATTAAATAATTCAACAT	189
MOH8	--TAAAGATTTAATCTTTTATT-AAAATTAAATTAAATAAAAACAAAATA	192
MOH17B	----- 73	
MOH16B	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAAT-AAAACAAAATA	187
MOH11A	----- TTGGCAGAATAAGTCATTGAAAGCTTCAAATATAAAGTTGATAAACTTT 54	
NGA7B	----- AGTCATTGAAAGCTTCAAATATAAAGTTGATAAACTTTTATTAAAATTAT	56
NGA2A	----- GGTGCATTGAAAGCTTCAAACATTAAGTTGATAAACTTTTATTAAAATTAT	56
NGA2B	----- CCT-GGAAATAACATTCAAAATAAAGTTGATAAACTTT 38	
ANJ20B	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAAT-AAAACAAAATA	188
MOH25A_(reversed)	--AAGTCATTGAAAGCTTCAAATATAAAGTTGATAAACTTTTATTAAAATTAT	66
NGA28A	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAAT-AAAACAAAATA	274
MOH9	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAAT-AAAACAAAATA	274
NGA21	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAAT-AAAACAAAATA	274
NGA11C	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAAT-AAAACAAAATA	274
NGA3	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAAT-AAAACAAAATA	274
ANJ29	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAAT-AAAACAAAATA	199
KT828418.1_(A1_MAD3)	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAAT-AAAACAAAATA	268
KT828427.1_(A1_MAD12)	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAAT-AAAACAAAATA	268
NGA14C	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAAT-AAAACAAAATA	262
NGA12	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAAT-AAAACAAAATAT	259
NGA5A	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAAT-AAAACAAAATAT	277
NGA8A	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAAT-AAAACAAAATAT	277
ANJ12	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAAT-AAAACAAAATAT	277
MOH11B	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAAT-AAAACAAAATAT	277
NGA4	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAAT-AAAACAAAATAT	278
MOH34	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAAT-AAAACAAAATAT	278
NGA15B	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAAT-AAAACAAAATAT	278
MOH19	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAAT-AAAACAAAATAT	278
MOH23B	----- ATAGTCATTGAAAGCTTCAAATATAAAGTTGATAAACTTT 46	
MOH36A	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAAT-AAAACAAAATAT	260
ANJ2	----- 0	
ANJ27	----- 59	
MOH37	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAAT-AAAACAAAATAT	239
MOH26	--TAAAGATTTAATCTTTTATTAAAATTAAATTAAATAAAAAT-AAAACAAAATA	239
AJ20C	----- AGAATAGTCATTGAAAGCTTCAAATATAAAGTTGATAAACTTT 49	



MOH16B	TAACAGAAATATATTAAAATTTAATTAACTTAAATTTCCACATGATTATTTATAT	247
MOH11A	TATTTAAAATTAATTTATAAATTTTATTAAATTTCTACATGATTATTTACAT	114
NGA7B	TAATTAACCTAATATTATAAATTTTATTAAATTTCTACATGATTATTTACAT	116
NGA2A	TAATTAACCTAATATTATAAATTTTATTAAATTTCTACATGATTATTTACAT	116
NGA2B	TATTTAAAATTAATATGATAAATTTTATTAAATTTCTACATGATTATTTACAT	98
ANJ20B	TAACAGAAATATATTATAAATTTTATTAAATTTCTACATGATTATTTACAT	248
MOH25A_(reversed)	TAATTAACCTAATATTATAAATTTTATTAAATTTCTACATGATTATTTACAT	126
NGA28A	TAACAGAAATATATTATAAATTTTATTAAATTTCTACATGATTATTTACAT	334
MOH9	TAACAGAAATATATTATAAATTTTATTAAATTTCTACATGATTATTTACAT	334
NGA21	TAACAGAAATATATTATAAATTTTATTAAATTTCTACATGATTATTTACAT	334
NGA11C	TAACAGAAATATATTATAAATTTTATTAAATTTCTACATGATTATTTACAT	334
NGA3	TAACAGAAATATATTATAAATTTTATTAAATTTCTACATGATTATTTACAT	334
ANJ29	TAACAGAAATATATTATAAATTTTATTAAATTTCTACATGATTATTTACAT	259
KT828418.1_(A1_MAD3)	TAACAGAAATATATTATAAATTTTATTAAATTTCTACATGATTATTTACAT	328
KT828427.1_(A1_MAD12)	TAACAGAAATATATTATAAATTTTATTAAATTTCTACATGATTATTTACAT	328
NGA14C	TAACAGAAATATATTATAAATTTTATTAAATTTCTACATGATTATTTACAT	322
NGA12	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	318
NGA5A	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	336
NGA8A	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	336
ANJ12	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	336
MOH11B	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	336
NGA4	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	337
MOH34	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	337
NGA15B	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	337
MOH19	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	337
MOH23B	ATTTAAATTA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	105
MOH36A	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	319
ANJ2	-----TTAGTAAAATTTCTACATGATTATTTACAT34	
ANJ27	-----ATATTATTATAAATTTCTACATGATTATTTACAT	99
MOH37	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	298
MOH26	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	298
AJ20C	ATTA-AAATTAATATTATAAATTTCTACATGATTATTTACAT	108
MOH18	ATTA-AAATTAATATTATAAATAGCATTATTAAATTTCTACATGATTATTTACAT	108
ANJ06	-----TAATTTATTAAATTTCTACATGATTATTTACAT	37
ANJ5	-----TTATTTGTAAAATTTCTACATGATTATTTACAT	37
ANJ7B	-----TTATTTATTAAATTTCTACATGATTATTTACAT	37
ANJ1	-----TTATTTATTAAATTTATACATGATTATTTACAT	37
MOH4	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	276
NGA19	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	345
MOH15A	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	344
NGA13	ATT-AAAATTAATATTATAAATATTAAATTTCTACATGATTATTTACAT	97
NC_001566.1_(RefSeq)	AACAGAAATATTTATTAAATTTAATTAAATTTCTACATGATTATTTACAT	3642
MOH30	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	314
NGA25_(reversed)	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	326
MOH36B	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	301
ANJ18B	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	296
ANJ8B	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	296
NGA34	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	296
MOH3	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	296
ANJ15A	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	296
MOH32	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	296
ANJ26A	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	296
ANJ22	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	296
ANJ25	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	296
ANJ28	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	296
ANJ23	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	217
ANJ19B	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	295
NGA17	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	295
NGA23	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	295
MOH25B	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	295
NGA22	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	295
NGA26A_(reversed)	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	295
ANJ33	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	295
MOH24	ATTTAAATTA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	88
ANJ35A_2	ATTTAAATTA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	101
ANJ35B	ATTTAAATTA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	101
ANJ34	ATTTAAATTA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	101
ANJ31	ATTTAAATTA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	101
ANJ30	ATTTAAATTA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	101
Anj21	ATTTAAATTA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	101
MOH28A	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	332
NGA11B	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	332
ANJ18A	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	332
MOH10	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	332
NGA26B_(reversed)	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	333
NGA16	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	333
MOH12C	AACAGAAATATA-TTATTAATTTTATTAAATTTCTACATGATTATTTACAT	264
	* *** * *** *	
NGA9	TTCAAGAACAAATTCTATATTATGCTGATAATTAAATTCTCATATAAGTTATAA	395

NGA6	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	395
NGA14A	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	395
NGA7A	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	395
NGA11A	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	395
NGA15	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	395
NGA14B	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	395
NGA20	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	395
NGA10	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	395
NGA5B	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	395
ANJ35A	TTCAACAACTCAAATTCTACTATGCTGATAACTTAATTTCATTCTATAATATAGTTATAA	118
ANJ126B	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	316
NGA7C	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	403
NGA31	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	385
MOH29	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	384
NGA1	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	384
MOH15B	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	330
MOH40	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	354
MOH42B	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	354
MOH39	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	354
ANJ39	TTCAACAATCAAATTCTACTATGCTGATAACTTAATTTCATTCCAG----AATAATAA	157
ANJ17	TTCAACAATCAAATTCTACTATGCTGATAACTTAATTTCATTCCAG----AATAATAA	157
ANJ37	TTCAACAATCAAATTCTACTATGCTGATAACTTAATTTCATTCCAG----AATAATAA	157
ANJ10	TTCAACAATCAAATTCTACTATGCTGATAACTTAATTTCATTCCAG----AATAATAA	155
MOH42A	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	339
MOH14	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	339
MOH27	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	344
MOH7	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	344
NGA8B	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	389
ANJ36	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	320
MOH23C	-----AAAATTAAACACAGAACATTCTATTATGCTT	286
M0H8	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	312
MOH17B	TTCAA----CAATCATATAATTCAATAACTTAATTTCATTCTGATAATATAGTTATAA	166
MOH16B	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	307
MOH11A	TTCGACAATCAAATTCTACTATGCTGATAACTTAATTGCATTC----CAGAATAATAA	169
NGA7B	TTCAACAACTCATATAATTCAATAATTAA---TTTCATTCCATA---ATATAGAATAA	169
NGA2A	TTCAACAACTCATATAATTCAATAATTAA---TTTCATTCCATA---ATATAGAATAA	169
NGA2B	TTCAACAACTCAAATTCTACAGATGCTGATAACTTAATTTCATTCC----AGAATAATAA	153
ANJ20B	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	308
MOH25A_(reversed)	TTCAACAACTCATAT----AATTCAATAATTAACTTCATTCTATA-ATATAGAATAA	179
NGA28A	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	394
MOH9	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	394
NGA21	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	394
NGA11C	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	394
NGA3	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	394
ANJ29	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	319
KT828418.1_(A1_MAD3)	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	388
KT828427.1_(A1_MAD12)	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	388
NGA14C	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	382
NGA12	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	378
NGA5A	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	396
NGA8A	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	396
ANJ12	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	396
MOH11B	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	396
NGA4	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	397
MOH34	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	397
NGA15B	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	397
MOH19	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	397
MOH23B	TTCGACAATCAAATTCTACTATGCTGATAACTTAATTTCATTCCAG----AATAATAA	160
MOH36A	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	379
ANJ2	TTCAACAACTCAGATTCTACGGTGCTGATAACT----TAATTTCATTCCAGAATAATAA	89
ANJ27	TTCAACAACTCAAATTCTACTATGCTGATAACTTAATTTCATTCCAG----AATAATAA	154
MOH37	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	358
MOH26	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	358
AJ20C	TTCAACAACTCAAATTCTACTATGCTGATAACTTAATTTCATTCCAGA----ATAATAA	163
MOH18	TTCAACAACTCAAATTCTACTATGCTGATAACTTAATTTCATTCCAGA----ATAATAA	163
ANJ06	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	97
ANJ5	TTGAACAACTCAAATTCTACTATGCTGATAACTTAATTTCATTCCAGA----TAATAG	92
ANJ7B	TTCAACAACTCAAATTCTACTATGCTGATAACTTAATTTCATTCCAGA----TAATAA	92
ANJ1	TTCAACAACTCAAATTCTATTATGCTGATAACTTAATTTCATTCCAGA----TAATAA	92
MOH4	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	336
NGA19	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	405
MOH15A	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	404
NGA13	TTCAACAACTCAAATTCTACTATGCTGAT----AACTTAATTTCATTCCAGAATAATAA	152
NC_001566.1_(RefSeq)	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	3702
MOH30	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	374
NGA25_(reversed)	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	386
MOH36B	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	361
ANJ18B	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	356
ANJ8B	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	356
NGA34	TTCAAGAACATCAAATTCTATTATGCTGATAATTAACTTCATTCTATAATATAGTTATAA	356

MOH3	TTCAAGAACATCAAATTCTATTGCTGATAATTAAATTCTATTCTATAATATAGTTATAA	356
ANJ15A	TTCAAGAACATCAAATTCTATTGCTGATAATTAAATTCTATTCTATAATATAGTTATAA	356
MOH32	TTCAAGAACATCAAATTCTATTGCTGATAATTAAATTCTATTCTATAATATAGTTATAA	356
ANJ26A	TTCAAGAACATCAAATTCTATTGCTGATAATTAAATTCTATTCTATAATATAGTTATAA	356
ANJ22	TTCAAGAACATCAAATTCTATTGCTGATAATTAAATTCTATTCTATAATATAGTTATAA	356
ANJ25	TTCAAGAACATCAAATTCTATTGCTGATAATTAAATTCTATTCTATAATATAGTTATAA	356
ANJ28	TTCAAGAACATCAAATTCTATTGCTGATAATTAAATTCTATTCTATAATATAGTTATAA	356
ANJ23	TTCAAGAACATCAAATTCTATTGCTGATAATTAAATTCTATTCTATAATATAGTTATAA	277
ANJ19B	TTCAAGAACATCAAATTCTATTGCTGATAATTAAATTCTATTCTATAATATAGTTATAA	355
NGA17	TTCAAGAACATCAAATTCTATTGCTGATAATTAAATTCTATTCTATAATATAGTTATAA	355
NGA23	TTCAAGAACATCAAATTCTATTGCTGATAATTAAATTCTATTCTATAATATAGTTATAA	355
MOH25B	TTCAAGAACATCAAATTCTATTGCTGATAATTAAATTCTATTCTATAATATAGTTATAA	355
NGA22	TTCAAGAACATCAAATTCTATTGCTGATAATTAAATTCTATTCTATAATATAGTTATAA	355
NGA26A_(reversed)	TTCAAGAACATCAAATTCTATTGCTGATAATTAAATTCTATTCTATAATATAGTTATAA	355
ANJ33	TTCAAGAACATCAAATTCTATTGCTGATAATTAAATTCTATTCTATAATATAGTTATAA	355
MOH24	TTCAACAATCATATAATTCAAT-AACCTT---AACCTTCATTCCATAATATAGTAATAA	142
ANJ35A_2	TTCAACAATCAAATTCTACTATGCTGAT----AACCTTAATTTCATTCCAGAATAATAA	156
ANJ35B	TTCAACAATCAAATTCTACTATGCTGAT----AACCTTAATTTCATTCCAGAATAATAA	156
ANJ34	TTCAACAATCAAATTCTACTATGCTGAT----AACCTTAATTTCATTCCAGAATAATAA	156
ANJ31	TTCAACAATCAAATTCTACTATGCTGAT----AACCTTAATTTCATTCCAGAATAATAA	156
ANJ30	TTCAACAATCAAATTCTACTATGCTGAT----AACCTTAATTTCATTCCAGAATAATAA	156
Anj21	TTCAACAATCAAATTCTACTATGCTGAT----AACCTTAATTTCATTCCAGAATAATAA	156
MOH28A	TTCAAGAACATCAAATTCTATTGCTGATAATTAAATTCTATTCTATAATATAGTTATAA	392
NGA11B	TTCAAGAACATCAAATTCTATTGCTGATAATTAAATTCTATTCTATAATATAGTTATAA	392
ANJ18A	TTCAAGAACATCAAATTCTATTGCTGATAATTAAATTCTATTCTATAATATAGTTATAA	392
MOH10	TTCAAGAACATCAAATTCTATTGCTGATAATTAAATTCTATTCTATAATATAGTTATAA	392
NGA26B_(reversed)	TTCAAGAACATCAAATTCTATTGCTGATAATTAAATTCTATTCTATAATATAGTTATAA	393
NGA16	TTCAAGAACATCAAATTCTATTGCTGATAATTAAATTCTATTCTATAATATAGTTATAA	393
MOH12C	TTCAAGAACATCAAATTCTATTGCTGATAATTAAATTCTATTCTATAATATAGTTATAA	324
	*	
NGA9	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	455
NGA6	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	455
NGA14A	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	455
NGA7A	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	455
NGA11A	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	455
NGA15	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	455
NGA14B	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	455
NGA20	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	455
NGA10	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	455
NGA5B	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	455
ANJ35A	TAATTATTATTATAATTCAACATTAACAGTTCATATTATTTAGATTTATTCTATAAATA	178
ANJ26B	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	376
NGA7C	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	463
NGA31	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	445
MOH29	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	444
NGA1	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	444
MOH15B	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	390
MOH40	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	414
MOH42B	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	414
MOH39	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	414
ANJ39	TAATTATTATTATAATTCAACATTAACAGTCATATTATTTAGATTTATTCTATAAATA	217
ANJ17	TAATTATTATTATAATTCAACATTAACAGTCATATTATTTAGATTTATTCTATAAATA	217
ANJ37	TAATTATTATTATAATTCAACATTAACAGTCATATTATTTAGATTTATTCTATAAATA	217
ANJ10	TAATTATTATTATAATTCAACATTAACAGTCATATTATTTAGATTTATTCTATAAATA	215
MOH42A	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	399
MOH14	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	399
MOH27	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	404
MOH7	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	404
NGA8B	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	449
ANJ36	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	380
MOH23C	CAATTATTCTTCTTCACTT-----TTAAAAATTCTATTCTATAATTGAAGATGA	337
M0H8	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	372
MOH17B	TAATCATTATTATAATTCAACATTAACAGTTATCTATTCTTAGATTTATTGTAAATA	226
MOH16B	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	367
MOH11A	TAATTATTATTATAATTCAACATTAACAGTTATCTATTCTTAGATTTATTGTAAATA	229
NGA7B	TAATCATTATTATAATTCAACATTAACAGTTATCTATTCTTAGATTTATTGTAAATA	229
NGA2A	TAATCATTATTATAATTCAACATTAACAGTTATCTATTCTTAGATTTATTGTAAATA	229
NGA2B	TAATCATTATTATAATTCAACATTAACAGTTATCTATTCTTAGATTTATTGTAAATA	213
ANJ20B	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	368
MOH25A_(reversed)	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTGTAAATA	239
NGA28A	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	454
MOH9	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	454
NGA21	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	454
NGA11C	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	454
NGA3	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	454
ANJ29	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	379
KT828418.1_(A1_MAD3)	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	448
KT828427.1_(A1_MAD12)	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	442
NGA14C	TAATCATTATTATAATTCAACATTAACGTGATATATTATTTAGATTTATTCTATAAATA	442

NGA12	TAATCATTATTATAATTCAACATTAACGTATATATTAGTTTAAATA	438
NGA5A	TAATCATTATTATAATTCAACATTAACGTATATATTAGTTTAAATA	456
NGA8A	TAATCATTATTATAATTCAACATTAACGTATATATTAGTTTAAATA	456
ANJ12	TAATCATTATTATAATTCAACATTAACGTATATATTAGTTTAAATA	456
MOH11B	TAATCATTATTATAATTCAACATTAACGTATATATTAGTTTAAATA	456
NGA4	TAATCATTATTATAATTCAACATTAACGTATATATTAGTTTAAATA	457
MOH34	TAATCATTATTATAATTCAACATTAACGTATATATTAGTTTAAATA	457
NGA15B	TAATCATTATTATAATTCAACATTAACGTATATATTAGTTTAAATA	457
MOH19	TAATCATTATTATAATTCAACATTAACGTATATATTAGTTTAAATA	457
MOH23B	TAATCATTATTATAATTCAACATTAACGTATATATTAGTTTAAATA	220
MOH36A	TAATCATTATTATAATTCAACATTAACGTATATATTAGTTTAAATA	439
ANJ2	TAATTATTATAATTCAACATTAACAGTCATATTAGTTTAAATA	149
ANJ27	TAATTATTATAATTCAACATTAACAGTCATATTAGTTTAAATA	214
MOH37	TAATCATTATTATAATTCAACATTAACGTATATATTAGTTTAAATA	418
MOH26	TAATCATTATTATAATTCAACATTAACGTATATATTAGTTTAAATA	418
AJ20C	TAATTATTATAATTCAACATTAACAGTCATATTAGTTTAAATA	223
MOH18	TAATTATTATAATTCAACATTAACAGTCATATTAGTTTAAATA	223
ANJ06	TAATCATTATTATAATTCAACATTAACGTATATATTAGTTTAAATA	157
ANJ5	TAATTATTATAATTCAACATTAACAGTCATATTAGTTTAAATA	152
ANJ7B	TAATTATTATAATTCAACATTAACAGTCATATTAGTTTAAATA	152
ANJ1	TAATTATTATAATTCAACATTAACAGTCATATTAGTTTAAATA	152
MOH4	TAATCATTATTATAATTCAACATTAACGTATATATTAGTTTAAATA	396
NGA19	TAATCATTATTATAATTCAACATTAACAGTCATATTAGTTTAAATA	465
MOH15A	TAATCATTATTATAATTCAACATTAACGTATATATTAGTTTAAATA	464
NGA13	TAATTATTATAATTCAACATTAACAGTCATATTAGTTTAAATA	212
NC_001566.1_(RefSeq)	TAATTATTATAATTCAACATTAACGTATATATTAGTTTAAACA	3762
MOH30	TAATCATTATTATAATTCAACATTAACGTATATATTAGTTTAAATA	434
NGA25_(reversed)	TAATCATTATTATAATTCAACATTAACGTATATTAGTTTAAATA	446
MOH36B	TAATCATTATTATAATTCAACATTAACGTATATTAGTTTAAATA	421
ANJ18B	TAATCATTATTATAATTCAACATTAACGTATATTAGTTTAAATA	416
ANJ8B	TAATCATTATTATAATTCAACATTAACGTATATTAGTTTAAATA	416
NGA34	TAATCATTATTATAATTCAACATTAACGTATATTAGTTTAAATA	416
MOH3	TAATCATTATTATAATTCAACATTAACGTATATTAGTTTAAATA	416
ANJ15A	TAATCATTATTATAATTCAACATTAACGTATATTAGTTTAAATA	416
MOH32	TAATCATTATTATAATTCAACATTAACGTATATTAGTTTAAATA	416
ANJ26A	TAATCATTATTATAATTCAACATTAACGTATATTAGTTTAAATA	416
ANJ22	TAATCATTATTATAATTCAACATTAACGTATATTAGTTTAAATA	416
ANJ25	TAATCATTATTATAATTCAACATTAACGTATATTAGTTTAAATA	416
ANJ28	TAATCATTATTATAATTCAACATTAACGTATATTAGTTTAAATA	416
ANJ23	TAATCATTATTATAATTCAACATTAACGTATATTAGTTTAAATA	337
ANJ19B	TAATCATTATTATAATTCAACATTAACGTATATTAGTTTAAATA	415
NGA17	TAATCATTATTATAATTCAACATTAACGTATATTAGTTTAAATA	415
NGA23	TAATCATTATTATAATTCAACATTAACGTATATTAGTTTAAATA	415
MOH25B	TAATCATTATTATAATTCAACATTAACGTATATTAGTTTAAATA	415
NGA22	TAATCATTATTATAATTCAACATTAACGTATATTAGTTTAAATA	415
NGA26A_(reversed)	TAATCATTATTATAATTCAACATTAACGTATATTAGTTTAAATA	415
ANJ33	TAATCATTATTATAATTCAACATTAACGTATATTAGTTTAAATA	415
MOH24	TAATTATTATAATTCAACATTAACAGTCATATTAGTTTAAATA	202
ANJ35A_2	TAATTATTATAATTCAACATTAACAGTCATATTAGTTTAAATA	216
ANJ35B	TAATTATTATAATTCAACATTAACAGTCATATTAGTTTAAATA	216
ANJ34	TAATTATTATAATTCAACATTAACAGTCATATTAGTTTAAATA	216
ANJ31	TAATTATTATAATTCAACATTAACAGTCATATTAGTTTAAATA	216
ANJ30	TAATTATTATAATTCAACATTAACAGTCATATTAGTTTAAATA	216
Anj21	TAATTATTATAATTCAACATTAACAGTCATATTAGTTTAAATA	216
MOH28A	TAATCATTATTATAATTCAACATTAACGTATATTAGTTTAAATA	452
NGA11B	TAATCATTATTATAATTCAACATTAACGTATATTAGTTTAAATA	452
ANJ18A	TAATCATTATTATAATTCAACATTAACGTATATTAGTTTAAATA	452
MOH10	TAATCATTATTATAATTCAACATTAACGTATATTAGTTTAAATA	452
NGA26B_(reversed)	TAATCATTATTATAATTCAACATTAACGTATATTAGTTTAAATA	453
NGA16	TAATCATTATTATAATTCAACATTAACGTATATTAGTTTAAATA	453
MOH12C	TAATCATTATTATAATTCAACATTAACGTATATTAGTTTAAATA	384
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NGA9	AATTTCAAAATTATTTT-ATTAaaaaaaatcataa-TATTGAAATTATTGAACAGTTAT	513
NGA6	AATTTCAAAATTATTTT-ATTAaaaaaaatcataa-TATTGAAATTATTGAACAGTTAT	513
NGA14A	AATTTCAAAATTATTTT-ATTAaaaaaaatcataa-TATTGAAATTATTGAACAGTTAT	513
NGA7A	AATTTCAAAATTATTTT-ATTAaaaaaaatcataa-TATTGAAATTATTGAACAGTTAT	513
NGA11A	AATTTCAAAATTATTTT-ATTAaaaaaaatcataa-TATTGAAATTATTGAACAGTTAT	513
NGA15	AATTTCAAAATTATTTT-ATTAaaaaaaatcataa-TATTGAAATTATTGAACAGTTAT	513
NGA14B	AATTTCAAAATTATTTT-ATTAaaaaaaatcataa-TATTGAAATTATTGAACAGTTAT	513
NGA20	AATTTCAAAATTATTTT-ATTAaaaaaaatcataa-TATTGAAATTATTGAACAGTTAT	513
NGA10	AATTTCAAAATTATTTT-ATTAaaaaaaatcataa-TATTGAAATTATTGAACAGTTAT	513
NGA5B	AATTTCAAAATTATTTT-ATTAaaaaaaatcataa-TATTGAAATTATTGAACAGTTAT	513
ANJ35A	AATTTCAAAATTATTTT-ATTAaaaaaaatcataa-----TTAACAGTAAT 222	
ANJ26B	AATTTCAAAATTATTTT-ATTAaaaaaaatcataa-TATTGAAATTATTGAACAGTTAT	434
NGA7C	AATTTCAAAATTATTTT-ATTAaaaaaaatcataa-TATTGAAATTATTGAACAGTTAT	521
NGA31	AATTTCAAAATTATTTT-ATTAaaaaaaatcataa-TATTGAAATTATTGAACAGTTAT	503
MOH29	AATTTCAAAATTATTTT-ATTAaaaaaaatcataa-TATTGAAATTATTGAACAGTTAT	502
NGA1	AATTTCAAAATTATTTT-ATTAaaaaaaatcataa-TATTGAAATTATTGAACAGTTAT	502
MOH15B	AATTTCAAAATTATTTT-ATTAaaaaaaatcataa-TATTGAAATTATTGAACAGTTAT	448

MOH40	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	472
MOH42B	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	472
MOH39	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	472
ANJ39	AATTTCAAACTTATTATT-ATTAaaaaattat-----TTAACAGTAAT 261	
ANJ17	AATTTCAAACTTATTATT-ATTAaaaaattat-----TTAACAGTAAT 261	
ANJ37	AATTTCAAACTTATTATT-ATTAaaaaattat-----TTAACAGTAAT 261	
ANJ10	AATTTCAAACTTATTATT-ATTAaaaaattat-----TTAACAGTAAT 259	
MOH42A	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	457
MOH14	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	457
MOH27	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	462
MOH7	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	462
NGA8B	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	507
ANJ36	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	438
MOH23C	A-ATTGCAATCCATTCTTCATTGGTCATCAATTGATGG-----	387
MOH8	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	430
MOH17B	AATTTCAAACTTATTATT-ATTAaaaaatcatga-TATTGAAATTATTTGAACAGTTAT	284
MOH16B	AATTTCAAACTTATTATT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	425
MOH11A	AATTTCAAACTTATTATT-ATTA-----AAAAAATTAAACAGTAAT 273	
NGA7B	AATTTCAAACTTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	287
NGA2A	AATTTCAAACTTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	287
NGA2B	AATTTCAAACTTATTATT-ATTAaaaaa-----ATTATTAAACAGTAAT 257	
ANJ20B	AATTTCAAACTTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	426
MOH25A_(reversed)	AATTTCAAACTTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	297
NGA28A	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	512
MOH9	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	512
NGA21	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	512
NGA11C	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	512
NGA3	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	512
ANJ29	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	437
KT828418.1_(A1_MAD3)	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	506
KT828427.1_(A1_MAD12)	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	506
NGA14C	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	500
NGA12	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	496
NGA5A	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	514
NGA8A	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	514
ANJ12	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	514
MOH11B	AATTTCAAAATTATTTT-ATTAaaaaatctaa-TATTGAAATTATTTGAACAGTTAT	514
NGA4	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	515
MOH34	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	515
NGA15B	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	515
MOH19	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	515
MOH23B	AATTTCAAACTTATTATT-ATTAaaaaa-----TATTAAACAGTAAT 264	
MOH36A	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	497
ANJ2	AATTTCAAACTTATTATT-ATTAaaaaa-----TATTAAACAGTAAT 193	
ANJ27	AATTTCAAACTTATTATT-ATTAaaaaa-----TTAACAGTAAT 258	
MOH37	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	476
MOH26	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	476
AJ20C	AATTTCAAACTTATTATT-ATTAaaaaa-----TTAACAGTAAT 267	
MOH18	AATTTCAAACTTATTATT-ATTAaaaaa-----TTAACAGTAAT 267	
ANJ06	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	215
ANJ5	AATTTCAAACTTATTATT-ATTAaaaaa-----TTAACAGTAAT 196	
ANJ7B	AATTTCAAACTTATTATT-ATTAaaaaa-----TTAACAGTAAT 196	
ANJ1	AATTTCAAACTTATTATT-ATTAaaaaa-----TTAACAGTAAT 196	
MOH4	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	454
NGA19	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	523
MOH15A	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	522
NGA13	AATTTCAAACTTATTATT-ATTAaaaaa-----TTAACAGTAAT 256	
NC_001566.1_(RefSeq)	AATTC TCAA ATT TTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACATTAT	3820
MOH30	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	492
NGA25_(reversed)	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	504
MOH36B	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	479
ANJ18B	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	474
ANJ8B	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	474
NGA34	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	474
MOH3	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	474
ANJ15A	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	474
MOH32	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	474
ANJ26A	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	474
ANJ22	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	474
ANJ25	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	474
ANJ28	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	474
ANJ23	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	395
ANJ19B	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	473
NGA17	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	473
NGA23	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	473
MOH25B	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	473
NGA22	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	473
NGA26A_(reversed)	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	473
ANJ33	AATTTCAAAATTATTTT-ATTAaaaaatcataa-TATTGAAATTATTTGAACAGTTAT	473
MOH24	AATTTCAAACTTATTATT-ATTAaaaaatcatga-TATTGAAATTATTTGAACAGTAAT	260

ANJ35A_2	AATTTCAAACTTATTATT-ATTAaaaaAATT-----	ATTTAACAGTAAT	260
ANJ35B	AATTTCAAACTTATTATT-ATTAaaaaAATT-----	ATTTAACAGTAAT	260
ANJ34	AATTTCAAACTTATTATT-ATTAaaaaAATT-----	ATTTAACAGTAAT	260
ANJ31	AATTTCAAACTTATTATT-ATTAaaaaAATT-----	ATTTAACAGTAAT	260
ANJ30	AATTTCAAACTTATTATT-ATTAaaaaAATT-----	ATTTAACAGTAAT	260
Anj21	AATTTCAAACTTATTATT-ATTAaaaaAATT-----	ATTTAACAGTAAT	260
MOH28A	AATTTCAAACTTTT-ATTAaaaATCATAA-TATTGAAATTATTGAACAGTTAT		510
NGA11B	AATTTCAAACTTTT-ATTAaaaATCATAA-TATTGAAATTATTGAACAGTTAT		510
ANJ18A	AATTTCAAACTTTT-ATTAaaaATCATAA-TATTGAAATTATTGAACAGTTAT		510
MOH10	AATTTCAAACTTTT-ATTAaaaATCATAA-TATTGAAATTATTGAACAGTTAT		510
NGA26B_(reversed)	AATTTCAAACTTTT-ATTAaaaATCATAA-TATTGAAATTATTGAACAGTTAT		511
NGA16	AATTTCAAACTTTT-ATTAaaaATCATAA-TATTGAAATTATTGAACAGTTAT		511
MOH12C	AATTTCAAACTTTT-ATTAaaaATCATAA-TATTGAAATTATTGAACAGTTAT		442
* * *** * *			
NGA9	TCCAATTATTCTATAATTATTGTTCCATCATAAAAATTATTAATTG		573
NGA6	TCCAATTATTCTATAATTATTGTTCCATCATAAAAATTATTAATTG		573
NGA14A	TCCAATTATTCTATAATTATTGTTCCATCATAAAAATTATTAATTG		573
NGA7A	TCCAATTATTCTATAATTATTGTTCCATCATAAAAATTATTAATTG		573
NGA11A	TCCAATTATTCTATAATTATTGTTCCATCATAAAAATTATTAATTG		573
NGA15	TCCAATTATTCTATAATTATTGTTCCATCATAAAAATTATTAATTG		573
NGA14B	TCCAATTATTCTATAATTATTGTTCCATCATAAAAATTATTAATTG		573
NGA20	TCCAATTATTCTATAATTATTGTTCCATCATAAAAATTATTAATTG		573
NGA10	TCCAATTATTCTATAATTATTGTTCCATCATAAAAATTATTAATTG		573
NGA5B	TCCAATTATTCTATAATTATTGTTCCATCATAAAAATTATTAATTG		573
ANJ35A	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		282
ANJ26B	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		491
NGA7C	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		578
NGA31	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		560
MOH29	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		559
NGA1	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		559
MOH15B	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		505
MOH40	TCCCATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		529
MOH42B	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		529
MOH39	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		529
ANJ39	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		321
ANJ17	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		321
ANJ37	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		321
ANJ10	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		319
MOH42A	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		514
MOH14	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		514
MOH27	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		519
MOH7	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		519
NGA8B	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		564
ANJ36	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		495
MOH23C	-----	387	
MOH8	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		490
MOH17B	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		344
MOH16B	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		480
MOH11A	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		333
NGA7B	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		347
NGA2A	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		347
NGA2B	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		317
ANJ20B	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		486
MOH25A_(reversed)	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		357
NGA28A	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		572
MOH9	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		572
NGA21	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		572
NGA11C	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		572
NGA3	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		572
ANJ29	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		497
KT828418.1_(A1_MAD3)	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		545
KT828427.1_(A1_MAD12)	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		545
NGA14C	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		560
NGA12	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		553
NGA5A	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		571
NGA8A	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		571
ANJ12	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		571
MOH11B	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT	529	
NGA4	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		572
MOH34	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		572
NGA15B	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		572
MOH19	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		572
MOH23B	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		324
MOH36A	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		554
ANJ2	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		253
ANJ27	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		318
MOH37	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		533
MOH26	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		533
AJ20C	TCCAATTATTCTTCATAATTCTTCCACTTTAAAATTATTAATT		327

MOH18	TCCAATTATTATTCTTCAATTATTTCTTCCCTCACCTTAAAGAATTTATTTAAT	327
ANJ06	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAA---AAATTITATATTTAAT	272
ANJ5	TCCAATTATTATTCTTCAATTATTCCTTCCACTTTAAAATTTATTTAAT	256
ANJ7B	TCCAATTATTATTCTTCAATTATTCCTTCCACTTTAAAATTTATTTAAT	256
ANJ1	TCCAATTATTATTCTTCAATTATTCCTTCCACTTTAAAATTTATTTAAT	256
MOH4	TCCAATTATTATTCTTCAATTATTCCTTCCACTTTAAAATTTATTTAAT	514
NGA19	TCCAATTATTATTCTTCAATTATTCCTTCCACTTTAAAATTTATTTAAT	583
MOH15A	TCCAATTATTATTCTTCAATTATTCCTTCCACTTTAAAATTTATTTAAT	582
NGA13	TCCAATTATTATTCTTCAATTATTCCTTCCACTTTAAAATTTATTTAAT	316
NC_001566.1_(RefSeq)	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	3880
MOH30	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	552
NGA25_(reversed)	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	564
MOH36B	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	539
ANJ18B	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	534
ANJ8B	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	534
NGA34	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	534
MOH3	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	534
ANJ15A	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	534
MOH32	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	534
ANJ26A	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	534
ANJ22	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	534
ANJ25	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	534
ANJ28	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	534
ANJ23	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	455
ANJ19B	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	533
NGA17	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	533
NGA23	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	533
MOH25B	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	533
NGA22	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	533
NGA26A_(reversed)	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	533
ANJ33	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	533
MOH24	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	316
ANJ35A_2	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	320
ANJ35B	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	320
ANJ34	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	320
ANJ31	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	320
ANJ30	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	320
Anj21	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	320
MOH28A	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	570
NGA11B	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	570
ANJ18A	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	570
MOH10	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	570
NGA26B_(reversed)	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	571
NGA16	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	571
MOH12C	TCCAATTATTATTCTTCAATTATTTGTTTCCATCATAAAATTTATTTAAT	502

NGA9	TGA----AATTGTAAATCCTTTTTCAATTAAATCAATTGGTCAC-----	616
NGA6	ATGA---AATTGTAAATCCTTTTTCAATTAAATCAATTGGTCATC-----	618
NGA14A	TG-A---AATTGTAAATCCTTTTTCAATTAAATCAATTGGTCATCAA-----	619
NGA7A	TG-A---AATTGTAAATCCTTTTTCAATTAAATCAATTGGTC-----	616
NGA11A	TG-A---AATTGTAAATCCTTTTTCAATTAAATCAATTGGTC-----	616
NGA15	TG-A---AATTGTAAATCCTTTTTCAATTAAATCAATTGGTCAC-----	616
NGA14B	TG-A---AATTGTAAATCCTTTTTCAATTAAATCAATTGGTC-----	614
NGA20	TG-A---AATTGTAAATCCTTTTTCAATTAAATCAATTGGTC-----	614
NGA10	TG-A---AATTGTAAATCCTTTTTCAATTAAATCAATTGGTCACATGAATGGGA	628
NGA5B	TG-A---AATTGTAAATCCTTTTTCAATTAAATCAATTGGTCACAA-----	618
ANJ35A	TGAAATGAAATTGCAATCCATTCTT-----CTATCTATTGGTCAT-----	325
ANJ26B	TGA-TGAAATTGTAATCCTTTTTCAATTAAATCAATTGGTC-----	536
NGA7C	TTA-TGAAATTGTAATCCTTTTTCAATTAAATCAATTGGTCAT-----	625
NGA31	TTA-TAAATTT-AAATCCTTTTTCAATTAAATCAATTGGTCATCAATGAATTT-	615
MOH29	TGA--TGAAATTGTAATCCTTTTTCAATTAAATCAATTGGTCATCG-----	607
NGA1	TGA--TGAAATTGTAATCCTTTTTTC-----586	
MOH15B	TGA--TGAAATTGTAATCCTTTTTCAATTAAATCAATTGGTCATCAATG-----	556
MOH40	TGA--TGAAA-----538	
MOH42B	TGA--TGAAATTGTAATCCTTTTTCAATTAAATCAATTGGTCATAA-----	577
MOH39	TG-----531	
ANJ39	TGAAATGAAATTGCAATCCATTCTT-----ATCTATTGGTCATAATGATATTG-	375
ANJ17	TGAAATGAAATTGCAATCCATTCTT-----ATCTATTGGTCATAATGATATTG-	375
ANJ37	TGAAATGAAATTGCAATCCATTCTT-----ATCTATTGGTCATA-----365	
ANJ10	TGAAATGAAATTGCAATCCATTCTT-----TATCTATTGGTCATAATGATATTGA	374
MOH42A	TGA--TGAAATTGTAATCCTTTTTCAATTAAATCAATTGGTCATC-----561	
MOH14	TGA--TGAAATTGTAATCCTTTTTCAATTAAATCAATTGGTCATC-----562	
MOH27	TGA--TGAAATTGTAATCCTTTTTCAATTAAATCAATTGGTCATC-----566	
MOH7	TGA--T-----523	
NGA8B	TGA--TGAAATTGTAATCCTTTTTCAATTAAATCAATTGGTCAGTGAATGGGG	622
ANJ36	TGA--TGAAATTGTAATCCTTTTTCAAT-----525	
MOH23C	-----387	
M0H8	TGAAATTGTAATCCTTTT-----TTTCAATTAAATCAATTGGTCATCAA-----536	
MOH17B	TGAAATTGTAATCATTCTT-----TATTAAATGCATTGGTCATCAATGATATTGA	404

MOH16B	ATTGATGAAATTGAAATCCTTTTCAATTAAATCAATTGGTCATCAATGA-----	534
MOH11A	TGAAATGAAATTGCAAATCC----ATTCTTCTATCTATTGGTCACAAATGATATT--	386
NGA7B	TGAAATGAAATTACAATCT----ATTCTTCTATCTATTGGTCAA-----	390
NGA2A	TGAAATGAAATTACAATCT----ATTCTTCTATCTATTGGTC-----	389
NGA2B	TGAAATGAAATTGCAAATCC----ATTCTTCTATCTATTGGTCAT-----	360
ANJ20B	TGAAATTGAAATCCTTTT---TTCAATTAAATCAATTGGTCACA-----	530
MOH25A_(reversed)	TGAAATGAAATTACAATCT----ATTCTTCTATCTATTGGTCATCAATGATATT--	410
NGA28A	TGAAATTGAAATCCTTTT---TTCAATTAAATCAATTGGTCATCAA-----	618
MOH9	TGAAATTGAAATCCTTTT---TTCAATTAAATCAATTGGTC-----	613
NGA21	TGAAATTGAAATCCTTTT---TTCAATTAAATCAATTGGT-----	612
NGA11C	TGAAATTGAAATCCTTTT---TTCAATTAAATCAATTGGC-----	613
NGA3	TGAAATTGAAATCCTTTT---TTCAATTAAATCAATTGGTCAC-----	615
ANJ29	TGAAATTGAAATCCTTTT---TTCAATTAAATCAATTGGTCATC-----	541
KT828418.1_(A1_MAD3)	-----	545
KT828427.1_(A1_MAD12)	-----	545
NGA14C	TGAAATTGAAATCCTTTT---TTCAATTAAATCAATT-----	597
NGA12	TGA-TGAAATTGAAATCCTTTTCAATTAAATCAATTGGTCACA-----	600
NGA5A	TGA-TGAAATTGAAATCCTTTTCAATTAAATCGATTGGTCATC-----	618
NGA8A	TGA-TGAAATTGAAATCCTTTTCAATTAAATCAATTGGTCAGTGAAT-----	624
ANJ12	TGA-TGAAATTGAAATCCTTTTCAATTAAATCAATTGGTCAC-----	617
MOH11B	-----	529
NGA4	TGA-TGAAATTGAAATCCTTTTCAATTAAATCAATTGGTCACAGT-----	621
MOH34	TGA-TGAAATTGAAATCCTTTTCAATTAAATCAATTGGTCACAA-----	620
NGA15B	TGA-TGAAATTGAAATCCTTTTCAATTAAATCAATTGGTCATC-----	619
MOH19	TGA-TGAAATTGAAATCCTTTTCAATTAAATCAATTAAAATCAAAA-----	616
MOH23B	TGAAATGAAATTGCAAATCCATTCTTCTATCTA---TTGGTCATCAATGATATTGA	379
MOH36A	TGA-TGAAATTGAAATCCTTTTCAATTAAATCAATTGGTCATC-----	602
ANJ2	TGAAATGAAATTGCAAATCCATTCTTCTATTGGT-----	292
ANJ27	TGAAATGAAATTGCAAATCCATTCTTCTATTGGT-----	361
MOH37	TGA-TGAAATTGAAATCCTTTTCAATTAAATCAATTGGTCATCAATG-----	584
MOH26	TGA-TGAAATTGAAATCCTTTTCAATTAAATCAATTGGTCAT-----	579
AJ20C	TGAAATGAAATTGCAAATCCATTCTTCTATC---TATTGGTCAT-----	370
MOH18	TGAAATGAAATTGCAAATCCATTCTTCTATC---TATTGGTCATCAATGATATTGA	382
ANJ06	TG-ATGAAATTGAAATCCTTTTCAATT-----AAATCAATTGGT-----	315
ANJ5	TGAAATGAAATTGCAAATCCATTCTTCTATC---TA---TTGGT-----	296
ANJ7B	TGAAATGAAATTGCAAATCCATTCTTCTATC---TA---TTGGT-----	296
ANJ1	TGAAATGAAATTGCAAATCCATTCTTCTATC---TA---TTGGT-----	296
MOH4	TGAAATTGAAATCCTTTTCAATTAAATC---AAT-----	550
NGA19	TGAAATTGAAATCCTTTTCAATTAAATC---AATTGGTC-----	624
MOH15A	TGAAATTGAAATCCTTTTCAATTAAATC---AATTGGTC-----	624
NGA13	TGAAATGAAATTGCAAATCCATTCTTCTATC---TATTGGTCATC-----	360
NC_001566.1_(RefSeq)	TGAAATTGAAATCCTTTTCAATTAAATC---AATTGGTCATCAATGATATTGA	3935
MOH30	TGAAATGAAATCCATTCTTCTATTAAATC-----	583
NGA25_(reversed)	TGAAATTGAAATCCATTCTTCTATTAAATC-----	589
MOH36B	TGAAATTGTATATCCTTTTCAATTAAATC---AATTGGTCATCAG-----	585
ANJ18B	TGAAATTGAAATCCTTTTCAATTAAATC---AATTGGTCATCAA-----	580
ANJ8B	TGAAATTGAAATCCTTTTCAATTAAATC---AATTGGTCATC-----	576
NGA34	TGAAATTGAAATCCTTTTCAATTAAATC---AATTGGTCATCAATGATATT--	587
MOH3	TGAAATTGAAATCCTTTTCAATTAAATC---AATTGGTCAT-----	577
ANJ15A	TGAAATTGAAATCCTTTTCAATTAAATC---AATTGGTCATCAAT-----	581
MOH32	TGAAATTGAAATCCTTTTCAATTAAATC---AATTGGTCATCAATGA-----	583
ANJ26A	TGAAATTGAAATCCTTTTCAATTAAATC-----	554
ANJ22	TGAAATTGTAGATCCATTCTTCAATTAAATC---AATTGGTCATC-----	579
ANJ25	TGAAATTGTAGATCCATTCTTCAATTAAATC-----	559
ANJ28	TGAAA-----	539
ANJ23	TGAAATTGAAATCCTTTTCAATTAAATC---AATTGGTC-----	496
ANJ19B	TGAAAATGAAGATCAATTCTTCAATTAAATA---AAT-----	569
NGA17	TGAAATTGAAATCCTTTTCAATTAAATC---AATTGGTCATAAT-----	579
NGA23	TGAAATTGAAATCCTTTTCAATTAAATC---AATTGGTCATCAATGAA-----	583
MOH25B	TGAAATTGAAATCCTTTTCAATTAAATC---AATTGGTCATC-----	577
NGA22	TGAAATTGAAATCCTTTTCAATTAAATC---AATTGGTCATC-----	577
NGA26A_(reversed)	TGAAATTGAAATCCTTTTCAATTAAATC---AATT-----	570
ANJ33	TGAAATTGAAATCCTTTTCAATTAAATC---AAT-----	569
MOH24	-----	316
ANJ35A_2	TGAAATGAAATTGCAAATCCATTCTTCTATC---TATTGGTCATAATGATATTGA	374
ANJ35B	TGAAATGAAATTGCAAATCCATTCTTCTATC---TATTGGTCAC-----	363
ANJ34	TGAAATGAAATTGCAAATCCATTCTTCTATC---TATTGGTCATCAATGAT-----	370
ANJ31	TGAAATGAAATTGCAAATCCATTCTTCTATC---TATTGGT-----	360
ANJ30	TGAAATGAAATTGCAAATCCATTCTTCTATC---TATTGGC-----	361
Anj21	TGAAATGAAATTGCAAATCCATTCTTCTATC---TATTGGTCAT-----	363
MOH28A	TGAAATTGAAATCCTTTTCAATTAAATC---AATTGG-----	609
NGA11B	TGAAATTGAAATCCTTTTCAATTAAATC---AATTGGTCATAATGAATT-----	622
ANJ18A	TGAAATTGAAATCCTTTTCAATTAAATC---AATTGGTC-----	611
MOH10	TGAAATTGAAATCCTTTTCAATTAAATC---AATTGGTCACA-----	614
NGA26B_(reversed)	TGAAATTGAAATCCTTTTCAATTAAATC---AATTGGTC-----	612
NGA16	TGAGATTGAAATCCTTTTCAATTAAATC---GGTTGGTCATCCGTGAA-----	621
MOH12C	TGAAATTGAAATCCTTTTCAATTAAATC---AATTGGTCATCAATG-----	550

APPENDIX IV: GeneAlex file for predicted queen genotypes data. The data was used for genetic diversity, population differentiation and population structure analysis. Individuals with \* were manually predicted from the genotyped workers whilst the rest were obtained as a result of sibship analysis in Colony version 2.0.6.4. 1 represent missing data

MOH15	MO H	210	21 0	285	28 5	152	15 4	132	13 6	125	12 5	16 9	17 1	181	18 1	240	24 0	148	15 2	104	10 8	221	22 3	169	17 3
MOH16	MO H	210	21 0	285	28 5	152	15 2	132	13 6	133	13 3	16 9	16 9	181	18 1	231	24 0	152	15 2	104	10 8	223	22 3	169	17 1
MOH17	MO H	210	21 0	285	28 5	150	15 2	126	13 4	133	13 3	16 9	16 9	181	18 1	240	24 0	152	15 2	108	10 8	233	23 3	173	17 3
MOH18	MO H	210	21 0	285	28 5	140	15 3	134	13 4	135	13 5	16 9	16 9	183	18 3	246	24 6	144	14 4	104	10 4	233	23 3	171	17 1
MOH19	MO H	210	21 0	285	28 5	150	15 0	126	13 6	133	13 3	17 1	17 1	171	17 1	240	24 0	144	14 4	104	10 4	223	23 7	169	17 5
MOH23	MO H	212	21 2	283	28 3	140	14 0	128	12 8	133	13 9	16 9	16 9	171	17 1	240	24 0	144	14 4	104	10 4	233	23 3	169	17 3
MOH24	MO H	212	21 2	285	28 5	140	15 4	126	13 6	133	13 3	17 1	17 1	181	18 1	240	24 0	148	14 8	104	10 4	223	22 3	157	15 7
MOH25	MO H	212	21 2	285	28 5	150	15 0	138	13 8	139	13 9	16 9	16 9	171	17 1	240	24 0	144	14 8	108	10 8	225	23 3	169	17 1
MOH26	MO H	212	21 2	285	28 5	140	14 0	132	13 4	139	13 9	17 1	17 1	171	17 1	246	24 6	152	15 2	104	10 8	233	23 3	169	16 9
MOH29	MO H	212	21 2	285	28 5	140	15 0	138	13 8	133	13 3	16 5	17 1	181	18 1	240	24 0	144	14 4	104	10 4	237	23 7	173	17 3
MOH30	MO H	212	21 2	285	28 5	140	14 0	134	13 6	139	13 9	16 9	16 9	177	17 7	240	24 6	144	15 2	104	10 4	237	23 7	173	17 3
MOH31	MO H	212	21 2	285	28 5	152	15 4	134	13 6	133	13 3	16 9	16 9	171	17 1	240	24 0	152	15 2	104	10 4	245	24 5	171	17 3
MOH34	MO H	212	21 2	283	28 5	152	15 2	126	12 6	139	13 9	17 3	17 3	177	17 1	240	24 0	144	15 2	108	10 8	233	23 3	157	15 7
MOH38	MO H	214	21 4	283	28 5	140	14 0	138	13 8	133	13 3	16 9	16 9	173	17 7	240	24 6	144	14 4	104	10 4	235	23 5	169	16 9
MOH4	MO H	210	21 2	285	28 5	140	14 0	134	13 6	133	13 3	16 6	16 6	181	18 1	240	24 0	144	14 4	104	10 8	237	23 7	169	17 3
MOH41	MO H	212	21 2	285	28 5	154	15 4	140	14 0	133	13 3	16 9	16 9	181	18 1	242	24 2	148	14 8	104	10 4	235	23 5	169	17 1
MOH42	MO H	212	21 2	291	29 1	152	15 2	128	12 2	135	13 5	16 9	16 9	173	17 3	240	24 0	148	14 8	104	10 8	225	22 5	169	17 3
MOH43	MO H	210	21 2	285	28 5	150	15 0	134	13 4	123	12 3	16 9	16 9	171	17 1	240	24 6	148	14 8	104	10 4	225	22 5	173	17 5
MOH5	MO H	210	21 0	283	28 3	140	14 0	136	13 6	123	13 9	16 9	17 1	173	17 3	240	24 0	152	15 2	108	10 8	233	23 3	169	16 9
MOH7	MO H	210	21 0	285	28 5	150	15 0	126	12 6	135	13 5	16 9	16 9	171	17 1	240	24 2	148	15 2	104	10 8	223	22 3	157	16 9
MOH8	MO H	210	21 0	285	28 5	152	15 2	132	13 4	139	13 9	16 9	16 9	171	17 1	240	24 0	142	14 4	104	10 8	233	23 3	169	16 9
MOHQ1	MO *	212	21 2	285	28 5	140	14 0	126	12 6	139	13 3	16 9	17 1	173	17 7	240	24 0	144	15 2	104	10 8	223	22 3	169	17 3
MHQ2*	MO H	210	21 2	283	28 5	140	15 2	126	12 6	133	13 9	16 9	16 9	171	17 7	240	24 0	144	15 2	104	10 8	235	23 7	169	16 9
MOHQ3	MO *	212	21 2	283	28 5	140	14 0	136	13 8	139	13 9	16 9	17 1	171	17 7	240	24 6	144	14 4	104	10 4	223	23 7	169	17 3
MOHQ4	MO *	210	21 0	285	28 5	140	14 0	136	13 8	139	13 3	16 9	16 9	171	17 7	240	24 0	144	15 2	104	10 8	235	23 7	157	16 9
NGA10	NGA	212	21 4	283	28 5	150	15 0	128	12 8	1	1	1	1	168	16 8	222	22 5	142	14 2	104	10 4	223	23 0	169	16 9
NGA11	NGA	212	21 4	285	28 5	150	15 2	128	12 8	1	1	1	1	169	16 9	222	22 2	142	14 2	104	10 4	225	22 5	169	16 9
NGA12	NGA	212	21 2	285	28 5	150	15 0	128	12 8	1	1	1	1	170	17 0	237	23 7	152	15 2	104	10 4	233	23 7	169	16 9
NGA13	NGA	212	21 2	283	28 3	150	15 0	128	12 8	1	1	1	1	173	17 3	222	22 2	126	12 6	104	10 4	223	23 3	169	17 1
NGA14	NGA	212	21 2	285	28 1	150	15 0	132	13 6	1	1	1	1	170	17 0	222	22 2	142	14 2	104	10 4	223	22 3	169	17 1
NGA16	NGA	212	21 2	283	28 3	150	15 0	130	13 4	1	1	1	1	168	16 8	237	23 7	152	15 2	104	10 4	225	22 5	157	17 7
NGA17	NGA	212	21 2	291	29 1	150	15 0	134	13 6	1	1	1	1	170	17 0	235	23 5	142	14 2	104	10 4	235	23 5	169	17 1

NGA18	NGA	212	21	2	285	28	5	150	15	0	134	13	4	1	1	1	1	170	17	0	235	23	5	148	15	2	104	10	4	235	23	5	171	17	1
NGA2	NGA	212	21	2	285	28	5	150	15	0	130	13	0	1	1	1	1	170	17	4	222	23	5	126	12	6	104	10	4	223	23	5	169	17	1
NGA20	NGA	212	21	2	283	29	3	150	15	0	134	13	6	1	1	1	1	172	17	2	235	23	7	152	15	2	104	10	4	237	23	7	173	17	3
NGA21	NGA	212	21	4	283	29	1	150	15	2	136	13	6	1	1	1	1	168	16	8	235	23	5	142	14	2	104	10	4	235	23	5	169	16	9
NGA23	NGA	212	21	4	283	28	3	150	15	2	136	13	6	1	1	1	1	170	17	0	222	22	2	142	14	2	104	10	4	223	22	3	171	17	5
NGA24	NGA	212	21	2	283	28	3	150	15	0	134	14	2	1	1	1	1	170	17	0	233	23	7	152	15	2	104	10	4	233	23	3	171	17	1
NGA26	NGA	218	21	8	285	29	1	140	14	0	128	12	8	1	1	1	1	170	17	0	237	23	7	126	15	2	104	10	4	223	22	3	171	17	1
NGA27	NGA	214	21	4	285	28	5	150	15	0	128	13	4	1	1	1	1	170	17	0	222	22	2	142	14	2	104	10	4	225	22	5	169	17	1
NGA3	NGA	212	21	2	283	28	3	150	15	0	128	13	6	1	1	1	1	170	18	2	235	23	5	126	12	6	104	10	4	235	23	5	169	17	1
NGA32	NGA	214	21	4	283	28	5	150	15	0	136	13	6	1	1	1	1	173	17	3	233	23	5	142	14	2	104	10	4	223	23	3	175	17	5
NGA33	NGA	214	21	4	285	29	1	150	15	0	134	13	6	1	1	1	1	168	16	8	222	22	2	152	15	2	104	10	4	223	22	3	169	16	9
NGA5	NGA	210	21	0	291	29	1	140	15	0	130	13	4	1	1	1	1	173	17	3	237	23	7	142	14	2	104	10	4	223	22	3	175	17	5
NGA7	NGA	210	21	0	283	28	3	140	14	0	136	13	6	1	1	1	1	168	16	8	235	23	5	142	15	2	104	10	4	235	23	5	169	17	1
NGAQ1*	NGA	210	21	2	285	29	1	140	15	2	132	13	6	1	1	1	1	168	17	0	222	22	4	142	14	2	104	10	4	223	22	5	157	16	9
NGAQ2*	NGA	210	21	2	291	28	7	140	14	0	134	14	2	1	1	1	1	168	16	8	222	23	3	142	14	2	104	10	4	233	23	7	169	16	9
NGAQ3*	NGA	210	21	0	283	29	1	140	15	2	136	13	8	1	1	1	1	170	17	4	222	23	5	142	15	2	104	10	4	223	23	5	1	1	1
NGAQ4*	NGA	210	21	4	285	29	1	140	14	0	136	13	6	1	1	1	1	170	17	0	235	23	5	142	14	2	104	10	4	223	23	5	169	17	1

APPENDIX V: Table showing the deviations from HWE equilibrium. The table was used to generate the heatmap

	<b>Anjouan</b>			<b>Moheli</b>			<b>Ngazidja</b>		
	$\chi^2$	df	Pr( $\chi^2$ )	$\chi^2$	df	Pr( $\chi^2$ )	$\chi^2$	df	Pr( $\chi^2$ )
A113	40.386	6	3.823750e-07	44.9382716	3	9.536458e-10	35.99902	6	2.757833e-06
A56	3.070	1	7.974608e-02	32.7769376	3	3.589166e-07	12.88838	6	4.484319e-02
A88	78.448845	15	1.340488e-10	46.0302217	15	5.258484e-05	13.39649	3	3.853113e-03
AP81	54.622631	15	2.066271e-06	67.6741650	21	8.251074e-07	33.97822	21	3.643426e-02
A28	16.102978	3	1.080168e-03	102.9570370	15	3.552714e-15	0.00000	0	1.000000e+00
A8	0.000000	0	1.000000e+00	66.1378512	10	2.454145e-10	0.00000	0	1.000000e+00
AC306	10.534224	10	3.949371e-01	56.8925000	10	1.396351e-08	91.22911	21	9.896095e-11
AP33	34.213994	10	1.698971e-04	16.5711662	6	1.099546e-02	23.36000	10	9.492987e-03
A43	32.293333	3	4.539011e-07	15.8556984	6	1.454981e-02	30.77310	6	2.800700e-05
AP23	2.135895	1	1.438865e-01	0.4288307	1	5.125633e-01	0.00000	0	1.000000e+00
B124	51.709126	15	6.309339e-06	87.0313933	21	5.221859e-10	43.36761	15	1.380352e-04
UN4987	34.157791	10	1.553367e-02	21.9168842	10	1.553367e-02	46.82195	10	1.017350e-06

APPENDIX VI: AMOVA results showing significant variations between populations, between samples within populations and within samples with populations

	Df	Sigma	Covariance (%)	Phi	p value
Between populations	2	2.27	24.97	0.64	0.001
Between samples within population	70	3.56	39.12	0.52	0.001
Within samples within populations	73	3.27	35.91	0.25	0.001
Total variations	145	9.11	100.00		